

Pathogenesis, prevalence, diagnosis, and management of exercise-induced bronchoconstriction: a practice parameter

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1. Practice parameters for the diagnosis and treatment of asthma. *J Allergy Clin Immunol.* 1995;96(suppl):S707–S870.
2. Practice parameters for allergy diagnostic testing. *Ann Allergy.* 1995;75:543–625.
3. Practice parameters for the diagnosis and management of immunodeficiency. *Ann Allergy.* 1996;76:282–294.
4. Practice parameters for allergen immunotherapy. *J Allergy Clin Immunol.* 1996;98:1001–1011.
5. Disease management of atopic dermatitis: a practice parameter. *Ann Allergy.* 1997;79:197–211.
6. The diagnosis and management of anaphylaxis. *J Allergy Clin Immunol.* 1998;101(suppl):S465–S528.
7. Algorithm for the diagnosis and management of asthma: a practice parameter update. *Ann Allergy.* 1998;81:415–420.
8. Diagnosis and management of rhinitis: parameter documents of the Joint Task Force on Practice Parameters in Allergy, Asthma and Immunology. *Ann Allergy.* 1998;81(suppl):S463–S518.
9. Parameters for the diagnosis and management of sinusitis. *J Allergy Clin Immunol.* 1998;102(suppl):S107–S144.
10. Stinging insect hypersensitivity: a practice parameter. *J Allergy Clin Immunol.* 1999;103:963–980.
11. Disease management of drug hypersensitivity: a practice parameter. *Ann Allergy.* 1999;83(suppl):S665–S700.
12. Diagnosis and management of urticaria: a practice parameter. *Ann Allergy.* 2000;85(suppl):S521–S544.
13. Allergen immunotherapy: a practice parameter. *Ann Allergy.* 2003;90(suppl):SI–S540.
14. Symptom severity assessment of allergic rhinitis: part I. *Ann Allergy.* 2003;91:105–114.
15. Disease management of atopic dermatitis: an updated practice parameter. *Ann Allergy.* 2004;93:S1–S21.
16. Stinging insect hypersensitivity: a practice parameter update. *J Allergy Clin Immunol.* 2004;114:869–886.
17. The diagnosis and management of anaphylaxis: an updated practice parameter. *J Allergy Clin Immunol.* 2005;115:S483–S523.
18. Practice parameter for the diagnosis and management of primary immunodeficiency. *Ann Allergy.* 2005;94:S1–S63.
19. Attaining optimal asthma control: a practice parameter. *J Allergy Clin Immunol.* 2005;116:S3–S11.
20. The diagnosis and management of sinusitis: a practice parameter update. *J Allergy Clin Immunol.* 2005;116:S13–S47.
21. Food allergy: a practice parameter. *Ann Allergy.* 2006;96:S1–S8.
22. Contact dermatitis: a practice parameter. *Ann Allergy.* 2006;97:S1–S38.
23. Allergen immunotherapy: a practice parameter second update. *J Allergy Clin Immunol.* 2007;120:S25–S85.
24. Allergy diagnostic testing: an updated practice parameter. *Ann Allergy.* 2008;100:S1–S147.
25. Diagnosis and management of rhinitis: an updated practice parameter. *J Am Coll Immunol.* 2008;121:S1–S84.
26. Kelso J, Li JT. Adverse reactions to vaccines. *Ann Allergy.* 2009;103:S1–S16.
27. Lieberman P, et al. The diagnosis and management of anaphylaxis practice parameter: 2010 Update. *J Allergy Clin Immunol* 2010; 126(3):477–480.e.42.
28. Solensky R, et al. Drug Allergy: An Updated Parameter. *Ann Allergy* 2010;105:273e.1–273e.78.

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CLASSIFICATION OF RECOMMENDATIONS AND EVIDENCE

Category of evidence

- Ia Evidence from meta-analysis of randomized controlled trials
- Ib Evidence from at least 1 randomized controlled trial
- IIa Evidence from at least 1 controlled study without randomization
- IIb Evidence from at least 1 other type of quasi-experimental study
- III Evidence from nonexperimental descriptive studies, such as comparative studies
- IV Evidence from expert committee reports or opinions or clinical experience of respected authorities or both

Strength of recommendation

- A Directly based on category I evidence
- B Directly based on category II evidence or extrapolated recommendation from category I evidence

-
-
- C Directly based on category III evidence or extrapolated recommendation from category I or II evidence
 - D Directly based on category IV evidence or extrapolated recommendation from category I, II, or III evidence
 - LB Laboratory based

GLOSSARY

Exercise-induced bronchoconstriction (EIB) is defined as a transient narrowing of the lower airway following exercise in the presence or absence of clinically recognized asthma. The term *exercise-induced asthma* (EIA) is not used in this document because it may imply incorrectly that exercise causes rather than exacerbates or triggers an attack of asthma.

Bronchial hyperresponsiveness (BHR) is an increase in sensitivity to an agent and is expressed as the dose or concentration of a substance that produces a specific decrease in forced expiratory volume in 1 second (FEV₁) (eg, provocation dose or provocative concentration causing a decrease of 20% [PD₂₀ or PC₂₀, respectively]).

Bronchial reactivity is the rate of change of the FEV₁ in relation to the dose or stimulus (eg, response dose ratio [RDR] with mannitol is the percentage decrease divided by the dose that achieves that decrease or the percentage decrease in exercise in response to the optimal stimulus).

Conditioning is defined as preparing the body for physical exercise and, in particular, sports performance.

Competitive athletes are individuals who engage in strenuous aerobic activity at any level from grade school age and older.

Elite athletes are highly competitive individuals who train and compete consistently at higher levels (eg, Olympics, professional aerobic sports).

Tolerance is a decrease in the degree and/or duration of response to an agent when used continuously instead of intermittently. Tolerance ordinarily refers to the inhibition of bronchoconstriction and in some cases bronchodilation to β_2 -adrenergic agents.

PREFACE

The goal of “Pathogenesis, Prevalence, Diagnosis, and Management of Exercise-Induced Bronchoconstriction: A Practice Parameter” is to empower health care specialty practitioners to provide outstanding health care services to their patients in diagnosing and managing EIB. This practice parameter is designed to accomplish this goal by providing the most up-to-date, evidence-based information and recommendations on the diagnosis and management of EIB. The term *EIA* is not used in this document because it implies a condition that does not exist—that of exercise that induces asthma. Instead, the term *EIB* is used throughout, and EIB is further differentiated based on whether the patient has chronic asthma in which exercise triggers bronchoconstriction or the patient does not have chronic asthma and only has bronchoconstriction associated with exercise.

This document was developed by a Workgroup under the aegis of the Joint Task Force on Practice Parameters. Three

national allergy and immunology organizations—the American Academy of Allergy, Asthma and Immunology (AAAAI), the American College of Allergy, Asthma and Immunology (ACAAI), and the Joint Council of Allergy, Asthma and Immunology (JCAAI)—have authorized the Joint Task Force to develop new parameters and update existing parameters. To date, the Joint Task Force has developed and published 28 practice parameters, including some that have been updated, for the specialty of allergy/immunology (see www.jcaai.org).

This parameter is the first such parameter for EIB. It provides the latest in evidence-based recommendations on the pathogenesis, prevalence, clinical presentation, diagnosis, differential diagnosis, and treatment (both nonpharmaceutical and pharmaceutical) of patients having or suspected of having EIB, with or without chronic asthma, in recreational and elite athletes and nonathletes. The document was written, edited, and reviewed by specialists in the discipline of allergy/immunology and was exclusively funded by the allergy and immunology organizations noted above.

The Workgroup, chaired by John M. Weiler, MD, MBA, developed the initial draft, which was then edited and reviewed by the Joint Task Force. In the development of this document, a comprehensive search of the medical literature was performed using the PubMed MEDLINE search engine and included Cochrane databases. Searches used keywords that were associated with EIB. Workgroup members, all of whom were experts in the area of EIB, contributed additional manuscripts to the list. Published clinical investigations were rated by category of evidence and the strength of clinical recommendations. After the Joint Task Force completed its review and editing of the manuscript, subject matter experts in the field of allergy and immunology and EIB were recruited to review the manuscript. These expert reviewers were appointed by the AAAAI and ACAAI. The authors of the document then diligently reviewed and evaluated additional comments and suggestions from these reviewer experts. The revised final document presented here was approved by the sponsoring organizations and reflects an evidence-based and generally accepted consensus parameter on the diagnosis and management of EIB encompassing the most contemporary evidence-based understanding of the pathogenesis, prevalence, clinical presentation, diagnosis, differential diagnosis, and treatment (both nonpharmaceutical and pharmaceutical) of patients having or suspected of having EIB, with or without chronic asthma, in recreational and elite athletes and nonathletes.

The term *EIB* is defined as a transient narrowing of the airway with increasing airway resistance after exercise. Exercise-induced bronchoconstriction most likely occurs because of dehydration of the airway resulting in a hyperosmolar environment, leading to mast cell mediator release and subsequent bronchoconstriction. The hyperosmolar theory of the pathogenesis of EIB is described in this document as the leading theory for EIB. The document also provides evidence that as many as 90% of asthmatic patients and 50% of competitive athletes may experience EIB. The diagnosis of

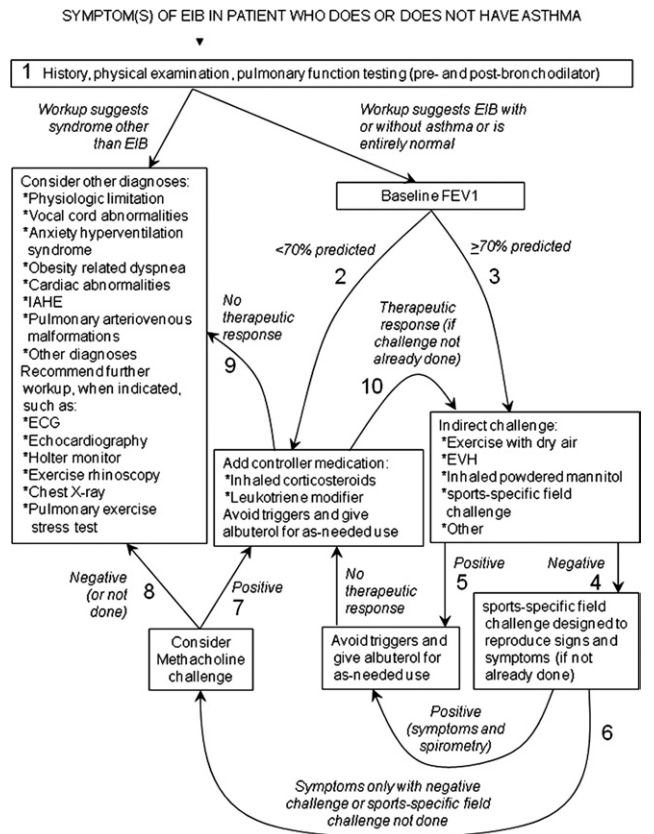


Figure 1. Algorithm: The major decision tree for the evaluation and management of individuals with suspected EIB in the presence or absence of chronic asthma.

EIB, which relies optimally on objective pulmonary function testing and exercise challenge, is discussed, as is the clinical presentation and specifics of diagnosis (eg, the value and conduct of exercise challenge or surrogate challenges). The differential diagnosis is elucidated, including other upper and lower airway disorders, cardiopulmonary conditions, and other diseases and conditions that mimic EIB. Finally, therapy is discussed, including the use of bronchodilators, anti-inflammatory agents, and the combination of pharmacotherapy and nonpharmacotherapy (eg, warm-up and cool-down maneuvers and dietary intervention). The diagnosis and particularly the management of EIB in elite athletes are discussed in detail in a separate section to explain how this condition may affect these highly conditioned individuals. Tables and figures are provided to illustrate key points.

The Executive Summary details the key items in summary statements in the Practice Parameter. The Executive Summary does not discuss all of the parameter topics in depth. An annotated algorithm in the document provides the major decision tree for the evaluation and management of individuals with suspected EIB in the presence or absence of chronic asthma (Figure 1). This is followed by a list of summary statements that provide the key points in

the evaluation and management of EIB. In the evidence-based commentary, the summary statements are repeated and are then followed by text that elaborates and supports each summary statement. The evidence-based commentary first discusses the general issues relating to EIB including pathogenesis, prevalence, clinical presentation, diagnosis, differential diagnosis, and treatment (both nonpharmaceutical and pharmaceutical) of patients having or suspected of having EIB, with or without chronic asthma, in recreational and elite athletes and nonathletes.

The Joint Task Force on Practice Parameters expresses gratitude to the AAAAI, ACAAI, and JACAAI, who provided support for the development of the parameter, and to the individuals who have dedicated time and effort to the drafting and review of this document.

ALGORITHM

Annotations for Algorithm

1 History, physical examination, and pulmonary function tests pre and postbronchodilator (with flow volume loops, and pre/postbronchodilator functions) are necessary if there is suspicion of asthma or EIB. History alone should not be used to diagnose or exclude the diagnosis of EIB. If pulmonary function equipment (i.e., spirometry) is not available in the clinic setting, patients should be referred to a pulmonary function laboratory or to a physician with office spirometry equipment. Peak flow is a poor surrogate for diagnosis.

2 and 3

Expert consensus is, if baseline FEV₁ is 70% predicted value, exercise challenge with dry air and EVH should not be done for safety reasons. Other challenges (e.g., exercise without the use of dry air, mannitol) should be performed with caution if baseline FEV₁ is 70% predicted value.³ Eucapnic voluntary hyperpnea may be a surrogate for exercise, especially for those who cannot exercise or for elite athletes; however, EVH can elicit a severe response. Inhalation of dry air as part of the exercise protocol is strongly recommended to diagnose or exclude EIB because it increases sensitivity to the challenge. Mannitol challenge is currently approved in 23 countries including the US. More than one indirect challenge may be required for diagnosis if a first challenge is negative. When predicted spirometry values are obtained outside of the range of the algorithm, judgment must be used when interpreting the results. Care must be taken when evaluating patients who are unusually tall or short, in particular, and have a predicted FEV₁ that is 70%.

4

If indirect challenge is negative, then a field challenge to reproduce signs and symptoms of the exercise-induced bronchospasm is recommended.³

5

If indirect challenge is positive, therapy with albuterol as needed with avoidance of triggers for EIB may be indicated.

6

If field challenge is negative but the patient is still symptomatic, a methacholine challenge may be performed to rule out BHR.

7

If methacholine challenge is positive, controller medications, including inhaled steroids, should be considered for probable asthma in conjunction with avoidance of triggers and use of albuterol as needed.

8

If methacholine challenge is negative, a different diagnosis should be considered. An extensive differential diagnosis should be considered if the evaluation (including history, physical examination, pulmonary function testing pre and postbronchodilator, exercise challenge, and methacholine) is not consistent with EIB. The differential diagnosis encompasses physiologic limitations including VCD, exercise dyspnea, anxiety, obesity or a gradient of poor conditioning, or cardiac abnormalities (e.g., tachycardia, idiopathic hypertrophic subaortic stenosis, hyperventilation syndrome, pulmonary arteriovenous malformation). The further evaluation may include ECG, echocardiography, Holter monitor, and exercise rhinoscopy as indicated.^{2,3}

9

If the controller medication is ineffective, then the steps described in Annotation 8 should be followed.

10

If the controller medication is effective, challenge should be performed to document EIB if the FEV₁ after controller therapy is greater than 70%. If the FEV₁ is not greater than 70%, additional consideration should be given to different diagnoses.

EXECUTIVE SUMMARY

Introduction and Definition (Summary Statement 1)

Exercise-induced bronchoconstriction is defined as the transient narrowing of the lower airways that occurs after vigorous exercise.¹⁻⁴ Exercise-induced bronchoconstriction may be observed in patients who have or do not have chronic asthma based on spirometry.¹⁻⁸ The term *EIA* should no longer be used because exercise does not induce asthma but rather is a trigger of bronchoconstriction. The diagnosis of EIB usually requires a decrease in FEV₁ after exercise of 10% to 15% of the preexercise value.^{3,6-8} Exercise-induced bronchoconstriction is a manifestation of BHR and is often the first sign of asthma.^{3,6-8} A β_2 -agonist by inhalation is the most widely used treatment given immediately before exercise to prevent EIB.⁷ Within 12 weeks of initiating daily treatment with inhaled corticosteroids (ICS) alone, the inflammation and resulting decrease in pulmonary function are attenuated in some but not all patients who have EIB with asthma.⁷⁻⁹

Pathophysiology of EIB (Summary Statements 2-7)

The pathophysiology of EIB has been elucidated during the past decade and a half. Exercise, especially strenuous exercise,

causes hyperpnea, which in turn causes drying of the airways, requiring humidification and often warming of large volumes of air during a short interval. The respiratory water loss that occurs at high ventilation rates, which may be associated with airway cooling and dehydration, leads to increases in osmolarity of the airway surface. The increase in osmolarity of the airway is postulated to induce degranulation of airway mast cells with release of chemical mediators, including prostaglandins (PGs), leukotrienes (LTs), and histamine, with bronchoconstriction of the airways. Inflammation of the airway may occur with the involvement of lymphocytes, eosinophils, epithelial cells, and perhaps neutrophils.¹⁰⁻¹³ This is the predominant explanation for EIB called the *osmotic theory*.¹⁰⁻¹³ The most important determinants of the EIB response and its severity are the water content of the inspired air and the level of work that is achieved and sustained during exercise.¹² It is important to understand that exercise itself is not necessary to cause the airways to narrow as demonstrated by the ability of eucapnic voluntary hyperpnea (EVH) of dry air to induce bronchoconstriction, similar to what is observed after exercise.¹⁰⁻¹³

It was previously postulated that warming the airway after inhalation of cold, dry air was the main mechanism causing EIB, called the *thermal theory*¹⁰⁻¹³; however, it is now known that cooling and rewarming of the airways are not required to provoke airway narrowing, in contrast to drying and osmotic changes with release of mediators of inflammation, which are required. It is now thought that airway rewarming may play at most only a limited role in the pathogenesis of EIB.¹⁰⁻¹³

In approximately half of patients who have EIB, there is an interval of refractoriness lasting approximately 1 to 3 hours immediately after an episode of EIB during which additional exercise results in little or no bronchoconstriction. This is called the *refractory period* and may also occur after strenuous exercise at a level that does not provoke EIB. This offers a nonpharmacologic approach to prevent attacks of EIB in athletes before competition.¹⁰⁻¹³

Genetics and Environment (Summary Statements 8-10)

It has been postulated that both genetics and the environment may contribute, together or alone, to the EIB phenotype.¹⁴⁻¹⁶ Unfortunately, there is limited information at present on the genetic makeup of individuals who have EIB, whether in the presence or absence of chronic asthma. Future genome-wide association studies should clarify the role of genetics in causing the EIB phenotype.¹⁴⁻¹⁶ In contrast, data exist to demonstrate that environmental conditions play an important role in some individuals in facilitating the occurrence of EIB. The environment in which training or participation occurs may contain allergens and/or pollutants that, when inhaled, are associated with oxidative stress, which may in turn enhance the development and severity of EIB.^{7,10-12,14-16} This is particularly true in elite athletes in whom the pathogenesis of EIB may relate to effects on the airway developing from conditioning large volumes of dry air over months or years of training with or without exposure to environmental irritants, allergens, and infectious agents.^{7,10-12,14-16}

Prevalence (Summary Statements 11–16)

Exercise-induced bronchoconstriction is reported to occur in as many as 90% of asthmatic patients,^{3,6–13} perhaps as a reflection of underlying asthma. Patients with more severe or poorly controlled asthma are more likely to manifest EIB than patients with less severe or well-controlled disease.^{3,6–13} The true prevalence of EIB in the general population is poorly defined; most epidemiologic studies of EIB have not differentiated asthmatic from non-asthmatic populations.^{1–3,7} In addition, prevalence data are affected by the lack of consensus on how a challenge is to be performed and what constitutes a positive response.^{1–3,6,7,17,18} Other factors determining prevalence of EIB in the general population include the environmental conditions, type of exercise, and intensity with which the exercise or surrogate challenge is performed.^{1–3,6,7,17,18} The prevalence of EIB may also be influenced by age, sex, and ethnicity. It has long been observed that the prevalence of EIB in elite athletes appears to be higher than in the general population and depends on the environmental conditions in which exercise is performed, the type of sport, and the maximum exercise level.^{1–3,6,7,17,18}

In reviewing data on the prevalence of EIB, it is important to recognize that the response to exercise in an unselected population forms a normally distributed curve, with some individuals even having increases in FEV₁ after the challenge; there is no specific cutoff that clearly distinguishes a positive from a negative test result.^{1–3,6,7,17,18} It is also important to recognize that symptoms are poor predictors of individuals who will have a positive exercise challenge because so many other conditions may occur that may be confused with EIB.^{1,2,7,19}

Diagnosis (Summary Statements 17–23)

Symptoms of EIB include cough, wheeze, chest pain primarily in children or chest tightness, shortness of breath, dyspnea, excessive mucous production, or feeling out of shape when the patient is actually in good physical condition.^{1,2,7,17,20} These symptoms also occur with other conditions so that diagnosis of EIB based only on symptoms lacks sensitivity and specificity to predict a positive exercise challenge. The diagnosis of EIB should never be made based on symptoms alone when unaccompanied by data from an objective exercise or surrogate challenge.^{1–3,6,7,17,18}

Diagnostic challenges are of 2 types: (1) direct challenges in which a pharmaceutical agent such as methacholine or histamine is the provoking agent that acts directly on airway smooth muscle and (2) indirect challenges in which exercise or a surrogate, such as EVH, inhalation of mannitol, inhalation of adenosine monophosphate (AMP), or inhalation of hypertonic saline, is the provoking agent that triggers mediator release. Mediator release in turn leads to bronchoconstriction. Indirect challenges are more specific in reflecting BHR because of airway inflammation and are preferred as a way to confirm underlying asthma.^{1–3,6,7,17,18} In addition, indirect challenges are recommended for monitoring asthma

therapy because BHR is most often associated with inflammation,^{1–3,6,17,18} which is diminished by ICS therapy.^{7–9,18}

Direct challenges using methacholine, an approved agent, may be performed in an office setting by trained personnel. The challenge, described in a consensus statement by the American Thoracic Society (ATS),⁶ requires administering increasing concentrations of methacholine by inhalation and following FEV₁ levels after each dose. Drug is administered using a nebulizer and either a dosimeter with a forced inhalation from end expiratory volume or alternatively by using tidal breathing over a timed interval. A decrease in FEV₁ of more than 20% from baseline is regarded as a positive challenge result. This is documented as the PC₂₀. A PC₂₀ of greater than 16 mg/mL is interpreted as normal or lack of BHR, between 4.0 and 16 mg/mL is interpreted as borderline BHR, between 1.0 and less than 4 mg/mL is considered positive with mild BHR, and less than 1.0 mg/mL is considered positive with moderate to severe BHR. Although the direct challenge is used as a screening test for chronic asthma, especially to rule out asthma, it is not useful to detect EIB because it has low sensitivity for EIB because it reflects the effect of only a single agonist.^{3,6,7,18}

Indirect challenges should also be conducted only by trained personnel and using standardized protocols. For example, laboratory-based exercise should be performed as described in the consensus statement published by the ATS.⁶ Such a laboratory challenge controls minute ventilation and water content of inhaled air.^{3,6,7,18} Exercise should be for 6 to 8 minutes, at 20°C to 25°C, while breathing dry air at 80% to 90% of estimated maximal heart rate (HR_{max}) as a surrogate for more than 40% of maximum voluntary ventilation (MVV).^{3,6,7,18} Maximal heart rate may be estimated using the formula 220 – age (in years); however, a more accurate equation, which was published recently, to predict HR_{max} is 208 – 0.7 × age.^{3,6,18} Ideally, the exercise ventilation should be above 60% of predicted maximum (ie, greater than 21 times FEV₁)^{3,6,18}; very well-conditioned individuals may require the exercise intensity to be above a 90% HR_{max}. A recent investigation in children reinforces the need to reach a target HR_{max} of 95% for children 9 to 17 years of age; in that study, the decrease in FEV₁ was 25.1% at 95% HR_{max} but 8.8% when the children reached only 85% HR_{max} (Figure 2).¹⁷

Spirometry should be performed at baseline, before exercise challenge, and at predetermined times after exercise, usually at 5, 10, 15, 30, and occasionally 45 to 60 minutes after exercise. The goal is to determine FEV₁ without having the patient perform full forced vital capacity (FVC) maneuvers to avoid causing the patient to become tired by the spirometry efforts. The International Olympic Committee Medical Commission (IOC-MC) Independent Panel on Asthma recommends that FEV₁ should be recorded beginning as soon as 3 minutes after completion of the challenge to overcome the problem of posttest respiratory fatigue. A pre-exercise value is obtained by performing a full FVC maneuver at baseline.^{3,6,7,18} A 10% or greater decrease in FEV₁ from the preexercise value at any 2 consecutive time points within

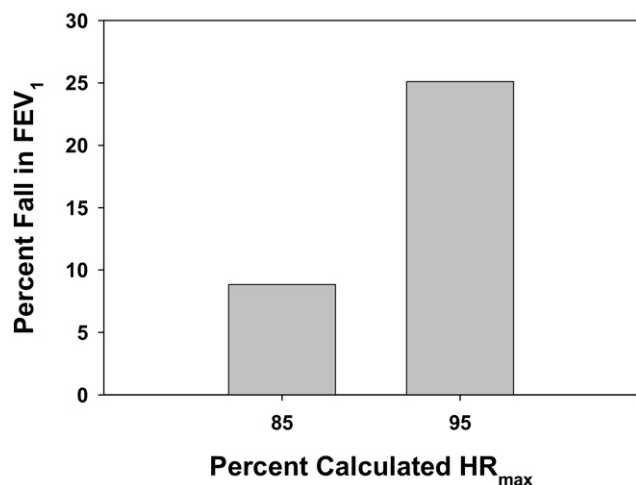


Figure 2. Exercise challenge of 20 asthmatic adolescent patients at 85% of estimated calculated maximum heart rate (HR_{max}) resulted in only 9 of 20 testing positive for exercise-induced bronchoconstriction (mean decrease in forced expiratory volume in one second [FEV₁] of 8.84%); however, all 20 asthmatic patients tested positive after exercise at 95% of HR_{max} (mean decrease in FEV₁ of 25.11%). Adapted from Carlsen et al.¹⁷

30 minutes of ceasing exercise may be considered diagnostic of EIB.^{3,6,7,18} It is preferable to perform FEV₁ testing after exercise without full FVC maneuvers to avoid fatiguing the patient.^{3,6,7,18} If a greater decrease in FEV₁ is required such as a decrease of 25% in FEV₁ as in some pharmaceutical studies, then only 1 time point may be necessary to be diagnostic of EIB. The area under the curve (AUC) of FEV₁ should be examined to determine whether the fall is consistent and not an artifact of an inadequate spirometry effort at 1 or more time points.^{3,6,7,18}

However, there is no single test that will identify all individuals who have EIB.⁷ Decreases in FEV₁ that are consistent with EIB may occur in individuals who are subsequently found to have other conditions.⁷ The FEV₁/FVC ratio may be useful for diagnosis of vocal cord dysfunction (VCD). A flat inspiratory loop on the flow volume curve may also suggest upper airway dysfunction rather than EIB.⁷

Exercise challenge by treadmill is most easily standardized for office practice or a hospital laboratory. Alternative exercise challenges using cycle ergometry may be more difficult to perform and may be less standardized than the treadmill challenge. Similarly, field challenge and free running are even more difficult to standardize.^{3,6,7,18}

Although sport governing bodies require specific cutoff values to diagnose EIB, there is no specific decrease in FEV₁ and there is no single absolute cutoff for a decrease in FEV₁ or change in some other spirometry measure that clearly and unequivocally distinguishes between EIB and lack of EIB.⁷ The ATS has suggested that the postexercise decrease in FEV₁ required to make the diagnosis must be 10%, whereas other groups have suggested a decrease of

13% to 15% is necessary to make the diagnosis.^{3,6,18} A decrease in FEV₁ of 15% after a “field” challenge and a decrease of 6% to 10% in the laboratory have also been recommended.^{3,6,7,18}

Surrogate challenges for exercise in which a hyperosmolar agent, mannitol, or EVH are used are increasingly being recommended by organizations that regulate drug use by elite athletes. Eucapnic voluntary hyperpnea should only be performed by highly trained specialists, and all safety precautions should be observed. Eucapnic voluntary hyperpnea may cause substantial decreases in FEV₁ in a patient with impairment caused by airway inflammation; the EVH test should be performed with caution, especially in patients with an FEV₁ that is below 80% of predicted, and should not be performed on patients in whom the FEV₁ is less than 70% of predicted.^{3,6,7}

Differential Diagnosis (Summary Statements 24–33)

A variety of conditions may mimic EIB. Exercise-induced laryngeal dysfunction (EILD), primarily VCD and other glottic abnormalities, can be clearly differentiated from EIB. An exercise challenge in which a fiberscope is introduced while the exercise is being performed will provide evidence for the anatomical disorder responsible for EILD. Loud inspiratory stridor is the characteristic hallmark of EILD and is rarely seen with EIB. Flattening of the inspiratory loop on a flow volume curve also suggests EILD rather than EIB. Failure to respond to asthma management, especially treatments that prevent or treat EIB, strongly suggests consideration of another diagnosis, such as EILD.^{7,19,20}

Some conditions are more difficult to differentiate from EIB with known asthma, and these conditions may be diagnoses of exclusion. Exercise-induced dyspnea and hyperventilation may simulate asthma and EIB, especially in children and adolescents.^{19,20} Exercise-induced dyspnea is seen as a physiologic limitation in otherwise healthy individuals or in obese individuals without bronchoconstriction. Shortness of breath with exercise may be caused by underlying pulmonary dysfunction unrelated to EIB, for example, restrictive lung physiology caused by obesity, skeletal defects (eg, pectus excavatum), diaphragmatic paralysis, and interstitial pulmonary fibrosis. Shortness of breath accompanied by pruritus and urticaria, particularly after ingestion of certain foods such as celery, may be caused by exercise-induced anaphylaxis (EIA_n). Dyspnea with exertion, with or without chest pain, may be related to cardiovascular, pulmonary, or gastroenterologic causes other than asthma and may require referral to a cardiologist, pulmonologist, or gastroenterologist. Rarely, mitochondrial enzyme deficiency with myopathy may need to be differentiated from EIB.^{7,19,20} Finally, psychological factors should be considered in the differential diagnosis of EIB.^{19,20}

Therapy Overview (Summary Statements 34–37)

Exercise-induced bronchoconstriction in a patient with chronic asthma suggests inadequate control of the underlying asthma or a wrong diagnosis.^{7,21,22} If the diagnosis of EIB can be confirmed, anti-inflammatory therapy should be maximized until control is observed.^{7,21,22}

Many pharmacotherapeutic agents are effective for the prophylaxis or attenuation of EIB even though these agents differ in their mechanisms of action and overall effectiveness. In addition, there is both inpatient and outpatient variability in the ability of the agents to prevent or control EIB when it occurs. These agents may differ in effectiveness over time because of variability of the underlying asthma, environmental conditions, intensity of the exercise stimulus, and the potential of each agent for tachyphylaxis.^{7,21–27}

β_2 -Adrenergic Receptor Agonists (Summary Statements 38–41)

Short-acting β_2 -agonists (SABAs), when administered by inhalation 5 to 20 minutes before exercise, depending on the agent, are effective for 2 to 4 hours in preventing EIB when used intermittently.^{7,21,22} Long-acting β_2 -agonists (LABAs), when administered by inhalation about 30 minutes before exercise, depending on the agent, are effective for as long as 12 hours in preventing EIB when used intermittently.^{7,21,22}

Inhaled SABAs are the single most effective agents to protect against EIB when given prophylactically immediately before exercise and to accelerate recovery of FEV₁ to baseline when administered after a decrease in FEV₁ after exercise. Short-acting β_2 -agonists will attenuate or protect against EIB in most patients.^{7,21,22,24} Their effectiveness may relate in large part to their action as functional agonists acting directly on bronchial smooth muscle (BSM) muscle receptors and by inhibition of mast cell mediator release.^{21,22}

Unfortunately, regular daily use of SABAs and LABAs results in relative loss of efficacy of the agents defined as tachyphylaxis, even when used in combination with ICS.^{7,21–27} Tolerance may be manifested as a reduction in duration and/or magnitude of protection against EIB when LABAs or SABAs are given before exercise and a prolongation of the time necessary for recovery from an attack of EIB when SABAs are given after exercise.^{7,21–27} Therefore, because of these potential concerns of tolerance, SABAs and LABAs are recommended for use only on an intermittent (ie, less than daily) basis as absolutely necessary for prophylaxis and treatment.^{7,21–27}

Long-acting β_2 -agonists should never be used as monotherapy to provide asthma control for chronic asthma but should only be combined with anti-inflammatory therapy, particularly ICS, to provide effective therapy for chronic asthma. Moreover, the addition of ICS has not been clearly demonstrated to diminish tolerance to the bronchoprotective effect of LABAs.^{7,23,26,28}

Daily use of LABAs and SABAs may actually increase the severity of EIB.^{7,23–27} Tolerance may increase with increasing use of SABAs and LABAs, potentially endangering patients

with attacks of severe asthma at the time of greatest need for an effective SABA.^{7,23–27} The onset of tolerance can be as rapid as 12 to 24 hours after beginning therapy with a SABA or LABA, and recovery may occur within 72 hours of discontinuation of SABA or LABA therapy.^{23–27}

Tolerance is often missed because the patient does not require SABAs continuously so that lack of efficacy is not observed by the patient or his or her physician.^{23–27} The mechanism of tolerance is postulated to be that, after long-term exposure to SABAs or LABAs, there is uncoupling and internalization or sequestration of β_2 -receptors in the mast cells and smooth muscle cells where the receptors are degraded. The net loss of functional β_2 -receptors results in downregulation of responsiveness of β_2 -agonists, manifesting as reduction in clinical protection to bronchoconstrictive stimuli. Resynthesis of receptors occurs within 72 hours of cessation of exposure to β_2 -agonists. Mast cell β_2 -receptor downregulation occurs more readily than with smooth muscle so that loss of bronchoprotection precedes loss of bronchodilation.^{23–27} Tolerance to bronchoprotection leads to shortening of duration of effect of β_2 -agonist, whereas tolerance to bronchodilation is demonstrated by prolongation of time to recovery from bronchoconstriction and by response to usual doses of β_2 -agonists.^{23–27}

There are no substantial differences in efficacy among the SABAs currently in use.^{7,21,22} Two LABAs are currently in use and differ primarily in their onsets of action. Formoterol has an onset of bronchodilation and bronchoprotective action of 5 to 15 minutes compared with salmeterol, which requires 15 to 30 minutes for onset of these actions. Both LABAs may protect for as long as 12 hours after the first dose in patients who have not received a SABA or LABA for at least 72 hours (ie, the drug-naïve patient). Unfortunately, even after the first dose of LABA, bronchoprotection for many patients may be for less than 12 hours and the optimal dosing interval for bronchoprotection for EIB may be as short as approximately 6 hours for many patients.^{7,21,23,26}

Leukotriene Inhibitors (Summary Statement 42)

The role of LTs in EIB is to sustain the bronchoconstrictive response. Leukotriene inhibitors are effective when used intermittently or daily to provide prophylaxis for asthma and EIB and do not lead to tolerance.^{7,11,21–24} Inhibitors of the LT pathway, including LT receptor antagonists (LTRAs) and lipoxygenase inhibitors, are effective as prophylactic agents. Leukotriene receptor antagonists, such as montelukast, have an onset of protection within 2 hours of dosing with duration of activity of 12 to 24 hours.^{7,11,21–24} However, the effectiveness of LTRAs is variable among patients, leading to complete inhibition of EIB in some individuals and only partial or no inhibition of EIB in others. Approximately half of patients who receive LTRA are considered responders, with 30% to 80% attenuation of EIB. These percentages may vary, depending on the FEV₁ decrease criteria used to make a diagnosis (eg, >10%, >15%, or >20% decrease in FEV₁) or used to define protection. Patients rarely experience complete

protection. This lack of total protection with the use of LTRAs is expected based on the observation that the pathogenesis of EIB involves additional mediators, including PGs and histamine.^{7,11,21–24}

The LTRAs may decrease the time to recovery from EIB when given prophylactically before exercise. However, they are not as effective as β_2 -agonists, and unlike β_2 -agonists, LTRAs have no ability to reverse airway obstruction when given after obstruction occurs.^{7,11,21–24}

Mast Cell Stabilizers (Summary Statement 43)

Cromolyn sodium and nedocromil sodium, when inhaled shortly before exercise, have bronchoprotective action in approximately half of patients to attenuate the severity of EIB, with rapid onset and a short duration of action rarely longer than 2 hours. Neither cromolyn nor nedocromil is currently available in the United States in an inhaled form. Neither agent has any bronchodilator activity. Mast cell stabilizers may be given with another agent to gain added benefits in obtaining bronchoprotective activity. Tolerance to mast cell stabilizers does not develop when the agents are given repeatedly, and these agents have an excellent safety profile.^{7,21,22,29,30}

Inhaled Corticosteroids (Summary Statements 44–45)

The occurrence of EIB may be a manifestation of lack of control of chronic asthma so that moderate to severe EIB suggests the need for assessment of therapy or another diagnosis. The National Heart, Lung, and Blood Institute Expert Panel Report 3 lists ICS as first-line therapy for chronic asthma alone or in combination with other agents.^{7–9,21,22,28–33} Inhaled corticosteroids have also been shown to have safety and efficacy in decreasing the frequency and severity of EIB in many patients with chronic asthma; ICS are dose and time dependent and may be associated with decreases in inflammatory mediators.^{7–9,21,22,28,31,32} Inhaled corticosteroid attenuates BHR to indirect challenge stimuli, including exercise, EVH, mannitol, and AMP and if given for a sufficiently long time, such as months, to direct stimuli such as methacholine. Inhaled corticosteroids have a rapid effect on exhaled nitric oxide (eNO) levels, another manifestation of the effect of ICS on inflammation.^{7–9}

Some bronchoprotective effect of ICS has been documented as early as 4 hours after the first dose.³² After 1 week of long-term use, ICS efficacy begins to plateau^{7–9,28,31,32}; however, the bronchoprotection may continue to increase slowly over weeks and even months until it reaches its final plateau.^{7–9,21,22,28,31,32} Bronchoprotection with ICS has been demonstrated to occur in 30% to 60% of asthmatic patients with EIB, with marked individual variability ranging from “complete” protection to little or no evidence of protection.^{7–9,21,22,28,31,32}

Inhaled corticosteroids do not obviate the need for short-term bronchoprotection for EIB. β_2 -agonists can be added, if necessary, for short-term prophylaxis of EIB.^{7,21,22} When maintenance ICS are not sufficiently effective to prevent

EIB, LTRAs or β_2 -agonists can be given to enhance bronchoprotection, if necessary.^{7,11,21,22}

Inhaled corticosteroids have not been found to protect against the development of tolerance to β_2 -agonist.^{21–23,26,31} Nevertheless, the combination of ICS and LABAs in a single inhaler is still recommended for treatment of moderate to severe persistent asthma by National Heart, Lung, and Blood Institute guidelines,³³ even when chronic asthma occurs in a patient who also has EIB. Notably, 1 study in adults²⁸ indicated better and more complete bronchoprotection and better asthma control when combination ICS-LABA therapy was given compared with use of ICS alone for 4 weeks. A similar study in children indicated a small persistent effect of bronchoprotection when combination ICS-LABA therapy was used compared with ICS alone.²³

Miscellaneous Therapeutic Agents (Summary Statements 46–47)

Other therapeutic agents have been reported to have efficacy in the prevention of EIB, including approved agents such as anticholinergic agents and theophylline and investigative agents such as antihistamines, inhaled furosemide, ascorbic acid, and inhaled heparin.⁷ Further prospective investigations with large populations are needed to confirm the safety, response rate, and degree of efficacy for these investigational agents.^{7,34}

Nonpharmacologic Therapy (Summary Statements 48–49)

Nonpharmacologic interventions may be effective in helping to control symptoms of EIB but are generally considered as adjunctive to pharmacotherapy. Preexercise warm-up may be helpful in reducing the severity of EIB for 1 to 3 hours because of the refractory period.⁷ Further reduction in EIB may be achieved with postexercise cool-down and breathing through the nose and covering the mouth, particularly in cold, dry weather.⁷ Reduction of sodium intake and ingestion of fish oil and ascorbic acid supplementation may be effective in diminishing the severity of EIB as well.⁷

Competitive and Elite Athletes (Summary Statements 50–53)

Exercise-induced bronchoconstriction alone in elite athletes who do not have chronic asthma may have different characteristics than EIB in elite athletes who have chronic asthma or EIB in the general population, including the pathogenesis, presentation, diagnosis, and management of the disorder and the requirement by sport-governing bodies to obtain permission to receive pharmaceutical agents to treat asthma. Airway inflammation in elite athletes may be related to the high intensity of physical training and minute ventilation and inhalation of airborne pollutants by athletes in training or competition environments.^{7,35} The diagnosis of EIB in elite athletes may be difficult because history and presentation are not reliable indicators of a diagnosis.^{1–3} Objective testing in an elite athlete to diagnose EIB and to secure permission to treat asthma is

mandated by relevant governing bodies of the specific sport in which the athlete is participating.³

The management of EIB in athletes who also have chronic asthma is similar in both recreational and elite athletes. However, the management of athletes at any level who only have EIB, unassociated with chronic asthma, is not well understood and requires additional study.^{7,35} It is unknown at present whether EIB alone represents a form of overuse syndrome of the lung in elite athletes.¹⁰

SUMMARY STATEMENTS

Pathophysiology of Exercise-Induced Bronchoconstriction

Definition and Overview

Summary Statement 1. Exercise-induced bronchoconstriction is defined as the transient narrowing of the lower airways that occurs after vigorous exercise. It may appear with or without asthma. The term *EIA* should not be used because exercise does not induce asthma but rather is a trigger of bronchoconstriction. D

Summary Statement 2. Exercise-induced bronchoconstriction occurs in response to heating and humidifying large volumes of air during a short period. The most important determinants of expression of EIB response and severity are the water content of the inspired air and/or the level of ventilation achieved and sustained during exercise. B

Summary Statement 3. Respiratory water loss at high ventilation is associated with airway cooling and dehydration and an increase in osmolarity of the airway surface. B The predominant theory of EIB is the osmotic theory, although the thermal theory may also play a role. C

Summary Statement 4. Exercise itself is not necessary to cause airways to narrow; voluntary hyperpnea of dry air may induce bronchoconstriction similar to exercise. Eucapnic voluntary hyperpnea is used as a surrogate for exercise in the diagnosis of EIB, particularly in athletes. B

Summary Statement 5. People who have EIB without asthma associated with airway inflammation and the presence of eosinophils are likely to be responsive to corticosteroids. B

Summary Statement 6. Exercise-induced bronchoconstriction is accompanied by release of mediators such as PGs, LTs, and histamine. B

Summary Statement 7. In approximately half of patients who have EIB, there is an interval of refractoriness lasting approximately 2 to 3 hours immediately after an episode of EIB during which additional exercise produces little or no bronchoconstriction. B

Genetics and Environment

Summary Statement 8. Gene expression and environmental interaction may be relevant to the EIB phenotype. D

Summary Statement 9. Oxidative stress caused by environmental pollutants that are inhaled during exercise may play an important role in the development and exaggeration of EIB. B

Summary Statement 10. The pathogenesis of EIB in elite athletes may relate to effects on the airways arising from humidifying large volumes of dry air over months of training with or without exposure to environmental irritants, allergens, and viral agents. D

Prevalence

Summary Statement 11. Exercise-induced bronchoconstriction is reported in most asthmatic patients. A

Summary Statement 12. Patients with more severe or less well-controlled asthma are more likely to manifest EIB than patients with less severe or better controlled disease. A

Summary Statement 13. The true prevalence of EIB in the general population is poorly defined because epidemiologic studies of EIB have not differentiated asthmatic vs nonasthmatic populations. In addition, there is no consensus for the end point indicative of a positive response, and the conditions under which exercise is performed frequently differ. B

Summary Statement 14. The prevalence of EIB in elite athletes appears to be higher than in the general population and depends on the type of sport, the maximum exercise level, and environmental conditions. B

Summary Statement 15. The prevalence of EIB varies with history, type of challenge, and conditions under which the challenge is performed. A

Summary Statement 16. The prevalence of EIB with and without asthma may be influenced by age, sex, and ethnicity. C

Diagnosis

Summary Statement 17. Self-reported symptoms alone are not reliable for diagnosis of EIB. B

Summary Statement 18. Optimal EIB management may require confirmation of the diagnosis using objective methods. A

Summary Statement 19. Self-reported symptom-based diagnosis of EIB in the elite athlete lacks sensitivity and specificity and establishes the necessity for standardized, objective challenges using spirometry. B

Summary Statement 20. The indirect challenge (eg, exercise or surrogate such as EVH) is preferred over a direct challenge (eg, methacholine) for assessing EIB in the elite athlete. D

Summary Statement 21. Eucapnic voluntary hyperpnea is the preferred surrogate challenge for the elite athlete participating in competitive sports. D

Summary Statement 22. The intensity of the exercise challenge for the elite athlete should be 95% or greater than actual or estimated HR_{max} , and dry medical-grade air should be used in performing the challenge. D

Summary Statement 23. Hyperosmolar aerosols may also be used as surrogates to exercise. C

Differential Diagnosis

Summary Statement 24. Exercise-induced laryngeal dysfunction, primarily VCD and other glottic abnormalities, may be elicited by exercise and mimic EIB. Inspiratory stridor is

a differentiating hallmark sign with EILD and not with EIB alone. Flattening of the inspiratory curve on spirometric maneuver may be seen concomitant with symptoms. Exercise-induced laryngeal dysfunction may occur alone or with EIB. Failure to respond to asthma management is a key historical feature suggesting EILD. C

Summary Statement 25. Exercise-induced dyspnea and hyperventilation can masquerade as asthma, especially in children and adolescents. C

Summary Statement 26. Shortness of breath with exercise may be associated with underlying conditions due to obstructive lung disease, such as chronic obstructive pulmonary disease (COPD), or restrictive lung physiology, such as obesity, skeletal defects (eg, pectus excavatum), diaphragmatic paralysis, and interstitial fibrosis. B

Summary Statement 27. Shortness of breath accompanied by pruritus and urticaria, with varying other systemic symptoms, suggests EIA rather than EIB. C

Summary Statement 28. In the absence of objective evidence of EIB, breathlessness with exercise, with or without chest pain, may be caused by cardiovascular, pulmonary, or gastroenterologic mechanisms other than asthma. Appropriate cardiopulmonary testing and/or referral to a cardiologist, pulmonologist, or gastroenterologist may be necessary. B

Summary Statement 29. Exercise-induced dyspnea is seen as a physiologic limitation in otherwise healthy active individuals without bronchospasm. C

Summary Statement 30. The association between EIB and gastroesophageal reflux disease (GERD) is controversial, and probably there is no relationship. A

Summary Statement 31. Psychological factors need to be considered in the differential diagnosis of EIB. D

Summary Statement 32. Dyspnea on exertion, which is prevalent in otherwise healthy, obese individuals, is not associated with EIB. C

Summary Statement 33. Mitochondrial enzyme deficiency with myopathy is a rare cause of exercise limitation. D

Therapy

Introduction

Summary Statement 34. Frequent EIB in asthmatic patients suggests inadequate asthma control and requires patient re-evaluation to determine the need for additional therapy. D

Summary Statement 35. Failure of appropriate pharmacotherapeutic agents to prevent EIB indicates the need to re-evaluate the diagnosis. D

Summary Statement 36. Several pharmacotherapeutic agents are effective when given for the prevention or attenuation of EIB. They differ in their mechanisms of action and overall effectiveness. In addition, there is both inpatient and outpatient variability in responsiveness. A

Summary Statement 37. Medications may differ in effectiveness over time because of variability of asthma, environmental conditions, intensity of the exercise stimulus, and tachyphylaxis. A

β_2 -Adrenergic Receptor Agonists

Summary Statement 38. Inhaled β_2 -adrenergic receptor agonists are the most effective group of agents for short-term protection against EIB and for accelerating recovery of FEV₁ to baseline when given after a decrease in FEV₁ after exercise. A

Summary Statement 39. When given as a single dose or on an intermittent basis, SABAs and LABAs may protect against or attenuate EIB; SABAs are usually effective for 2 to 4 hours and LABAs for up to 12 hours. A

Summary Statement 40. Daily use of β_2 -adrenergic agents alone or in combination with ICS usually will lead to tolerance manifested as a reduction in duration and/or magnitude of protection against EIB and a prolongation of recovery in response to SABA after exercise. Therefore, monotherapy with adrenergic agents is generally recommended for use only on an intermittent basis for prevention of EIB. A

Summary Statement 41. Regular (ie, daily) use of β_2 -agonists for EIB leads to relative loss of efficacy of the agent. A

Leukotriene Inhibitors

Summary Statement 42. Daily therapy with LT inhibitors does not lead to tolerance and can be used for intermittent or maintenance prophylaxis; however, it provides protection that may not be complete and has no use to reverse airway obstruction when it occurs. A

Mast Cell Stabilizers

Summary Statement 43. Cromolyn sodium and nedocromil sodium (currently not available in the United States in an inhaled form) when inhaled shortly before exercise attenuate EIB but have a short duration of action. They do not have a bronchodilator activity. They may be effective alone or as added therapy with other drugs for EIB. A

Inhaled Corticosteroids

Summary Statement 44. Although ICS therapy can decrease the frequency and severity of EIB, its use does not necessarily eliminate the need for additional acute therapy with β_2 -adrenergic agonists or other agents. A

Summary Statement 45. Inhaled corticosteroid therapy does not prevent the occurrence of tolerance from daily β_2 -agonist use. A

Anticholinergic Agents

Summary Statement 46. Although ipratropium bromide has been inconsistent in attenuating EIB, a few patients may be responsive to this agent. A

Methylxanthines, Antihistamines, and Other Agents

Summary Statement 47. Drugs in several other pharmacotherapeutic classes, including theophylline, antihistamines, calcium channel blockers, -adrenergic receptor antagonists, inhaled furosemide, heparin, and hyaluronic acid, have been examined for actions against EIB with inconsistent results. B

Nonpharmacologic Therapy

Summary Statement 48. Preexercise warm-up may be helpful in reducing the severity of EIB. A

Summary Statement 49. Reduction of sodium intake and ingestion of fish oil and ascorbic acid supplementation may be helpful in reducing the severity of EIB. A

Competitive and Elite Athletes

Summary Statement 50. Exercise-induced bronchoconstriction alone in elite athletes may have different characteristics than EIB with asthma in elite athletes or EIB in the general population. These divergent characteristics may include pathogenesis, presentation, diagnosis, management, and the requirement by governing bodies to obtain permission to receive pharmaceutical agents. D

Summary Statement 51. Airway inflammation in elite athletes may be related to the high intensity of physical training, high minute ventilation, and inhalation of airborne pollutants and allergens. D

Summary Statement 52. The diagnosis of EIB, whether alone or with asthma, in elite athletes may be difficult because history and presentation are not reliable. Objective testing is necessary to diagnose the condition accurately. A

Summary Statement 53. In general, the treatment of EIB in patients who have asthma is similar in both recreational and elite athletes. However, the efficacy of therapy for EIB alone in athletes at any level is not well established. A

Pathophysiology of Exercise-Induced Bronchoconstriction

Definition and Overview

Summary Statement 1. Exercise-induced bronchoconstriction is defined as the transient narrowing of the lower airways that occurs after vigorous exercise. It may appear with or without asthma. The term *EIA* should not be used because exercise does not induce asthma but rather is a trigger of bronchoconstriction. D

Exercise-induced bronchoconstriction is usually measured as a reduction in FEV₁ after exercise and defined as a decrease of 10% to 15% of the preexercise value.^{1–6,36–40} Exercise-induced bronchoconstriction is a manifestation of BHR and is often the first sign of asthma. A β_2 -agonist, by inhalation, is the most widely used treatment given immediately before exercise to prevent EIB. In clinically recognized asthmatic patients, EIB is associated with airway inflammation, and decrement of EIB will occur in most cases within 12 weeks of daily treatment with ICS alone.^{9,28,41,42}

Summary Statement 2. Exercise-induced bronchoconstriction occurs in response to heating and humidifying large volumes of air during a short period. The most important determinants of expression of EIB response and severity are the water content of the inspired air and/or the level of ventilation achieved and sustained during exercise. B

At resting ventilation under most inspired air conditions, the air is usually fully conditioned by the time it has passed

over the nasal mucosa. During exercise, there is a switch from nasal to mouth breathing as ventilation increases to 30 L/min or more.⁴³ The number of generations of the lower airways required to complete the conditioning of the air depends on the following: ventilation rate, water content and temperature of the inspired air, temperature of the airway wall, and availability of airway surface liquid (ASL) to provide humidification.⁴⁴ The cumulative surface area of the airways and the ASL volume are sufficiently large that it is unlikely that unconditioned air will enter the alveoli. In contrast, the cumulative surface area and the volume of the ASL in the first 10 generations are surprisingly small—at approximately 190 cm² with less than 1 mL in volume of ASL—compared with the cumulative surface area to generation 17 (3,100 cm²).⁴⁵

The most important determinants of EIB severity are the water content of the inspired air and the level of ventilation achieved and sustained during exercise. The dryer the inspired air and the higher the ventilation during the 6 to 8 minutes of exercise, the greater the likelihood of a positive response to exercise in a susceptible person.^{46–49}

Direct measurements of airway temperature have shown that inspired air at 22°C and 9 mg H₂O at 60 L/min is not fully conditioned at the fifth airway generation.⁵⁰ Mathematical models have demonstrated that under these conditions the air is fully conditioned by the 12th airway generation. Airways of generation 12 have a diameter about 1 mm.⁵¹ Full conditioning of inspired air of subfreezing temperature involves more than 12 generations, and these very small airways that have a diameter less than 1 mm are susceptible to injury from the cold air.

When air fully saturated with water is inspired at body temperature during exercise, EIB is markedly inhibited or even completely prevented.^{48,52,53} Any intervention that reduces respiratory water loss reduces severity of EIB, such as reducing the intensity and duration of exercise,^{17,54} reducing the ventilation by aerobic conditioning,⁵⁵ increasing water content of the inspired air, and breathing via the nose or using a face mask.^{56–58} All of these interventions are consistent with the observation that the greater the number of generations of airways recruited to condition the air, the more likely EIB will occur and the more severe it will be for a given individual. The involvement of peripheral airways in the conditioning process is probably important because the density of mast cells is the greatest in the periphery.^{12,51,59–61}

Summary Statement 3. Respiratory water loss at high ventilation is associated with airway cooling and dehydration and an increase in osmolarity of the airway surface. B The predominant theory of EIB is the osmotic theory, although the thermal theory may also play a role. C

Respiratory water loss at high ventilation is associated with cooling of the airways⁵⁰ and dehydration of the ASL volume with subsequent increase in osmolarity.⁶² Cooling and hyperosmolarity probably act independently as stimuli for the airways to narrow. Cooling is a mechanical stimulus that involves reactive hyperemia of the bronchial vasculature,^{63,64}

whereas hyperosmolarity induces mediator release and subsequent contraction of the BSM.^{12,13}

When air of subfreezing temperature is inspired during exercise, airway cooling causes vasoconstriction of the bronchial vasculature.⁶⁵ On cessation of exercise, when ventilation falls and the airways rewarm, a reactive hyperemia with vascular engorgement and edema of the airway wall occurs.⁶⁶ This is known as the thermal theory of EIB. The proponents of this theory consider that the vascular events explain all the airway narrowing seen with EIB.^{63,64} The thermal theory of EIB does not include BSM contraction or mediator release in the mechanism of EIB.^{63,64}

The osmotic theory of EIB developed when it was demonstrated that abnormal cooling of the airways was not a prerequisite for EIB and that the airways of asthmatics were sensitive to small changes in osmolarity.^{62,67–81} The osmotic theory has been supported by observations that mast cells and eosinophils release mediators in response to an increase in osmolarity.^{82–84}

Dehydration of the ASL causes an increase in its ion content and osmolarity when water on the airway surface is evaporated faster than it is returned by condensation or from the epithelial cell or submucosa.^{51,85} Dehydration of the ASL has been demonstrated by a marked reduction in mucociliary clearance during dry air breathing both in asthmatic and healthy patients.⁸⁶

The cooling and osmotic theories of EIB may operate together under conditions of breathing cold dry air when the vascular effects may amplify the contractile effect of the mediators.^{12,13} As the temperature of the inspired air increases toward body temperature, the osmotic effects of water loss will become more important than cooling. Airway cooling does not play a significant role in the EIB that occurs with breathing hot dry air.^{69–71,87,88}

Summary Statement 4. Exercise itself is not necessary to cause airways to narrow; voluntary hyperpnea of dry air may induce bronchoconstriction similar to exercise. Eucapnic voluntary hyperpnea is used as a surrogate for exercise in the diagnosis of EIB, particularly in athletes. B

Exercise itself is not necessary to cause airways to narrow; voluntary hyperpnea of dry air may induce bronchoconstriction similar to exercise.^{49,89–92} Thus, EVH of dry air containing 4.9% to 5% carbon dioxide is often used as a surrogate for exercise in the diagnosis of EIB, particularly in athletes.^{93–96} The EVH test for EIB was standardized many years ago to evaluate military recruits for EIB.^{92,97,98} Eucapnic voluntary hyperpnea is recommended by the European Respiratory Society/European Academy of Allergy and Clinical Immunology Task Force⁹⁹ to identify EIB in athletes and is included in the World Anti-Doping Agency assessment of asthma (www.wada-ama.org/Documents/Science_Medicine/). Eucapnic voluntary hyperpnea should not be performed on a patient who has an FEV₁ below 70% of predicted and only with caution if FEV₁ is below 80% of predicted because of the risk of a serious decrease in FEV₁.

Summary Statement 5. People who have EIB without asthma associated with airway inflammation and the presence of eosinophils are likely to be responsive to corticosteroids. B

In people with clinically recognized asthma, EIB is related to the presence of eosinophils; those with EIB tend to have a greater concentration of eosinophils in sputum relative to those without EIB.¹⁰⁰ Sputum eosinophils have also been reported to increase acutely in response to exercise,¹⁰¹ although this has not been a universal finding.¹⁰² There is an association between the percentage of eosinophils in sputum and the severity of EIB.¹⁰³ An association between peripheral blood eosinophil counts and severity of EIB has also been reported.¹⁰⁴ As expected, the reduction in EIB severity after treatment with ICS is also accompanied by a reduction in percentage of eosinophils in sputum.^{8,9,28,32,41,42,103,105,106}

Summary Statement 6. Exercise-induced bronchoconstriction is accompanied by release of mediators such as PGs, LTs, and histamine. B

There is both indirect and direct evidence for mediators being important in EIB. Some studies show the benefit of specific pharmacologic antagonists and mast cell-stabilizing agents on EIB, and other studies show increases in mediator concentrations in sputum and urine after exercise.

Mast cells have long been thought to be an important source of mediators for EIB because drugs that prevent mast cell release of mediators or antagonize the effects of these mediators reduce the severity of EIB.^{79,80,107–118} Prostaglandin 2 (PGD₂), is the major mast cell specific mediator in EIB. Mast cells also generate LTs and histamine.^{83,119} In addition to mast cells, eosinophils are also a potential source of LTs.⁸⁴ These same mediators, beyond acting on BSM to cause contraction and the airways to narrow, can also increase microvascular permeability (MVP), leading to edema.¹²⁰

The increased urinary excretion of LTE₄ and the stable urinary metabolite of PGD₂, 9 α ,11 β PGF₂ have provided evidence that arachidonic acid metabolites are probably involved in EIB.^{11,30,32,80,116,117} Although early studies found only small increases in histamine in arterial plasma in response to exercise,^{121,122} recent studies of sputum have confirmed that EIB is associated with release of histamine.^{116,117}

Importantly, inhibition of the release of these mediators is associated with reduction in severity of EIB.³² Sodium cromoglycate and the related compound nedocromil sodium inhibit EIB by stabilizing mast cells.^{79,80,109,111,114,115} This has been demonstrated by the finding that PGD₂ is released in response to the osmotic stimulus mannitol.^{30,123}

Leukotriene antagonists modify the maximum decrease in FEV₁ and the time of recovery after EIB.^{112,124} Histamine antagonists have been observed to have variable activity on EIB with few investigators reporting complete inhibition of EIB in most patients.^{107,110,113} It is of interest that histamine antagonists may be more effective in preventing EIB in those exercising strenuously.¹²⁵ With strenuous exercise, the more peripheral airways are likely to be involved in conditioning the air and histamine release from mast cells is more likely to occur.^{61,125}

Many mediators increase MVP,¹²⁰ an event that is thought to play a role in EIB. An increase in MVP is usually identified by a change in markers of microvascular leakage. Thus, an increase in MVP after exercise has been demonstrated by a change in sputum-serum ratio of albumin.¹²⁶ The increase in this ratio has been shown to relate to severity of EIB. Furthermore, vascular endothelial growth factor and angiotensin 2 also stimulate MVP, and levels of these substances measured after exercise are also related to severity of EIB.^{126–128} The potential role for increased MVP in the pathophysiology of EIB is clear in that it could act to amplify the airway narrowing caused by BSM contraction. However, MVP may also play a role in the pathogenesis of EIB in relation to airway injury (see below).

Summary Statement 7. In approximately half of patients who have EIB, there is an interval of refractoriness lasting approximately 2 to 3 hours immediately after an episode of EIB during which additional exercise produces little or no bronchoconstriction. B

In approximately half of patients who have EIB after the initial episode of EIB, there is an interval of refractoriness lasting approximately 1 to 3 hours during which additional exercise produces little or no bronchoconstriction.^{129–131} The precise mechanism for this refractory period is not fully understood; however, recent findings suggest that the refractory period could be explained by BSM becoming tolerant to the effect of mediators.¹³² The refractory period may be prevented by taking nonsteroidal anti-inflammatory drugs for 3 days. Nonsteroidal anti-inflammatory drugs inhibit the cyclooxygenase pathway and thus production of several PGs. Some investigators propose that PGE₂, an endogenous mediator that causes smooth muscle relaxation, may play a role in the development of refractoriness.^{133–136}

Genetics and Environment

Summary Statement 8. Gene expression and environmental interaction may be relevant to the EIB phenotype. D

The few studies that have investigated gene expression linked to EIB have used only asthmatic patients who express the EIB phenotype. Some studies have identified increased expression of genes involved in eicosanoid release in patients who have asthma, whereas others found no relationship.^{15,137} The polymorphism IL-13^{+2044G/A} has been strongly associated with atopy and severity of EIB in Korean children, whereas the IL-13^{1112C/T} and the haplotype of IL-13 polymorphisms were associated with LTRA responsiveness.¹³⁸

One group of researchers¹⁶ demonstrated increased expression of mucin 5AC (MUC5AC) after an exercise challenge that was associated with the severity of EIB. Mucins such as MUC5AC may significantly contribute to airflow obstruction.¹³⁹ Moreover, associations were found between levels of MUC5AC and cys-LTs and neurokinin A in the airways and suggested a mechanism whereby MUC5AC is released by cys-LT activation of sensory airway nerves.¹⁶ Using phenotypically distinct asthmatic patients, those without and with

EIB, the same authors¹⁴⁰ identified different patterns of gene expression. In particular, after exercise challenge, expression of transglutaminase 2 was increased. Transglutaminase 2 was further shown to augment enzymatic activity of phospholipase A₂ group X (sPLA₂-X), an enzyme that catalyzes eicosanoid (cysteinyl-LTs) formation. High sPLA₂-X has been identified in asthmatic patients and has been shown to increase further after exercise challenge in asthmatic EIB phenotypes.¹⁴¹

Summary Statement 9. Oxidative stress caused by environmental pollutants that are inhaled during exercise may play an important role in the development and exaggeration of EIB. B

Oxidative stress is an important feature in the pathogenesis of asthma¹⁴² and may enhance EIB-related mediator production.¹⁴³ Oxidative stress probably results from both an increase in reactive oxygen/nitrogen species and depletion of antioxidants in the airways. Inhalation of high particulate matter (PM) air during exercise has been shown to reduce alveolar contribution to eNO, decrease total nitrate, and increase lipid peroxidation in exhaled breath condensate and was related to postexposure decrease in FEV₁.¹⁴⁴ Increased concentration of 8-isoprostane, a marker of airway oxidative stress, in exhaled breath condensate is associated with EIB.¹⁴³

Ozone is a major oxidant generated from a photochemical reaction of hydrocarbons and nitrogen oxides from combustion emissions that has been shown to trigger EIB.^{145,146} Exposure studies show decrements in FEV₁ ranging from 0% to 48% with mild ozone exposure.^{147,148} Glutathione S-transferases are found in high concentrations in the lung and are involved in antioxidant defense pathways¹⁴⁹ and glutathione homeostasis.¹⁵⁰ A single nucleotide polymorphism of glutathione S-transferase π ile105 has recently been implicated in the development of new onset asthma and EIB in children who participate in team sports in high ambient ozone, with a hazard ratio for new-onset asthma of 6.15 (range, 2.2–7.4).¹⁵¹

Airborne PM generated from combustion emissions is associated with the formation of reactive oxygen species that influence mediator release from airway cells.^{152,153} Exposure to high levels of PM is associated with increased emergency department visits,^{154,155} increased bronchodilator use,¹⁵⁶ and bronchoconstriction after exercise in asthmatic and nonasthmatic individuals.¹⁵⁷ High levels of airborne PM have been identified at indoor ice rinks that use a fossil-fueled (gas, propane, or natural gas) ice resurfacer^{158–160} and at athletic fields that are close to high automobile and truck traffic.¹⁶¹ The ice rink air exposure during exercise coupled with a genetic susceptibility could account for the observed high prevalence of asthma and EIB in athletes exercising in an ice rink.^{159,160,162–165} Cysteinyl-LTs appear to be important mediators in PM-induced bronchoconstriction; 1 study¹⁶⁶ found increased protection against EIB in a high pollution environment by the LTRA montelukast. Diesel exhaust particles have been shown to enhance the induction of allergy and to increase inflammatory cytokines and cysteinyl-LTs in animal models.¹⁶⁷ Likewise, secondhand tobacco smoke has been shown to increase cysteinyl-LT-related albuterol usage

and montelukast responsiveness in children exposed to tobacco smoke.¹⁶⁸

Summary Statement 10. The pathogenesis of EIB in elite athletes may relate to effects on the airways arising from humidifying large volumes of dry air over months of training with or without exposure to environmental irritants, allergens, and viral agents. D

Exercise-induced bronchoconstriction is commonly seen in highly competitive athletes, especially in those competing at the national and international level.^{1,2,95,164,169–171} The reason that this condition is seen in such well-trained athletes is not fully understood, but airway injury and the environment may play an important role, particularly in athletes reporting onset of asthma at older than 22 years of age.^{10,35}

Injury of the airways may arise from conditioning large volumes of dry air over months of training, with or without exposure to environmental irritants, allergens, and viral agents.^{158,159,171–178} Repair follows injury and the process is associated with microvascular leakage. It is possible that BHR to inflammatory mediators and pharmacologic agonists may develop as a result of repeated exposure to plasma-derived products that cause a change in the contractile properties of BSM.^{10,126,179–183}

A hypothesis has now been postulated relating pathogenesis of BHR and EIB to airway injury in athletes. This hypothesis is based on observations reported by many investigators¹⁰ that recruitment of the very small airways (<1 mm) in the air conditioning process may be accompanied by injury of the epithelium. Such injury would be associated with plasma exudation, and in the process of repetitive injury and exudation, the contractile properties of the BSM would be altered, rendering it more sensitive. One potential outcome may be BHR to methacholine as demonstrated in winter athletes.^{178,184–186} It is important to know whether methacholine responsiveness in athletes is a manifestation related to airway injury rather than a sign of “classic” asthma. The notion that BHR may be related to injury is supported by the return to normal responsiveness when athletes are out of season¹⁸⁷ or after retirement.¹⁸⁸ For summer athletes who are more likely to be atopic and have higher IgE levels, the hypothesis proposes that “passive” sensitization of the BSM can occur in vivo during the injury/exudation process. In this case, the BSM may become more responsive to contractile mediators such as LTE₄ released into the circulation in response to intense exercise.¹⁸⁹ This may explain why some athletes can benefit from use of LT antagonists and other agents that prevent release of mediators or antagonize their action.¹⁶⁶

One of the important concepts in this hypothesis is the presence of mast cells in the small airways of healthy people and their proximity to the BSM in these airways.⁶¹ The presence of mast cells adjacent to the smooth muscle or a mast cell myositis is a relatively new concept in defining asthma.^{60,190} As noted above, mast cell mediators released in the periphery provoke smooth muscle contraction and airway inflammation, which are the hallmarks of asthma.

The potential for β_2 -agonists to inhibit or enhance development of BHR and EIB also needs to be considered. There are well-known beneficial effects of β_2 -agonists: these include relaxation of BSM, stabilization of mast cells, and stimulation of Cl⁻ ion secretion and water to the airway surface. It is possible that given in the short term, β_2 -agonists reduce airway injury and microvascular leakage.¹⁹¹ By contrast, the same drugs given in the long term may cause mast cells to become more sensitive or unstable to stimuli because β_2 -receptors are downregulated with long-term use.^{192,193} The sensitivity of the smooth muscle may be increased because of an increase in number of histamine receptors¹⁹⁴ or cross talk between the relaxing and contractile pathways through the Gs protein.¹⁹⁵

There are other mechanisms whereby elite athletes could develop BHR. One is respiratory tract infection. There is a higher frequency of infections in athletes compared with controls. The reason is thought to be due to the suppressed immunity that occurs between 3 and 72 hours after exercise, known as the “open window” theory.¹⁷⁷ The role of viruses in changing reactivity may relate to effects on the β_2 -receptor. Bronchial hyperresponsiveness to histamine is prolonged when training was continued during an upper respiratory tract infection.¹⁹⁶ A low serum IgA secretion rate during a marathon was shown to be the best predictor of infection.¹⁹⁷ The finding of lymphoid aggregates in cross-country skiers¹⁹⁸ may indicate an immune response to infection.^{199,200}

The difference in the pathogenesis of EIB and BHR may be the reason there are differences in response to treatment between athletes and patients with classic asthma.^{201,202} For example, neutrophils rather than eosinophils may be more common in those with exercise-induced airway injury, making the problem less amenable to treatment with inhaled steroids.²⁰¹ By contrast, macrolides may have potential for modifying the inflammation of neutrophils.²⁰³

Forty years ago, EIB was thought to involve the release of mast cell mediators⁷ because sodium cromoglycate, a mast cell-stabilizing agent with no action on BSM, was effective in inhibiting EIB. The recognition that the rate of water loss from the airway surface was the determining factor for EIB severity led to the thermal (airway cooling) and osmotic (airway dehydration) hypotheses of EIB. After the introduction of ICS, it became clear that the pathophysiology of EIB also involved inflammation of the airways. During the last 10 to 15 years, the techniques used to measure mediators have become much more sensitive, allowing PGs, LTs, and histamine to be identified in sputum and urine after exercise. Observations that these mediators are indeed present confirm the role of the mast cell and other inflammatory cells in EIB. The increasing recognition that EIB can occur in healthy people (eg, elite athletes, military recruits, and schoolchildren) without classic signs and symptoms of asthma has led to airway injury being suggested as important in the pathogenesis of EIB. Conditioning vast volumes of dry air has the potential to injure the peripheral airways in athletes, as does

Table 1. Prevalence of EIB in Different Populations

Population	Challenge test	Sport	Prevalence of EIB, %	Reference
Adults	EVH	Fitness	19	212
High school athletes	EVH	Not stated	38	213
Adolescents	Exercise	High school	28	214
College athletes	EVH	Various athletes	35–36	2
Children from India	Exercise	Urban/rural	13/10	215
Adolescents	Exercise	Various sports	9.4	216
Olympics	Exercise	Winter sports	18–50	164
Elite	Field exercise/methacholine	Skiers	37.5/8.3	178
Elite	Exercise	Figure skaters	35	162
Elite	Exercise	Figure skaters	30	163
Elite	EVH	Summer athletes	50	4
Olympics	EVH/exercise	Winter sports	45/29	94

Abbreviations: EIB, exercise-induced bronchoconstriction; EVH, eucapnic voluntary hyperpnea.

the environment, which is increasingly being considered as important in the pathogenesis of EIB.

Prevalence

Exercise-induced bronchoconstriction is reported in most asthmatic patients but also occurs in the absence of chronic asthma. Exercise-induced bronchoconstriction is very common in athletes. The prevalence of EIB is significantly influenced by the criteria used for diagnosis. The use of self-reported symptoms to make the diagnosis of EIB will likely misdiagnose asthma in patients who do not have EIB and miss people who have the condition.^{1,204–206} Self-reported symptoms should not be relied on solely for the diagnosis of EIB without the concomitant use of spirometry and bronchial provocation challenge, including reversibility after β_2 -agonist, to confirm the diagnosis.

The prevalence of EIB is also influenced by age, sex, ethnicity, and the environmental conditions (eg, air temperature and humidity, allergen content, and pollution) in which exercise is performed. This largely affects the prevalence of EIB in different population samples (Table 1).

Summary Statement 11. Exercise-induced bronchoconstriction is reported in most asthmatic patients. A

Summary Statement 12. Patients with more severe or less well-controlled asthma are more likely to manifest EIB than patients with less severe or better controlled disease. A

Summary Statement 13. The true prevalence of EIB in the general population is poorly defined because epidemiologic studies of EIB have not differentiated asthmatic vs nonasthmatic populations. In addition, there is no consensus for the end point indicative of a positive response, and the conditions under which exercise is performed frequently differ. B

The occurrence of bronchoconstriction, especially with symptoms during or after exercise, is one of the common characteristics of asthma, but it also occurs in the absence of asthma. In asthmatic patients, EIB in itself is a marker of poor control and suggests the need to initiate or step up therapy.^{33,207} The reported prevalence of EIB varies consider-

ably, depending on the severity and control of asthma in the patients being evaluated.

Exercise-induced bronchoconstriction is a common disorder in children.^{208,209} The few data available from cross-sectional studies performed over time seem to indicate that the prevalence of EIB is increasing, in parallel with the increasing prevalence of asthma. A survey in 1973²¹⁰ and again in 1988²¹¹ examined 12-year-old children in South Wales who had a decrease of 15% in peak expiratory flow rate after a free run for 6 minutes. On the basis of this measure, they found an increase in prevalence of EIB from 6.7% to 7.7%. The lower prevalence of EIB observed in 2003 (4.7%) may be confounded by the increased number of children taking ICS. Accordingly, the data from these studies showed that the percentage of children who had asthma in the last 12 months was 4.2%, 9.1%, and 15.4% and the percentages who ever had asthma was 5.5%, 12.0%, and 27.3% in 1973, 1988, and 2003, respectively. Otherwise, there is a paucity of data from cohort studies in children examining changes in prevalence of EIB over time and virtually no data in adults. Table 1 indicates that prevalence varies widely, depending on the factors that are being studied and controlled, such as level of intensity of exercise, population under study, and environmental condition.

A study using EVH²¹² reported a presumed prevalence of EIB of 19.4% in 212 adults without a history of asthma. In 2007, another researcher^{217,218} reported that individuals with a family history of asthma may have a higher prevalence of EIB after running than those who do not have a positive family history. Exercise-induced bronchoconstriction is also more frequent in atopic individuals,^{171,219} including those with allergic rhinitis,²²⁰ and after respiratory viral infections and other respiratory diseases.²²¹ It is unclear what the relationship is between the natural history of EIB when not associated with asthma and the subsequent development of asthma.

Summary Statement 14. The prevalence of EIB in elite athletes appears to be higher than in the general population

and depends on the type of sport, the maximum exercise level, and environmental conditions. B

The need for objective data to diagnose EIB is especially important in the elite athlete because of the establishment of definitive criteria that must be met to allow the use of β_2 -agonists in or out of competition.³⁵ No test for EIB is fully sensitive or specific, and thus, no test is sufficient to substantiate or exclude the diagnosis of EIB in all athletes. Additional variability, such as the passage of time and different environments, may affect the ability to detect BHR.

Reports of the occurrence of asthma symptoms in elite athletes have varied from none to 61%,^{222,223} depending on the sport and environment.^{1,158,160,162,164,174,222,224–228} In fact, higher prevalence rates are reported in certain populations, such as in elite endurance athletes, and in unique environments, such as competitive skaters and cross-country skiers.^{35,165,228} Some investigators^{222,223} suggested that endurance athletes, whether summer or winter, had considerably more symptoms than athletes participating in less aerobic sports. Overall, this study suggested that based on symptoms as many as 1 in 4 athletes had EIB. Using a questionnaire in recreational athletes, a similar prevalence of asthma symptoms was reported.²²⁹ However, these data do not suggest that the individuals who report asthma are the same as those who have a positive exercise challenge. However, a similar prevalence of EIB in winter Olympic athletes has been reported based on objective data using an exercise challenge.¹⁶⁴ Certain populations have a higher than expected prevalence based on unique circumstances, such as the high prevalence of EIB in skaters (20%–35%) that has been attributed to high emission pollution from ice cleaning equipment and cold dry air.^{158,159} A similar example is the extremes of cold dry air, such as that to which cross-country skiers are exposed, which may increase the prevalence of EIB to 30% to 50%.²³⁰ Similarly, another study¹⁷⁰ found as many as 78% of elite cross-country skiers have symptoms and/or hyperresponsive airways.

Athletes participating in the 1996 summer sports also have variations in EIB prevalence, which may depend on the sport in which they participate. For example, long distance runners may have a prevalence of 17%, whereas speed runners may have a prevalence of 8%.¹⁷⁴ Whether these differences are significant may depend on how the test was performed rather than on a difference in the sports for athletes who expend a similar amount of work. By survey, none of the US Olympic divers and weightlifters had symptoms, whereas 45% of

Table 2. Percentage of Athletes Participating in the Olympic Summer Games Notifying or Making Application to Use IBAs

Games	Total No. of Athletes	No. (%) Applying to Use IBAs
Atlanta (notified)	10,677	383 (3.6)
Sydney (notified)	10,649	607 (5.7)
Athens (approved)	10,653	445 (4.2)
Beijing (approved)	10,810	781 (7.2)

Abbreviation: IBAs, inhaler β_2 -agonists.

Table 3. Athletes Notifying or Making Application to Use IBAs in 3 Sequential Olympic Summer Games Based on Type of Sport

Sport	No. (%) Applying to Use IBAs		
	Sydney (2000)	Athens (2004)	Beijing (2008)
Triathlon	24 (24.0)	10 (10.1)	28 (25.7)
Cycling	81 (17.0)	63 (13.5)	87 (17.3)
Modern pentathlon	7 (14.6)	8 (12.5)	9 (19.1)
Swimming	163 (11.2)	121 (8.4)	202 (19.3)
Rowing	48 (8.8)	34 (6.1)	70 (12.8)
Canoeing	23 (7.0)	16 (4.9)	9 (2.8)
Athletics (track and field)	87 (4.1)	87 (4.4)	117 (5.8)
Field hockey	15 (4.3)	12 (3.1)	41 (10.7)

Abbreviation: IBAs, inhaler β_2 -agonists.

mountain bikers experienced symptoms, consistent with the hypothesis that endurance sports have a higher prevalence of associated EIB during sport participation.²²²

Poor air quality may also be associated with a high prevalence of EIB.²³¹ In swimmers, chloramines above the water may trigger EIB. Interestingly, swimmers with longer duration of exposure tend to have a higher prevalence of EIB.²³² Discontinuation of swimming resulted in a decreased incidence of EIB.¹⁸⁸ As noted above, the high prevalence of EIB in skaters (20%–35%) has been attributed to high emission pollution from ice cleaning equipment and cold dry air.^{158,159} Seasonal variation of EIB is also described in Olympic athletes.¹⁷¹ For example, when using a 6.5% decrease in FEV₁ with running, 28% of runners had probable EIB. Of these runners, 22% had EIB that occurred only in the winter and 7% had EIB only during the pollen season.¹⁷¹ Seasonal difference was also demonstrated by another investigation,²³³ which found that 35% of runners training in the cold reported an increased prevalence of EIB compared with summer when the prevalence was less.

Data collected from the summer Olympics between the years 1992 and 2008 have documented an increasing percentage of elite athletes reporting asthma. This trend follows a similar increase noted in the general population. Confounders, such as the requirement of the Olympic Committee to have an objective diagnosis of EIB to permit utilization of albuterol, may account for the large increase of percentages between 2004 and 2008. Similarly, the difference in reporting requirements over the years makes the data difficult to interpret. Table 2 contains the data on asthma reporting in the Summer Games. Table 3 contains data demonstrating the influence that particular sports have on the reporting of asthma from athletes. Again, the data may be biased by differences in reporting requirements for each of the Games.

Summary Statement 15. The prevalence of EIB varies with history, type of challenge, and conditions under which the challenge is performed. A

By examining responses to history questions on their intake forms required by the US Olympic Committee athletes participating in the 1996 Summer Olympic Games, investi-

gators²²² found 0% to 45% of summer athletes, depending on sport, answered questions compatible with having EIB. The prevalence varied significantly among different sports, with nonendurance sports having minimal levels while endurance sports had higher prevalences. By using the same data extraction method, the same researcher²²³ found that up to 60.7% of athletes participating in Nordic skiing events responded to questions that suggested they had EIB.

A limitation of determining prevalence by survey is evident by work performed by investigators,¹ who found that those who had symptoms did not necessarily have a positive challenge result and those who had a positive challenge result did not necessarily have symptoms. Other researchers²³⁴ demonstrated this in adolescent athletes with and without a history suggesting EIB. Those with a history of abnormal baseline spirometry suggesting a risk for EIB had a positive exercise challenge (at least a 15% decrease in FEV₁) in 8 of 48 tested (17%), whereas in those students who had no risk factors, 14 of 118 (12%) had a positive challenge result. Thus, symptoms suggest a positive challenge result, but most of those who had positive challenge results were not at risk. This suggests that patients with a history of chronic asthma are more likely to have positive exercise challenges, but individuals without a history of asthma will also have a positive challenge result.

Another study⁴ used different techniques in an attempt to clarify prevalence of EIB. These investigators challenged 50 elite athletes with and without a history of asthma documented by questionnaire with methacholine provocation and with EVH. The results showed that of the 42 athletes who reported respiratory symptoms, 9 had a positive methacholine test result and 25 had a positive EVH test result. Methacholine had an excellent negative predictive value, but only a 36% sensitivity for identifying those with a positive EVH test result.

These findings are consistent with the observations found in 2 more studies,^{227,235} which demonstrated that EIB and asthma symptoms do not correlate well with eNO, results of bronchial lavage, or challenges with AMP or histamine challenge. Data from this group confirmed that typical signs and symptoms consistent with asthma are not always found in athletes who have EIB. Conversely, some athletes thought to have EIB probably do not. It has been demonstrated that symptoms are neither sensitive nor specific to suggest a positive EVH test result as evidence for EIB.² This testing found that 36% of college athletes without symptoms had a comparable decrease of 10% in FEV₁ and a similar number (35%) of those with symptoms had such a decrease with EVH. Other investigators²³⁶ challenged 33 elite swimmers with EVH challenge, 8-minute swim challenge, and 8-minute laboratory cycle challenge. A 10% decrease in FEV₁ was considered positive; 55%, 3%, and 12% were considered positive by EVH, swim test, and cycle test, respectively.

Summary Statement 16. The prevalence of EIB with and without asthma may be influenced by age, sex, and ethnicity. C

Researchers²³⁷ exercised 15,241 children using a 6-minute run and a decrease of 15% for peak flow as an indicator of EIB. In this cohort, girls (8.5%) were more likely than boys (6.4%) and those from urban settings (8.9%) were more likely than those from rural environs (7%) to have a positive challenge result. As expected in all populations, symptoms were a poor predictor of positive challenge results.

The frequency and severity of asthma may vary by sex, with males having greater frequency during childhood but females having more severe asthma during adulthood.^{238,239} In contrast to the above findings,²³⁷ another study²⁴⁰ failed to demonstrate sex differences in EIB. However, these investigators found that with increasing age, the frequency of EIB decreased. Frequency of asthma and EIB may also vary by sex in elite athletes. In winter sports, females appear to exceed males in prevalence of EIB. In US Olympic winter sports, the prevalence of EIB by exercise challenge test was 26% in female and 18% in male athletes with a combined percentage of 23%.¹⁶⁴ Using questionnaires and methacholine challenges, investigators²⁴¹ found that the prevalence of exercise-associated asthma symptoms and BHR was higher in female than male athletes. When using EVH as a surrogate challenge for EIB, another study² failed to find such a difference in prevalence between the sexes.

Urbanization has been shown to be associated with an increase in prevalence of asthma and EIB. Using a free run and peak flow testing for EIB, rural children, urban poor, and wealthy urbanites had a prevalence of 0.1%, 3.1%, and 5.8%, respectively.²⁴²

A standardized free running test with peak flow monitoring demonstrated that African Americans had a higher prevalence of EIB than European Americans (13% and 2%, respectively).²⁴³ When assessing 9-year-old children with cycle ergometry in Great Britain, ethnicity differences in EIB were also evident, with Asian children (originating from the Indian subcontinent) having a prevalence of 3.6 times higher than that of white, inner-city children.²⁴⁴

In summary, EIB prevalence is affected by age, sex, ethnicity, and urbanization. Elite athletes may have a high prevalence of EIB, which can be associated with extreme atmospheric conditions, such as high levels of pollen, pollution, cold air, and chemicals, particularly in the training environment. Exercise-induced bronchoconstriction can be demonstrated in people without symptoms, but symptoms are not a sensitive predictor of EIB. Prevalence of EIB varies by the test used to detect it.

Diagnosis

Summary Statement 17. Self-reported symptoms alone are not reliable for diagnosis of EIB. B

Exercise-induced bronchoconstriction in the athlete may be accompanied by symptoms of cough, wheeze, chest tightness, shortness of breath, or excessive mucous production. However, a clinical diagnosis of EIB based on self-reported symptoms after exercise alone may be misleading because often objective measures cannot confirm asthma as the cause of the

symptoms.¹ Likewise, absence of self-reported symptoms also may be misleading because this could reflect lack of patient perception of EIB. Approximately half of elite winter sport athletes who report symptoms with exercise have normal airway function, and approximately half of those who report no symptoms will demonstrate bronchoconstriction after exercise or other indirect challenges.¹ This study documents the necessity of objective spirometry using a standardized protocol for diagnosis of EIB. Another investigation² corroborated that symptoms were not predictive of EIB in a study of college students. Using EVH with a 10% or greater decrease in FEV₁ to diagnose EIB, this study² identified a prevalence of 36% in those with no symptoms and 35% in those reporting symptoms of EIB. Athletes with high ventilation activities were significantly more symptomatic (48%) than athletes in low ventilation sports (25%; $P = .02$) but showed no difference in EIB prevalence between the groups when challenged. It is therefore essential to confirm EIB by objective measures generally following FEV₁ by using standardized tests.³

Summary Statement 18. Optimal EIB management may require confirmation of the diagnosis using objective methods. A

Although there is no single test that will identify all individuals who have EIB, indirect challenges, including exercise, EVH, inhaled powdered mannitol, nebulized hypertonic saline, or AMP, are more effective in identifying EIB than are direct challenges such as histamine or methacholine.^{18,245–247} In addition, indirect challenges are recommended³ for monitoring asthma therapy because BHR is most often related to inflammation and inflammation is diminished by ICS treatment for asthma.

The indirect challenges release mediators of inflammation, including LTs, PGs, and histamine, that provoke airway smooth muscle contraction.¹³ Therefore, indirect challenges may reflect the severity of airway inflammation.²⁴⁷ In contrast, direct challenge agents act directly on airway smooth muscle receptors to provoke bronchoconstriction, exclusive of airway inflammation.³

Challenges with pharmacologic agents that act directly on airway smooth muscle such as methacholine may be performed in the office, clinic, or hospital laboratory. These challenges are usually recommended to exclude a diagnosis of asthma rather than to exclude or include a diagnosis of EIB; however, they may be the only tests available. The direct challenge requires inhaling increasing concentrations of methacholine and then performing FEV₁ but not the full FVC maneuver after the administration of each dose. A decrease in FEV₁ of greater than 20% from baseline is regarded as a positive challenge result. This is documented as the PC₂₀. A PC₂₀ of greater than 16 mg/mL is interpreted as normal bronchial responsiveness, between 4.0 and 16 mg/mL is considered borderline BHR, between 1.0 and 4.0 mg/mL is considered mild BHR (positive test result), and a less than 1.0 mg/mL is considered moderate to severe BHR.⁶ Although the direct challenge is used as a screening test for chronic asthma, it

has low sensitivity for EIB because it reflects the effect of only a single agonist and is not recommended as a screening tool for EIB.^{4,246}

Exercise Challenge

The lack of strict adherence to the components of the consensus statement published by the ATS⁶ on a standardized exercise challenge for EIB has led to variability in documented prevalence of EIB.³⁵ Diagnosis by exercise should be made by using a challenge that is standardized for duration of exercise and for intensity by standardizing minute ventilation and water content of inhaled air.^{17,49,53,54,248,249} Spirometry is performed at baseline before the exercise challenge and at a series of defined postchallenge times, such as often as 1 to 3, 5, 10, 15, 20, and 30 to 45 minutes after 8 minutes of exercise, with 2 repeatable FEV₁ efforts (and not full FVC maneuvers) at each time point. The goal is to collect spirometry data as early and as late as practical within the time in which EIB typically occurs.³ For example, the IOC-MC Independent Panel on Asthma recommends that FEV₁ should be first recorded at 3 minutes after the completion of the challenge to overcome the problem of possible posttest respiratory muscle fatigue. Post-exercise FEV₁ values are compared with preexercise FEV₁ values to calculate a percentage change from baseline. A 10% or greater decrease in FEV₁ from an FVC or FEV₁ maneuver at any 2 consecutive time points within 30 minutes of ceasing exercise may be considered diagnostic of EIB.⁶ If a greater decrease in FEV₁ is required (ie, a decrease of 25%), then only one time point may be necessary to be diagnostic of EIB. In any case, it is important to examine the AUC to determine whether the decrease is consistent and is not likely an artifact of an inadequate spirometry effort.

A flattened or truncated inspiratory loop and a decrease in FEV₁ with no alteration in the FEV₁/FVC ratio are consistent with, but not diagnostic of, VCD.^{250,251} However, the absence of this finding does not rule out VCD as described below. The presence of stridor suggests upper airway dysfunction with or without the concomitant presence of EIB. If VCD is a consideration, then FVC maneuvers should be used to compare preexercise and immediate postexercise inspiratory loops.

Expert panels and reports from experts have developed consensus guidelines for diagnosis of EIB based on exercise challenges.^{6,252,253} Panels have suggested that the postexercise decrease in FEV₁ required to make the diagnosis must be 10%, whereas other groups have suggested that a decrease of 13% or 15% is necessary to make the diagnosis.^{37,253} A 15% decrease in FEV₁ after a “field” exercise challenge⁴⁰ and 6% to 10% decrease in FEV₁ for laboratory challenges have been recommended.^{171,226} Although there is no absolute standard cutoff postexercise decrease in FEV₁, the change in FEV₁ after exercise is normally distributed. Lowering the cutoff will result in an increased likelihood of false-positive test results for EIB, whereas raising the cutoff will result in the increased likelihood of false-negative test results for EIB. The ATS⁶ and the European Respiratory Society⁹⁹ have rec-

ommended a 10% decrease in FEV₁ after exercise as criterion for EIB, based on 2 standard deviations from the mean percentage decrease in FEV₁ in healthy individuals.³⁸

As noted above, it is essential to confirm the diagnosis of EIB using objective measures because of the lack of sensitivity and specificity of self-reported symptoms. The exercise challenge for diagnosis of EIB can be sensitive and specific if exercising minute ventilation and water content of inhaled air are standardized.^{248,249} Standardized laboratory-based EIB challenges have been performed using 6 to 8 minutes of exercise in ambient conditions (20°C-25°C, relative humidity [RH] <50%) at 80% to 90% of estimated HR_{max} as a surrogate standard to V_E.²⁵⁴⁻²⁵⁸ Maximal heart rate is often estimated by the equation of 220 - age; however, more recent analysis suggests that a more accurate regression equation to predict HR_{max} is 208 - 0.7 × age.²⁵⁹ Ideally, the exercise ventilation should be above 60% of predicted maximum (ie, greater than 21 times FEV₁).⁶ Some investigators have suggested that exercise intensity should be less than 85% of predicted HR_{max}²⁶⁰; these recommendations are based on the inappropriate assumption that exercise intensity affects catecholamine release and causes bronchodilation. This assumption is most invalid for individuals who routinely exercise because 85% HR_{max} may not be adequate to elicit a bronchoconstrictive response. Although ATS guidelines⁶ and investigations by others^{17,224,252,256-258} suggest that laboratory-based EIB testing should include an exercise challenge of 6 to 8 minutes in ambient conditions (20°C-25°C, RH <50%) at 80% to 90% predicted HR_{max}; very well-conditioned individuals may require the exercise intensity to be above a HR_{max} of 90%. Investigators¹⁷ evaluated the exercise load in relationship to EIB severity in children (aged 9-17 years old) with asthma and found that treadmill tests at 85% and 95% of calculated HR_{max} resulted in greatly different decreases in FEV₁ (Figure 2). Decrease in FEV₁ was 25.1% at 95% HR_{max} but only 8.8% at 85% HR_{max}. Only 9 of 20 individuals had decreases greater than 10% at 85% HR_{max}, whereas all 20 individuals had decreases greater than 10% at 95% HR_{max} (Figure 2). This investigation supports the necessity for a high exercise intensity, which elicits high V_E.

Dry air delivered by the use of Douglas bag, as described below in the "Laboratory Challenges" section, also may enhance the ability to achieve a positive challenge because a dehydrating stimulus is thought to be a primary trigger of EIB. However, increased vagal activity from facial cooling during cold air exercise^{261,262} or after water emersion²⁶³ has been shown to result in significant bronchoconstriction in healthy individuals and asthma patients. It has been demonstrated that facial cooling during exercise resulted in significantly lower FEV₁ after exercise with either cold or warm dry air inhalation than did similar exercise without facial cooling.²⁶¹

Field-Based Challenges

Free running^{40,226,253} and sports-specific exercise challenges^{162-164,230,264} have been shown to be valid for the evaluation

of EIB. These challenges have been demonstrated to be more sensitive than laboratory challenges at ambient temperature and RH in the laboratory in elite winter athletes.²²⁶ One study²²⁶ compared sports-specific field-based exercise challenges of varied duration to a standardized 6- to 8-minute laboratory exercise challenge. Laboratory conditions were 21°C, 60% RH, and exercise intensity of 95% of peak heart rate. Although 5 athletes were positive by both challenges, 18 of 23 athletes who tested positive for EIB by field-based challenge tested normal by the ambient condition laboratory challenge (Figure 3). These results provided strong support for the notion that water content of inhaled air is a primary stimulus for EIB. Sport-specific tests have been used with success by researchers¹⁶⁴ who studied winter Olympic athletes participating in Nordic skiing, speed skating, ice hockey, and ice skating to evaluate EIB, as did another group of researchers²⁶⁴ who used a 15-minute Nordic ski field exercise to identify EIB in cross-country skiers. Free running has been used to screen large populations of individuals because of the ease of administration. Several groups have used free running to evaluate schoolchildren.^{40,237,253,265} Good validity and reliability of a free run test to evaluate airway responsiveness in large groups of children 8 to 11 years of age have been established; a 15% decrease in FEV₁ was used

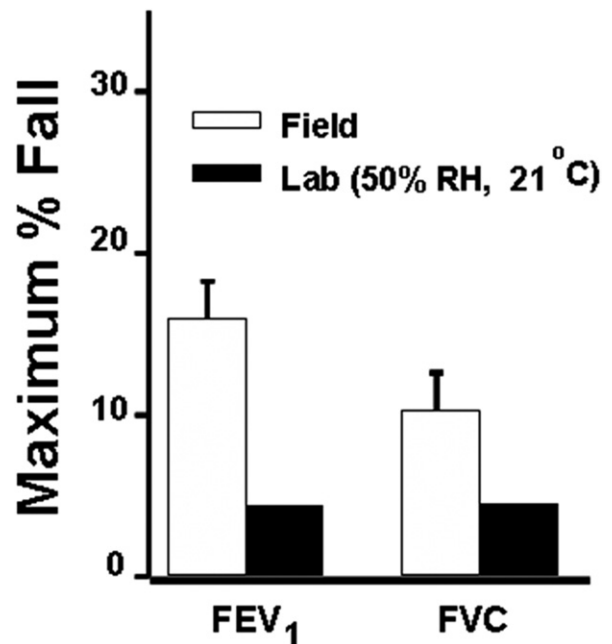


Figure 3. Of 23 elite winter athletes who tested positive for exercise-induced bronchoconstriction by a sports-specific field-based exercise challenge in cold dry air, only 5 (approximately 22%) tested positive by a laboratory treadmill run of equal intensity but under ambient conditions of 21°C, 50% relative humidity (RH). This figure displays data from the 18 athletes who tested negative by laboratory challenge. Adapted from Rundell et al.²²⁶ FEV₁ indicates forced expiratory volume in 1 second; FVC, forced vital capacity.

as cutoff criteria,²⁵³ and the 95% probability for repeated tests was a $\pm 12\%$ decrease in FEV₁.^{40,253} Researchers³⁷ also evaluated 8 studies with 232 young asthma patients and determined that a decrease in FEV₁ of 13% after exercise provided a sensitivity of 63% and a specificity of 94%, thus providing objective data to evaluate bronchial provocation challenge results. Furthermore, the cutoff value was not affected by the severity of asthma. However, the uncontrolled nature of the stimulus and changing ambient and environmental conditions impose variability that may limit free run tests as a means of monitoring therapy. A more reproducible stimulus and physiologic outcome may be needed to monitor treatment effectively.³

Laboratory Challenges

The laboratory exercise challenge for EIB can have high specificity and high sensitivity when standardized. The water content of inspired air should be below 5 mg·L⁻¹ of air.^{51,59} Because a primary trigger for EIB is the increase in osmolarity of the periciliary fluid, secondary to loss of water due to conditioning of relatively dry inspired air during exercise or hyperventilation,^{12,52} the level of minute ventilation achieved and maintained should be approximately 60% to 85% MVV.²²⁶ As noted above, this can be estimated by multiplying the baseline FEV₁ by 21 and 30, respectively.

Others have demonstrated a 50% reduction in severity of EIB when comparing exercise challenge conditions of 40% and 95% RH at ambient temperature (24% vs 12% decreases in FEV₁, respectively).²⁶⁶ Others^{248,249} examined inhaled airway temperature and EIB response and noted that severity of EIB was related to water content and not air temperature per se during exercise or EVH challenges. In these studies,^{248,249} subjects exercised or performed EVH while breathing room temperature (22°C) or cold (-1°C) dry air, with water content for both conditions less than 5 mg/L of air; exercising minute ventilations were not different among conditions. No significant difference in postexercise FEV₁ between inhaled room temperature and cold air conditions was found. Similarly, there was no difference between exercise and EVH with matched minute ventilations (Figure 4).

Treadmill or Cycle Ergometer Exercise Challenge

Current recommendations indicate that an 8-minute exercise challenge that approaches 90% or more of estimated HR_{max} by 2 minutes and is maintained for the remaining 6 minutes of the 8-minute challenge is optimal for identifying EIB. A shorter duration of 6 or 7 minutes is often used in children. This protocol should ensure a minute ventilation of approximately 60% to 85% of MVV, with the exceptions of the elite athlete for whom a higher intensity may be required or the asthmatic patient who may not be able to achieve the high ventilation. For the elite or highly conditioned athlete, an intensity of approximately 95% HR_{max} is recommended. Exercising target minute ventilation can be estimated by multiplying the

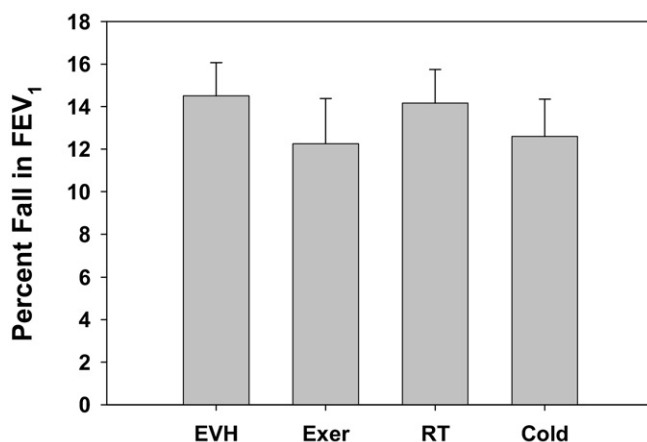


Figure 4. Athletes with mild exercise-induced bronchoconstriction underwent exercise (Exer) and eucapnic voluntary hyperpnea (EVH) challenges breathing either cold (Cold) or room temperature (RT) dry air. The first bar is all EVH (Cold and RT), the second bar is all Exer (Cold and RT), the third bar is all RT (EVH and Exer), and the fourth bar is all cold (EVH and Exer). Similar decreases in FEV₁ were seen for combinations of both challenge modes and both temperature conditions with no significant differences being observed. Adapted from Evans et al.²⁴⁹

baseline FEV₁ by 21 and 30 times for 60% and 85% MVV, respectively.^{94,169,267}

Treadmill Protocol. The treadmill speed and incline are selected according to the tester's subjective assessment of the individual's physical fitness with the ultimate goal of obtaining a total test duration of 8 minutes with the final 4 to 6 minutes at an intensity equal to 80% to 90% of estimated HR_{max}. This intensity should result in a ventilation equal to 40% to 60% of MVV.⁶ A reasonable protocol is to begin the test at 2.5 mph (4 kph) at an incline of 2.5%, adjusting speed and incline to elicit a heart rate of 80% to 90% HR_{max} within 2 minutes and maintain it for the remaining duration of the 8-minute challenge.³⁹ There are nomograms that can be used to estimate oxygen consumption in relation to body weight and treadmill speed both for children²⁶⁸ and adults.²⁶⁹ Patients can terminate the challenge at any time during the test if they feel unable to continue.²⁸ Because water content and minute ventilation are major determinants of the severity of the decrease in FEV₁, it is important to increase the minute ventilation rapidly. Thus, protocols using progressive exercise such as the Bruce Protocol and the Jones Progressive Exercise Protocols²⁷⁰ are inadequate to evaluate EIB because they do not increase the minute ventilation fast enough and may lead to refractoriness without provoking a decrease in FEV₁. Unfortunately, ventilation is the stimulus, but it is not usually measured because of equipment limitations.

Bicycle Ergometer Protocol. The designated workload of the bicycle ergometer challenge is based on the relationship between oxygen consumption and workload, as well as between ventilation and FEV₁.²⁷¹ For example, for a patient with predicted FEV₁ of 3 L, the target workload would be 150 W.

As described by 2 sets of authors,^{6,271} the test begins at 60% of target workload (eg, 90 W) for the first minute, then 70% (105 W) and 90% (135 W) for second and third minutes, and 100% (150 W) for minutes 4 to 8. This equation has some limitations and only acts as a guide for a target workload. For some patients, it may generate a workload that is too high, yet for others it may be too low so adjustments may be needed during exercise.

Inhaled air during both challenges should be dry medical-grade air (<5 mg H₂O+L) administered via a gas cylinder with a reservoir bag (Douglas bag) and 1-way valve apparatus; nose clips should be worn during the test. It is recommended that the challenge is not performed while breathing ambient air (eg, 22°C, 50% RH) because of the increased likelihood of a false-negative test result.²²⁶ Heart rate, arterial oxygen saturation by pulse oximetry, and minute ventilation (if possible) should be measured throughout the test; heart rate and arterial saturation should be monitored after challenge to recovery. Monitoring of oxygen saturation is of primary importance in the differential diagnosis of exercise-induced hypoxemia (EIH). Exercise-induced hypoxemia, a significantly compromised gas exchange during heavy exercise, affects approximately 40% of elite endurance athletes²⁷² and is commonly misdiagnosed as EIB. Tight control of test conditions enhances high test-retest reliability important for monitoring patient treatment on subsequent follow-up.²⁷¹ Either cycle ergometry or treadmill running is an acceptable mode of challenge; however, careful monitoring of cycle load or treadmill running speed and elevation is important to test validity and reproducibility. The most important determinants of the response to exercise are the ventilation achieved and sustained for 6 to 8 minutes and the water content of the inspired air; thus, the ventilation should be considered the measure of the stimulus when dry air is inspired and heart rate as only a secondary means of monitoring the intensity of the stimulus.

Pulmonary function tests should follow ATS standards for FEV₁ and FVC maneuvers.⁶ Two reproducible efforts with FEV₁ within 3% of each other should be obtained before the exercise challenge; the best effort should be used to calculate postexercise decrease in FEV₁. Before conducting the exercise challenge, it is useful to calculate some target values from the preexercise FEV₁; for example, the FEV₁ value that represents (1) a positive test result (ie, a 10% decrease in FEV₁), (2) an alert for severe bronchoconstriction and need for administering a bronchodilator (ie, 50% decrease in FEV₁), and (3) an indication of recovery (ie, the 95% value for recovery of FEV₁). Two reproducible expiratory efforts should be obtained after exercise usually at 1 to 3, 5, 10, 15, 20, and perhaps 30 to 45 minutes after completion of the challenge or at least until the nadir in FEV₁ is reached and recovery has started. A positive test result is best confirmed by a 10% decrease in FEV₁ from baseline at 2 consecutive time points. Full follow-up of FEV₁ to recovery permits the maximal decrease in FEV₁ to be identified. This value is useful for identifying benefit of a medication because a 50% reduction in the decrease after treatment is considered a

significant benefit. Further follow-up of FEV₁ for 60 minutes allows the calculation of area under the FEV₁ time recovery curve (AUC_{0-60 min}) and permits calculation of the time to 95% recovery in FEV₁. The patient should remain at the office or laboratory until the FEV₁ has returned to 95% of baseline value or the clinician feels that it is safe for the patient to leave the clinic.

Eucapnic Voluntary Hyperpnea Challenge

Eucapnic voluntary hyperpnea was developed and standardized at the Walter Reed Hospital in Washington to assess EIB in recruits with a history of asthma.²⁷³ Eucapnic voluntary hyperpnea was later identified as a useful test to identify EIB in elite athletes who were often limited by the lack of an appropriate ergometer that would challenge them sufficiently.^{4,94,274} Eucapnic voluntary hyperpnea has since been recommended by the Independent Panel on Asthma of the IOC-MC as the optimal test to identify EIB for athletes seeking approval to inhale a β_2 -agonist before an event.^{165,169}

The profile of the airway response to EVH shares many similarities with exercise. This includes the stimulus itself, the time course of the airway response and the recovery, the mediators involved, and the inhibitory effects of drugs.^{30,32,97,114} However, EVH does not result in the same cardiovascular response as exercise or the same sympathetic drive, and there are a few subjects who test negative to EVH but positive to exercise.^{93,94}

An EVH challenge requires the subject to breathe dry medical-grade air (containing 4.9%–5% carbon dioxide, 21% oxygen, and balance nitrogen) at an exercising V_E for 6 minutes. Because hyperventilation-induced hypocapnia can cause bronchoconstriction in people with and without asthma,²⁷⁵ eucapnia should be ensured by using EVH air mixture containing 5% carbon dioxide.^{94,274,276} It has been demonstrated that a single fraction of inspired carbon dioxide (4.89%) will provide near normal alveolar carbon dioxide over a wide range of voluntary hyperventilation (30–110 L/min).⁹¹ This may not be applicable to the elite athlete. This was a primary study that demonstrated that the EVH challenge produced similar decreases in FEV₁ to a dry air exercise challenge.

The general procedure for the EVH challenge involves voluntary breathing of the air administered through a 2-way valve connected to a gas cylinder through a reservoir bag for 6 minutes. Prechallenge and postchallenge spirometry are performed. Recommended ventilation rate for EVH is between 21 and 30 times FEV₁ (representing 60% and 85% MVV, respectively). A target value of 30 times FEV₁ is best used only for trained or elite athletes who regularly perform exercise without premedication. The lower value of 21 times FEV₁ is used in known asthmatic patients or those with a history of EIB not taking regular treatment. Patients may have suboptimal challenges when lower levels of ventilation are used as the target.⁹⁷ The ventilation reached and sustained are important determinants of the response to exercise.¹³ However, prolonged periods of hyperpnea can result in a significant refractoriness to the stimulus; it has been demon-

strated²⁷⁷ that 2 consecutive EVH challenges within 93 minutes resulted in respective decreases in FEV₁ of 27.4 ± 9.8 and 16.1 ± 5.9, whereas three 2-minute interrupted EVH challenges resulted in an 18.9 ± 10.6 decrease in FEV₁ after the last challenge.²⁷⁷

It is important to measure minute ventilation to ensure adequate and consistent ventilation throughout the 6-minute test. This can be accomplished by using either a rotameter between the gas cylinder and patient to maintain a constant predetermined flow and/or by measuring continuous V_E on exhalation. One variation of EVH is to chill inspired air, which has been hypothesized to lead to a greater change in airway osmolality or vascular response. However, others^{248,249} found no difference in postEVH decreases in FEV₁ when the room temperature or cold air was inhaled (Figure 4). Because the cold temperature inhaled air did not have an additive effect on the bronchoconstrictive response, the same authors^{248,249} confirmed earlier work⁷⁰ demonstrating that water content, and not temperature of inhaled air, is the essential test condition from bronchoconstrictive response. It must be reiterated that EVH should be performed with caution in individuals who have a baseline FEV₁ less than 80% of predicted and should not be performed in those with baseline FEV₁ less than 70% of predicted.

Hypertonic Saline Challenge

The hypertonic saline challenge is economical and an easily administered indirect challenge that increases ASL osmolality and triggers sensitized cells to release inflammatory mediators.^{278–280} There are many similarities among challenges with hypertonic saline, EVH, and exercise.^{281,282}

In addition, hypertonic saline allows for concurrent collection of sputum samples for mediator and cellular analysis.^{283–285} The hypertonic saline (4.5%) aerosol is generated using a high-output ultrasonic nebulizer and the exposure times are progressively increased (ie, 30 seconds, 60 seconds, 2 minutes, 4 minutes, and 8 minutes, which together totals 15.5 minutes) so that the total test time is usually less than 20 minutes; moreover, decreases in FEV₁ are usually not severe.^{283–285} Initial exposure to the 4.5% hypertonic saline lasts for 30 seconds²⁸⁴; this interval is doubled if FEV₁ decreases less than 10% between exposures. Forced expiratory volume in 1 second is measured 60 seconds after exposure. If FEV₁ decrease is 10% or more, the exposure time is repeated; the test is terminated if decrease is 15% or more in FEV₁ or when a total dose of 23 g has been administered in 15.5 minutes.²⁸⁴ Inhaled saline produces a dose-dependent response, allowing for classification of the severity of BHR in populations of those not using ICS. An epidemiologic study²⁷⁹ found that children positive to hypertonic saline were approximately 4.26 times more likely to have EIB. A later study by the same investigators corroborated a positive response to saline and diagnosis of asthma.²⁸⁶

Thus, hypertonic saline has been used to identify the potential for BHR in patients with a history of asthma who are currently symptomatic²⁸⁷ and asymptomatic.²⁸⁸ This is partic-

ularly useful in those who wish to participate in scuba diving because 18% of intending scuba divers have been shown to have asthma and hypertonic saline is useful to assess the effect of treatment.²⁸⁹

Inhaled Powder Mannitol Challenge

Mannitol is a stable sugar alcohol found in most vegetables that is a hyperosmolar agent that has been developed into a bronchial provocation challenge.²⁹⁰ The mannitol test is approved in Europe, Asia, Australia, and the United States for the assessment of bronchial hyperresponsiveness.³ Like exercise, EVH, and hypertonic saline, inhaled powder mannitol is an indirect challenge that produces airway smooth muscle contraction by creating a hyperosmolar environment that leads to release of mediators from inflammatory cells of the airways.⁸¹ Mannitol challenge is associated with mast cell activation and mediator release; after mannitol challenge, the levels of urinary 9α,11βPGF₂, and LTE₄ were increased similarly in both asthmatic and nonasthmatic subjects.⁸¹ The results of this study suggest that the difference between asthmatic and nonasthmatic subjects is not in the release of mediators per se but more in the responsiveness of airway smooth muscle to bronchoconstrictive mediators. As an indirect challenge, inhaled powder mannitol reflects the neural and cellular aspects of the airway responsiveness rather than just smooth muscle hyperresponsiveness independent of mediator release.^{283,290}

Mannitol is administered from a dry powder inhaler device in progressive doubling doses of 5, 10, 20, 40, 80, 160, 160, and 160 mg, with a maximal total dose of 635 mg, depending on the airway response.^{39,290} One minute after each dose, FEV₁ is measured in duplicate by the FEV₁ maneuver. Full FVC maneuvers should not be obtained to avoid tiring patients who are performing any of these tests and to avoid taking too long to perform spirometry. The baseline FEV₁ obtained after the initial capsule containing no mannitol is used to calculate the target for the 15% decrease in FEV₁ (0.85 × 0-mg dose FEV₁) after subsequent doses. The challenge is discontinued when there is a 15% or greater decrease in FEV₁ from baseline or a between dose decrease of 10% or greater in FEV₁ or a cumulative dose of 635 mg is administered.³⁹ The data are expressed as the dose required to provoke a 15% decrease in FEV₁ (PD₁₅), which is obtained by plotting the change in FEV₁ against the log cumulative dose of mannitol delivered. The following equation⁶ is used to calculate the PD₁₅:

$$PD_{15} = \text{antilog} \left[\log D_1 + \frac{(\log D_2 - \log D_1)(15 - F_1)}{F_2 - F_1} \right]$$

where D₁ is the mannitol dose preceding D₂, D₂ is the final mannitol dose resulting in a 15% or greater decrease in FEV₁, F₁ is the percentage decrease in FEV₁ after D₁, and F₂ is the percentage decrease in FEV₁ after D₂.

The inhaled powder mannitol challenge has a number of inherent advantages over other indirect and direct challenges.

It is practical for office administration because of the ease of use, short time to perform the challenge, and no requirement for specialized and costly equipment, such as a treadmill or cycle ergometer. The only requirements are spirometer, nose clips, calculator, and mannitol kit.

There is a significant relationship between sensitivity to mannitol and reactivity to exercise in known asthmatic patients not being treated with inhaled steroids.^{247,291} Mannitol as a challenge test has also been used to identify EIB in elite athletes.²⁹² There is a lower sensitivity for mannitol to identify EIB in those with symptoms but no definite diagnosis of asthma.³⁹ Mannitol has been used successfully to identify BHR in individuals with exercise-induced wheeze.²⁹³

Of potential importance is the capability to use the mannitol challenge to document treatment effectiveness.²⁹⁴ Because BHR is dependent on the presence of airway inflammation, a decrease in sensitivity to mannitol may be used as an index of reduction in airway inflammation. This can be documented by change in PD₁₅ and RDR (calculated by dividing the final percentage decrease in FEV₁ by the cumulative dose of mannitol producing that decrease). There was a significant reduction in sensitivity (PD₁₅) and airway reactivity (RDR) in asthmatic patients after treatment with ICS; the PD₁₅ increased from a pretreatment value of 78 mg to 289 mg after treatment and a 4.2-fold improvement in RDR.²⁹⁴

Summary Statement 19. Self-reported symptom-based diagnosis of EIB in the elite athlete lacks sensitivity and specificity and establishes the necessity for standardized, objective challenges using spirometry. B

The elite athlete with EIB typically presents with symptoms of cough, wheeze, chest tightness, dyspnea, and excess mucus.¹ However, the same symptoms may be present in elite athletes without reversible airway obstruction, and the athlete may not be adept at reporting those symptoms.^{1,2} Investigators¹ demonstrated that approximately half of elite athletes who reported EIB symptoms had normal spirometry after a challenge test and approximately half who reported no symptoms of EIB tested positive. Others² demonstrated essentially the same results in Division I college athletes. These data demonstrate that the diagnosis of EIB in the elite athlete must be confirmed with a positive challenge test result.

Summary Statement 20. The indirect challenge (eg, exercise or surrogate such as EVH) is preferred over a direct challenge (eg, methacholine) for assessing EIB in the elite athlete. D

Challenge tests for asthma and EIB are either direct or indirect. The direct challenge involves methacholine or histamine inhalation and acts on smooth muscle receptors in the muscle. This challenge is independent of airway inflammation. The direct challenge involves inhalation of methacholine or histamine in progressively increasing amounts until a decrease in FEV₁ of 20% or greater from baseline occurs. If a PC₂₀ of methacholine is less than or equal to 4 mg/mL when the patient is not taking ICS or 4 to 16 mg/mL when the patient has been taking ICS for 1 month or longer, the test result is considered positive according to the IOC-MC.²⁹⁵

The methacholine challenge is not recommended for the athlete with EIB without the presence of known asthma. Methacholine has been shown to have a low sensitivity and high specificity for EIB without asthma.²⁰⁶ A study examining 50 elite summer athletes⁴ found that 42 reported 1 or more symptoms, 9 had positive methacholine challenge results, and 25 had positive EVH challenge results with a mean decrease in FEV₁ of 25.4% ± 15%. Methacholine had a negative predictive value of 61% and a sensitivity of 36% for identifying athletes responsive to EVH.

Summary Statement 21. Eucapnic voluntary hyperpnea is the preferred surrogate challenge for the elite athlete participating in competitive sports. D

Indirect challenges are preferred for assessing EIB in elite athletes. The most frequently used indirect challenges for elite athletes include sports-specific field exercise, laboratory exercise, EVH, and inhaled powder mannitol. Eucapnic voluntary hyperpnea is considered the most useful and predictive of these indirect challenges,^{2,4,94,236,274,296} and because of the high sensitivity to identifying athletes with EIB, it is the challenge recommended by the IOC-MC Independent Asthma Panel.²⁹⁵ Sports-specific field-based exercise challenges at race-pace intensity are successfully used,¹⁶⁴ but the lack of environmental control affects results and test-retest consistency.³ Using competitive swimmers as subjects, an investigation²³⁶ demonstrated that a sports-specific challenge was not effective at diagnosing EIB in swimmers. Eucapnic voluntary hyperpnea was more likely to produce a positive response than a swim challenge or a laboratory cycle challenge at more than 85% estimated HR_{max}, temperature of 21°C, and RH of 50%; 1 of 33 subjects had a positive field swim challenge result, 18 had a positive EVH challenge result, and 4 had a positive laboratory cycle challenge result.

Ventilation for EVH should be at more than 85% MVV when testing the elite athlete, and inhaled air contains less than 5 mg H₂O/L of air. The EVH test uses compressed air with 4.95% to 5% carbon dioxide to ensure eucapnia, as described earlier, and the test should be of a 6-minute duration and follow the procedure previously described. The EVH test should be performed with caution, especially in patients with FEV₁ that is below 80% of predicted, and should not be performed on patients in whom FEV₁ is less than 70% of predicted.

Summary Statement 22. The intensity of the exercise challenge for the elite athlete should be 95% or greater than actual or estimated HR_{max}, and dry medical-grade air should be used in performing the challenge. D

Laboratory exercise challenge should follow the same procedures as described earlier; however, exercise intensity should be at or above 95% of HR_{max} for the last 4 minutes of the 8-minute test. This heart rate–based intensity will ensure adequate ventilation for a reliable test. Cycle ergometry tests should be reserved for athletes who are cyclists, skaters, or alpine skiers, whereas treadmill run tests should be performed on Nordic skiers and runners. Triathletes can be tested by cycle ergometry or treadmill running. Although cutoff criteria

have been shown to be statistically justified at values less than a 10% decrease in FEV₁,^{1,231} the IOC-MC dictates a 10% or greater decrease in FEV₁ 3 to 30 minutes after exercise as a positive challenge result.

Summary Statement 23. Hyperosmolar aerosols may also be used as surrogates to exercise. C

Inhaled mannitol has been successfully used as a challenge surrogate for EIB in elite athletes.²⁹² A decrease in FEV₁ of 15% or more after inhaling 635 mg of dry powdered mannitol is considered positive for EIB. The mannitol response is reported as PD₁₅ or a between dose fall of 10%,³⁹ and these challenges are supported by the IOC-MC.^{35,169}

In conclusion, a diagnosis of EIB must include objective lung function measures using a standardized indirect challenge. Although the standardized dry air exercise challenge and EVH are effective in diagnosing EIB, equipment is expensive and may not be practical in many clinical settings. Nevertheless, the hypertonic saline challenge and inhaled powdered mannitol require less equipment and space and can be easily performed in the office environment.

Differential Diagnosis

Summary Statement 24. Exercise-induced laryngeal dysfunction, primarily VCD and other glottic abnormalities, may be elicited by exercise and mimic EIB. Inspiratory stridor is a differentiating hallmark sign with EILD and not with EIB alone. Flattening of the inspiratory curve on spirometric maneuver may be seen concomitant with symptoms. Exercise-induced laryngeal dysfunction may occur alone or with EIB. Failure to respond to asthma management is a key historical feature suggesting EILD. C

Since the initial description of VCD as a functional disorder that mimicked attacks of asthma,²⁹⁷ VCD and glottic structural abnormalities elicited seen with exercise have been increasingly recognized. These functional and structural disorders can be grouped as EILD, including (1) paradoxical VCD, (2) exercise-induced laryngeal prolapse,²⁹⁸ (3) exercise-induced laryngomalacia,²⁹⁹ and (4) variants, including arytenoid collapse while the vocal cords move normally.³⁰⁰ Exercise-induced laryngeal dysfunction occurs in all age groups, especially among young adult female elite athletes.²⁵⁰

Bronchial provocation challenge with methacholine, exercise, and EVH may be negative in patients with EILD who do not otherwise have BHR. The onset of breathing difficulties occurs and peaks during exercise with EILD, rather than peaking after exercise with EIB. Medications used to treat asthma such as β_2 -agonists are ineffective to prevent or reverse EILD. Exercise-induced laryngeal dysfunction can be suspected by bronchial provocation challenges with isocapnic hyperventilation of frigid air, methacholine, and/or exercise demonstrating variable extrathoracic airway obstruction.³⁰¹

Inspiratory stridor with throat tightness during maximal exercise resolves within approximately 5 minutes of discontinuation of exercise in patients with EILD. Inspiratory stridor with EILD contrasts with EIB, in which case dyspnea generally occurs after exercise, peaks 5 to 20 minutes after

stopping, and involves expiration rather than inspiration.⁷ There may be variations in the timing of the manifestation of EILD symptoms, depending on such factors as the duration and intensity of the exercise.

In VCD, direct observation of the vocal cord adduction by laryngoscopy and flattening or truncation of the inspiratory portion of the spirometric flow-volume loop are the hallmarks for diagnosis. These findings may only be seen during symptomatic periods. Methacholine challenge can be used to elicit VCD.^{302,303} Additional evidence of VCD may be suggested by examining a video of the patient recorded while exercising in the natural setting at the time that inspiratory stridor is heard.³⁰⁴ Diagnosis can be made directly by using continuous laryngoscopy during exercise challenge.³⁰⁵ Spirometry and laryngoscopy with sound recording can be performed during exercise, detecting minor and major aryepiglottic and vocal cord abnormalities.

Exercise-induced laryngeal prolapse has been seen in otherwise healthy athletes and can present with subtotal occlusion of the larynx. This condition may result from mucosal edema from the aryepiglottic folds being drawn into the endolarynx (laryngochalasia).²⁹⁸ Laryngoscopic evaluation at rest may be normal, and various laryngeal abnormalities may only be elicited with exercise challenge.³⁰⁶

Laryngomalacia is associated with diminished laryngeal tone, resulting in supraglottic collapse, and is usually a congenital condition.³⁰⁷ Laryngomalacia is the most common cause of inspiratory stridor in infants³⁰⁸ but may not manifest until later childhood with participation in competitive sports.^{299,307,309–311} Although the typical anatomical features of congenital laryngomalacia (shortened aryepiglottic folds or retroflexed epiglottis) may not be seen, other presentations, such as profound arytenoid redundancy and prolapse, can be seen during nasolaryngeal endoscopy. As in infants with laryngomalacia, supraglottoplasty can improve late-onset disease.^{307,308} Laryngomalacia can also be seen in adults.³¹²

Concurrent laryngeal abnormalities may be seen in patients with VCD. Laryngoscopy may identify findings suggestive of GERD, chronic laryngitis, laryngomalacia, vocal cord motion impairment, nodules, and subglottic stenosis, especially in patients in whom exercise induces symptoms.³¹³ Exercise-induced laryngeal dysfunction may coexist with EIB. Inspiratory stridor is the signature clinical feature suggesting EILD rather than EIB.

Gastroesophageal reflux anatomical findings may be seen on laryngoscopy in children and adults with EILD, but whether they are causative or concomitant is difficult to establish. Empiric pharmacologic treatment of GERD in juveniles having VCD has been recommended because posterior laryngeal changes associated with GERD are common in juveniles with VCD.³¹⁴

Although laryngopharyngeal reflux may be a contributing factor in many EILD cases, there is little objective evidence supporting this. The sensitivity and specificity of the laryngoscopic examination to diagnose laryngopharyngeal reflux is also controversial.³¹⁵ Although there may be a clinical

suspicion of laryngopharyngeal reflux, there is an absence of an objective “gold standard” to establish this diagnosis.

Summary Statement 25. Exercise-induced dyspnea and hyperventilation can masquerade as asthma, especially in children and adolescents. C

Chest discomfort perceived as dyspnea during vigorous exercise may be associated with hypocapnia from hyperventilation without bronchoconstriction, especially in children and young adolescents previously diagnosed as having and having been treated for EIB.^{19,20,316–319}

It has been demonstrated that exercise-induced lactic acidosis is causally involved in the hyperventilation. However, lactic acidosis does not represent the only additional stimulus of ventilation during intense exercise. Sensory input from exercising muscles such as muscle afferents may also trigger hyperventilation.^{320,321}

Idiopathic hyperventilation is a poorly understood condition in which patients have sustained hyperventilation, hypocapnia, and dyspneic drive.³²²

Summary Statement 26. Shortness of breath with exercise may be associated with underlying conditions due to obstructive lung disease, such as COPD, or restrictive lung physiology, such as obesity, skeletal defects (eg, pectus excavatum), diaphragmatic paralysis, and interstitial fibrosis. B

Dyspnea with exertion in some obese patients may not be a manifestation of EIB.³²³ Idiopathic pectus excavatum may be associated with exercise symptoms, including chest pain, dyspnea, or impaired endurance. Even in the absence of clinical symptoms, restrictive lung defects and lower airway obstruction are common.^{324,325}

Scoliosis has been associated with decreased exercise tolerance. Patients who have mild scoliosis may be asymptomatic at rest but with exercise may have decreased tidal volume, as well as hypercapnia and hypoxia.³²⁶

Although diaphragmatic paralysis has a predictable effect on lung function, the symptoms depend on the preexisting heart-lung diseases and may mimic various cardiorespiratory processes, including EIB.³²⁷

Patients with interstitial lung disease frequently have dyspnea with exercise. These patients' exercise limitations appear to be related to arterial hypoxemia and not respiratory mechanics. Their dyspnea is often fixed and reproducible.^{328,329}

Summary Statement 27. Shortness of breath accompanied by pruritus and urticaria, with varying other systemic symptoms, suggests EIA_na rather than EIB. C

Exercise-induced anaphylaxis is characterized by the exertion-related onset of cutaneous pruritus and warmth, generalized urticaria, and appearance of such additional manifestations as shortness of breath, upper respiratory tract distress, vascular collapse, and gastrointestinal tract symptoms. This must be differentiated from asthma, cholinergic urticaria, angioedema, and cardiac arrhythmias, which are recognized as exertion-related phenomena in predisposed patients but are distinct from EIA_na.^{330,331} There is variability in the reproducibility of EIA_na symptoms given similar testing conditions.

Episodes of food-dependent exercise-induced anaphylaxis (FDEIA_na) may or may not be dependent on ingestion of identifiable foods.^{332,333} The cumulative effect of exercise and food ingestion may trigger the mediator release and anaphylaxis, whereas each of these triggers independently does not. Foods reported as predisposing factors range from shellfish, eaten 4 to 24 hours before EIA_na, to seemingly benign foods, such as celery, eaten before or after exercise.^{332,333} Skin testing with foods may be helpful in eliciting the trigger when history taking does not.³³⁴ Serum tryptase measurements may help in confirming the diagnosis of EIA_na.³³⁵ FDEIA_na occurs in both children and adults.^{330–336}

Wheat gliadin has been identified as the cause of FDEIA_na due to wheat.³³⁷ It has been further determined that cross-linking between tissue transglutaminase and ω -5 gliadin-derived peptides increases IgE binding. The tissue transglutaminase becomes activated in the patients' intestinal mucosa, and large allergen complexes become capable of eliciting anaphylaxis.³³⁸ Exercise and aspirin have been shown to increase the levels of circulating gliadin peptides in patients with wheat FDEIA_na, suggesting facilitated allergen absorption from the gastrointestinal tract.³³⁹

Although skin testing to the specific foods, commercial or fresh food extracts, or crude gliadin is most used, measurement of IgE levels specific to epitope peptides of ω -5 gliadin or recombinant ω -5 gliadin may be useful as an in vitro diagnostic method.^{339,340}

Food-dependent exercise-induced anaphylaxis can have a delayed onset for an unpredictable number of hours. It has therefore been suggested that, in such patients, exercise should be avoided 4 to 6 hours after specific food ingestion. In patients with wheat gliadin-associated EIA_na, a gluten-free diet is recommended.^{337,341}

Susceptible patients may be advised to take an antihistamine before exercise and should carry self-injectable epinephrine, which is the primary treatment for anaphylaxis.³⁴¹

Summary Statement 28. In the absence of objective evidence of EIB, breathlessness with exercise, with or without chest pain, may be caused by cardiovascular, pulmonary, or gastroenterologic mechanisms other than asthma. Appropriate cardiopulmonary testing and/or referral to a cardiologist, pulmonologist, or gastroenterologist may be necessary. B

Although the incidence of cardiac-related dyspnea with exercise in young, healthy patients is minimal, it remains an important differential in EIB. Primary pulmonary hypertension (PPH) may occur in both adults and children. Patients with PPH may demonstrate peripheral airway obstruction, poor oxygenation, and early physiologic aerobic limits restricting exertion and, in children, documentation of significant reversibility of lower airway obstruction.^{342–344} A case report documents how PPH can masquerade as asthma. Two adult nonsmokers presenting with wheezing, chronic cough, and irreversible obstructive lung disease were diagnosed as having adult-onset severe refractory asthma but actually had dilation of the central pulmonary arteries, compressing the mainstem bronchi.³⁴⁵

Congestive heart failure may present with dyspnea on exertion. Hyperpnea with exercise may occur without lung function impairment. Ventilation-perfusion mismatch in exercise may be enhanced with increased treatment of heart failure.³⁴⁶

Hypertrophic cardiomyopathy is well known to cause sudden death in young athletes, with an annual 1% mortality rate.³⁴⁷ Patients may have dyspnea and chest pain that improve with β -blockers.³⁴⁸

Cardiac dysrhythmias may also cause dyspnea with exercise. Supraventricular tachycardia may cause EIB in children.¹⁹ Young adults with complete heart block may have shortness of breath, dyspnea on exertion, syncope, dizziness, or fatigue.³⁴⁹

Vascular rings of the aorta are rare but may present as asthma. Spirometry in these patients reveals a decreased peak expiratory flow and truncation of the expiratory flow volume loop with normal FVC, FEV₁, and FEV₁/FVC. Chest radiographs are significant for a right aortic arch.³⁵⁰

Pulmonary arteriovenous malformations and disorders with right to left shunts may cause exercise-induced dyspnea due to hypoxemia, without associated bronchoconstriction. Hereditary hemorrhagic telangiectasia, atrial septal defects, ventricular septal defects, and Osler-Rendu-Weber syndrome are among the primary causes. Cardiopulmonary exercise testing, as differentiated from pulmonary testing for EIB, is an appropriate noninvasive tool to begin and guide the evaluation of these patients presenting as undiagnosed dyspnea. The evaluation may require procedures such as cardiac catheterization to further delineate the right to left shunt.^{351,352}

Exertional dyspnea in symptomatic patients with COPD may be due to the combined deleterious effects of higher ventilatory demand and abnormal ventilatory dynamics but not temporally attributable to bronchoconstriction.³⁵³ Patients with COPD may have evidence of small airway dysfunction with increased ventilatory requirements during exercise likely on a basis of greater ventilation and perfusion abnormalities. These abnormalities also involve changes in end-expiratory lung volume and breathing patterns that are more shallow and rapid than in a comparatively healthy cohort.

Summary Statement 29. Exercise-induced dyspnea is seen as a physiologic limitation in otherwise healthy active individuals without bronchospasm. C

Perhaps the most common reason for exercise-induced dyspnea in children is physiologic (poorly conditioned) limitation without bronchospasm or underlying disease.¹⁹ Limits in exercise performance and respiratory system oxygen transport may occur in highly fit adults.³⁵⁴ This may be due to flow limitation in intrathoracic airways because of narrowed, hyperactive airways or secondary to excessive ventilatory demands superimposed on a normal maximum flow-volume envelope. In addition, exercise-induced arterial hypoxemia occurs as a result of an excessively widened alveolar-arterial oxygen pressure difference. This inefficient gas exchange may be attributable in part to small intracardiac or intrapulmonary shunts of deoxygenated mixed venous blood during

exercise. Finally, fatigue of the respiratory muscles resulting from sustained, high-intensity exercise and the resultant vasoconstrictor effects on lung muscle vasculature will also compromise oxygen transport and performance. Exercise in the hypoxic environment of even moderately high altitudes will greatly exacerbate the negative influences of these respiratory system limitations to exercise performance, especially in highly fit individuals.

Summary Statement 30. The association between EIB and GERD is controversial, and probably there is no relationship. A

Although there are reports of exertional gastroesophageal reflux in healthy individuals, most studies have demonstrated no significant correlations between GERD and EIB.³⁵⁵⁻³⁵⁷ Although acid reflux may be common in EIB patients, many patients with exercise-related respiratory symptoms may be misdiagnosed as having asthma when they truly have exercise-onset GERD.³⁵⁸ Some controversies exist in treatment of GERD and EIB. Symptoms of acid reflux related to running were relieved by proton pump inhibitor, but the respiratory symptoms of EIB were not relieved by proton pump inhibitors.³⁵⁹ In contrast, other investigators have reported improvements in exercise-related breathing symptoms when patients were treated with proton pump inhibitors.³⁵⁸

Summary Statement 31. Psychological factors need to be considered in the differential diagnosis of EIB. D

Psychological factors may obfuscate the diagnosis in patients with apparent exercise intolerance. Scenarios such as individuals, particularly young women, complaining of shortness of breath while running without having stridor, wheezing, or relief with trial bronchodilators are not uncommon but may be vexing to the patients, their parents, and their physicians. Although VCD and EIH may have functional triggers, differentiating EIB requires subjective and objective assessment.³⁶⁰ Mental stress may be one trigger factor where hyperventilation is seen in patients with asthma-like symptoms with negative asthma test results.³⁶¹ If objective testing does not reveal any bronchoconstriction or other physiologic explanations, then psychological etiology should be considered and addressed with the patient, which may involve a recommendation for psychological consultation.

Summary Statement 32. Dyspnea on exertion, which is prevalent in otherwise healthy, obese individuals, is not associated with EIB. C

Dyspnea on exertion is present in obese patients. This dyspnea has been strongly associated with an increased oxygen cost of breathing, without bronchoconstriction, in otherwise healthy obese women.³⁶² Exercise capacity has been variously reported as unchanged in obese females to being reduced at or near maximal effort.³⁶³

Summary Statement 33. Mitochondrial enzyme deficiency with myopathy is a rare cause of exercise limitation. D

Impaired oxidative phosphorylation in working muscle disrupts the normal regulation of cardiac output and ventilation relative to muscle metabolic rate in exercise.³⁶⁴ Deficiencies of mitochondrial enzymes cause a number of severe neurologic syndromes in pediatric patients. Isolated myopa-

thies secondary to enzymatic deficiency have been recognized in adults and may be more prevalent than reported.^{365,366}

Therapy

Introduction

Summary Statement 34. Frequent EIB in asthmatic patients suggests inadequate asthma control and requires patient re-evaluation to determine the need for additional therapy. D

Exercise-induced bronchoconstriction is a reflection of BHR and, in asthmatic children and adults, ordinarily is due to underlying inflammation. Exercise-induced bronchoconstriction in these individuals, most of whom are not elite athletes, may represent inadequacy of overall asthma control.^{33,207}

Summary Statement 35. Failure of appropriate pharmacotherapeutic agents to prevent EIB indicates the need to re-evaluate the diagnosis. D

The goal of therapy for EIB is to prevent symptoms induced by exercise, to enhance overall control of asthma, and to ameliorate symptoms rapidly when they occur. Pharmacotherapeutic agents that are effective in controlling chronic asthma generally have bronchoprotective activity for EIB. If asthma is otherwise well controlled, bronchoprotective therapy is administered only as needed. This therapy may be delivered by inhalation or by oral administration minutes to hours before exercise, respectively. Nonpharmacologic therapies can also be helpful in preventing EIB when used alone or in combination with pharmacotherapy; these are described in the “Nonpharmacologic Therapy” Section.

Summary Statement 36. Several pharmacotherapeutic agents are effective when given for the prevention or attenuation of EIB. They differ in their mechanisms of action and overall effectiveness. In addition, there is both inpatient and outpatient variability in responsiveness. A

Pharmacotherapeutic agents act to prevent or attenuate EIB by various mechanisms with different degrees of effectiveness. None of the available therapies completely eliminates EIB. Pharmacotherapy shifts the dose-response relationship to a more favorable position after exercise.^{367,368} The efficacy of a given agent in protecting against EIB can vary at different times and among different individuals.

Summary Statement 37. Medications may differ in effectiveness over time because of variability of asthma, environmental conditions, intensity of the exercise stimulus, and tachyphylaxis. A

The variability of effectiveness within an individual may be due to changes in airway responsiveness over time, environmental conditions, and intensity of the exercise stimulus.³⁶⁹ The variability among individuals may result from differences in baseline airway responsiveness and susceptibility to tachyphylaxis and perhaps to genetic differences.⁷ Pharmacotherapeutic studies supported by pharmaceutical sponsors generally have used parallel groups or crossover designs to compare active drugs with placebo or to compare 2 (or more) active drugs.³⁶⁹ The primary end point is most

commonly the maximum percentage decrease in FEV₁, especially for studies submitted to the US Food and Drug Administration (FDA)³⁶⁹ in support of a bronchoprotective end point. Peak expiratory flow has also been used as an end point in some studies but not as a primary end point, and it is used less commonly than FEV₁.

In addition to the maximum absolute decrease in FEV₁, expressed as a percentage of baseline, the results may also be expressed as the AUC^{0–60 min} or until return to baseline and the time to recover to 95% baseline FEV₁ before and after therapy.³⁷⁰ Baseline lung function may also be compared before and after therapy if a bronchodilator response is also being investigated.^{6,24,29,112,369,371–374} Some studies have examined the percentage of individuals protected from EIB after therapy (responder analysis).

The maximum decrease in FEV₁ required to produce a positive test result varies with the situation in which the test is performed. In a clinical setting, the decrease in FEV₁ from baseline required to diagnose EIB is usually 10%⁶ or perhaps 13%³⁷ or 15%.⁴⁰ As an inclusion criterion in a pharmaceutical trial, a 20% decrease in FEV₁ is usually required to define a positive challenge result (as described in FDA guidance).³⁶⁹ In a clinical setting, it is desirable to produce as complete protection as possible so that there is no decrease in FEV₁ after exercise with treatment. In a pharmaceutical trial, protection may be defined as less than a 10% decrease in FEV₁ after exercise or 50% protection compared with placebo³⁷⁵ for patients who are required to have a 20% decrease at the screening visit. When other end points are used (eg, AUC or time to recovery), the percentage of protection must also be adjusted for the situation in which the test is performed. Protection has been defined in some studies based only on statistically significant differences between responses after pretreatment with active drug compared with placebo. Attempts to define protection that have clinical relevancy have resulted in concepts such as “complete protection” and “clinical protection.”^{21,24} Complete protection may be suggested by predefined decreases of FEV₁ percentage within an accepted reference range (eg, <10% for FEV₁). Clinical protection has been defined as a 50% inhibition of the placebo response to exercise by drug pretreatment.^{374,375} The 10% decrease is based on the mean plus 2 standard deviations of the decrease in healthy individuals,³⁸ and the 50% protection is based on the coefficient of variation for repeated tests.³⁷⁵

Numerous studies have been assessed in developing the evidence-based recommendations for therapy in this document, but there is little information on the consistency, presence, or absence of drug effect from recurrent testing in individuals. Also, most published evidence is from EIB studies in patients with clinically diagnosed (and usually atopic) asthma, and recommendations made herein are based on these data.^{2,7,204} Information suggesting possible differences in pathogenesis of EIB and response to pharmacotherapies in apparently nonasthmatic elite athletes is summarized elsewhere in separate sections (“Pathophysiology of Exercise-Induced Bronchoconstriction” and “Competitive and Elite Athletes”). It is recommended that this

section on therapy be used largely as a starting point on which to base a trial of best therapy for a patient with EIB tempered by the responsiveness and needs of that individual patient over time. Failure to demonstrate inhibition or significant attenuation of apparent EIB by effective bronchoprotective agents should indicate the need for reevaluation of the diagnosis.

β_2 -Adrenergic Receptor Agonists

Summary Statement 38. Inhaled β_2 -adrenergic receptor agonists are the most effective group of agents for short-term protection against EIB and for accelerating recovery of FEV₁ to baseline when given after a decrease in FEV₁ after exercise. A

β_2 -Adrenergic receptor agonists are the single most effective therapeutic group of agents for acute prevention of intermittent EIB.^{29,271,372–380} They attenuate or protect against EIB in most patients.^{13,29,271,376,378,381} Their effectiveness may relate in large part to their action as functional agonists acting directly on BSM receptors and probably by inhibiting mast cell mediator release.^{123,382,383} These agents also are the most effective group of agents for enhancing recovery of FEV₁ to baseline values when given after a decrease after exercise.^{371,372}

Summary Statement 39. When given as a single dose or on an intermittent basis, SABAs and LABAs may protect against or attenuate EIB; SABAs are usually effective for 2 to 4 hours and LABAs for up to 12 hours. A

Early investigations of β_2 -adrenergic drugs developed for asthma showed that these agents were highly effective in protecting against EIB when inhaled 5 to 20 minutes before exercise.^{373,376–378} Protection was found to last from 2 to 4 hours after inhalation, with most studies showing a duration at the lower end of this interval.^{384–386} There appear to be no substantial differences among SABAs currently in use.^{386–388} Mast cell stabilizers, described below, have been used as add-on therapy to supplement SABAs in increasing the degree of bronchoprotection.^{29,380}

Two LABAs are currently in use and differ in their actions, mainly in their onsets of effect. Formoterol has a more rapid onset of bronchodilator and bronchoprotective action; in contrast, salmeterol requires 15 to 30 minutes.^{389,390}

Summary Statement 40. Daily use of β_2 -adrenergic agents alone or in combination with ICS usually will lead to tolerance manifested as a reduction in duration and/or magnitude of protection against EIB and a prolongation of recovery in response to SABA after exercise. Therefore, monotherapy with adrenergic agents is generally recommended for use only on an intermittent basis for prevention of EIB. A

Prolonged duration of a bronchoprotective effect, for as long as 12 hours, has been shown for these drugs after the first dose in β_2 -agonist-naïve patients.^{385,391–396} However, many patients are not protected for this entire dosing interval, and the optimal dosing interval for bronchoprotection for EIB may be closer to 6 hours on average.^{385,392,393,395} Prolonged protection with intermittent use of LABAs is sustained,^{392,395–398} but daily maintenance use of LABAs (and SABAs) often results in

some loss of bronchoprotection (“tolerance”) with cross-tolerance to other β_2 -agonists.^{25–27,393,399–402} Moreover, daily use of LABAs and SABAs may actually increase the severity of EIB.^{401,402} Of additional concern, the degree of tolerance may increase with increasing bronchoconstriction, potentially putting patients in severe asthma attacks at risk of less bronchodilator responsiveness at the time of greatest need.⁴⁰³ Therefore, only intermittent use of adrenergic agonists is recommended for bronchoprotection. Although some individuals may have a greater propensity than others to develop tolerance, only a small number of patients are required to demonstrate tolerance,^{27,401–406} suggesting that tolerance occurs in most patients. Tolerance occurs even when patients are also receiving ICS.^{26,28}

The onset of tolerance can be rapid. By extrapolation of the effects on methacholine-induced bronchoconstriction²⁷ and other challenges such as AMP⁴⁰⁷ it may occur within 12 to 24 hours after a first dose.^{27,407–409} The degree appears to increase with constant β_2 -agonist use before it reaches a plateau.²⁷ By similar extrapolation, sensitivity to β_2 -agonists may recover within 72 hours after the last dose of β_2 -agonist use.²⁷

Summary Statement 41. Regular (ie, daily) use of β_2 -agonists for EIB leads to relative loss of efficacy of the agent. A

Tolerance may not develop when β -agonist use is limited to an interval of 48 to 72 hours.⁴¹⁰ However, a longer period for recovery may be required for other stimuli such as allergen challenges.⁴¹¹ Tolerance is manifested most strikingly by a decrease in effectiveness of SABA⁴¹² and by a shortening of duration of LABA effect,^{26,28,391,393,396} in 1 study to less than 3 hours,⁴¹³ and by prolongation of recovery from bronchoconstriction.^{27,401} The presence of tolerance is often missed clinically because a patient is rarely challenged at the point of care; consequently, the shorter duration of protection and the prolonged recovery time are not revealed. Importantly, prescribing additional doses of adrenergic aerosol immediately before exercise may unintentionally contribute to further generation of tolerance.

The mechanism(s) by which long-term (daily) use of β_2 -agonists lead to tolerance is unclear. A number of observations have led to suggestions for possible mechanisms involved in the development of “tolerance.” Long-term exposure of β -receptors to β_2 -agonists results in uncoupling and internalization or sequestration in the cells where they are degraded.⁴¹⁴ This net loss in the number of available functional β -receptors⁴¹⁵ results in “downregulation” of responsiveness to β_2 -agonists, which manifests as a lack of clinical protection to bronchoconstrictive stimuli. Restoration of sensitivity requires resynthesis of the receptor to the active state. This resynthesis is observed clinically within 72 hours of cessation of exposure to β_2 -agonist.^{27,410}

Stimulation of mast cell β -receptors normally inhibits mediator release. The process of β -receptor desensitization varies markedly among different cell types; bronchial mast cells are more easily desensitized than are BSM cells.^{414,416} Downregulation appears to occur more readily in mast cells as may occur with therapeutic administration of β_2 -agonists. For this

reason, the clinical effects of downregulation are evident more rapidly on mast cells with an effect on bronchoprotection than on smooth muscle and bronchodilation.⁴¹⁷ The downregulation of mast cell β -receptors not only enhances mediator release but potentially enhances bronchoconstriction as well.^{192,193,383,401,418}

This downregulation or tolerance is demonstrated clinically as a reduction in duration of β_2 -agonist bronchoprotection to stimuli such as exercise, which depends on mast cell mediator release for bronchoconstriction. Tolerance to bronchodilation is demonstrated by prolongation of the time of recovery from bronchoconstriction in response to usual doses of β_2 -agonists.^{27,401,402,419}

Downregulation of the β_2 -receptor is accompanied by augmentation of pathways mediated through the LT, histamine, and thromboxane receptors. Activation of these receptors has the added potential to enhance bronchoconstriction.^{194,195,382,401,402,420} Also, BHR may be induced by non-mast cell mediator mechanisms involving cholinergic agonists, for example, independently or with mast cell mediator mechanisms.^{421–423}

The complexity of mechanisms involved in β -adrenoreceptor functioning, however, is emphasized in a review.⁴²⁴ In investigations primarily using a mouse model, long-term β_2 -agonist exposure caused an increase, rather than decrease, in contractile signaling. Unexpectedly, long-term exposure to a highly selective β -receptor antagonist (ie, β -blocker) resulted in upregulation of β_2 -adrenoreceptors and bronchoprotection.⁴²⁵ An open-label investigation in humans with chronic mild asthma reported results consistent with this latter finding in the mouse model.⁴²⁵ Consequently, it is clear that a better appreciation of the mechanisms of β -adrenoreceptor functioning is needed. A more thorough understanding of the difference between the effects of intermittent and long-term use of β_2 -adrenergic agonists, BHR, and the production of tolerance is necessary.

From a clinical point of view, potential concerns associated with tolerance with long-term use of SABAs or LABAs need to be recognized. Intermittent use of β_2 -agonists is preferred and recommended for bronchoprotection.

The use of LABAs as daily monotherapy to provide overall asthma control is not recommended.³³ When ICS alone are not adequate in controlling chronic asthma, LABAs are often combined with ICS to provide effective maintenance therapy; however, there is no convincing clinical evidence that this combination diminishes tolerance to the bronchoprotective effect of LABAs in asthma or EIB with asthma.^{26,28,426} Long-acting β_2 -agonists alone used intermittently, up to 3 times a week, do not appear to be associated with tolerance and may be prescribed for EIB.^{410,427}

LT Inhibitors

Summary Statement 42. Daily therapy with LT inhibitors does not lead to tolerance and can be used for intermittent or maintenance prophylaxis; however, it provides protection

that may not be complete and has no use to reverse airway obstruction when it occurs. A

The role of LTs in EIB is to sustain the bronchoconstrictive and inflammatory response, although their role appears to vary significantly among patients. Correspondingly, inhibitors of the LT pathway (LTRAs and lipoxygenase inhibitors) are effective in enhancing recovery, and again, there is much variability in their effectiveness, completely blocking EIB in some asthmatic individuals but less so or not at all in others. There is a 30% to 80% attenuation of EIB, with approximately 50% of patients being responders. These percentages may vary, depending in part on the FEV₁ decrease required to make a diagnosis of EIB (>10%, >15%, or >20%) or used to define protection.^{24,379,400,428} Most patients do not experience complete protection.²⁴ This is not surprising given that other mediators (eg, PGD₂ and histamine)^{116,429} are involved in EIB.

Various LTRAs have been found to be effective in attenuating EIB.^{430,431} Most studies have examined specific LTD₄ receptor antagonist, particularly montelukast. Montelukast is approved by the FDA for treatment of EIB in adolescents and adults. These LTD₄ antagonists are administered orally, although bronchoprotection has been reported when some inhibitors have been given by inhalation.^{432–437} Montelukast acts within 1 to 2 hours of oral administration^{112,434,437} and has a bronchoprotective activity for 24 hours.^{112,124,370,438,439} Maximum protection may not be retained toward the end of this period.⁴⁴⁰ Leukotriene receptor antagonists also accelerate the time to recovery from EIB.^{124,412} Whereas LTRAs are not as effective overall in attenuating EIB as β_2 -agonists,²⁴ tolerance does not develop with long-term use.^{124,399,400,441} The variability in effect on EIB suggests populations of responders and nonresponders similar to that shown for the LT effects on overall asthma control.^{138,442,443}

A second group of agents that affects the LT pathway by inhibiting synthesis are the lipoxygenase inhibitors. Lipoxygenase inhibitors have also been shown to attenuate EIB when given orally^{428,436,444,445} or intravenously,⁴⁴⁶ but the duration of inhibition of these compounds is relatively short^{428,436} and they are not currently recommended for this indication.

Mast Cell Stabilizers

Summary Statement 43. Cromolyn sodium and nedocromil sodium (currently not available in the United States in an inhaled form) when inhaled shortly before exercise attenuate EIB but have a short duration of action. They do not have a bronchodilator activity. They may be effective alone or as added therapy with other drugs for EIB. A

Cromolyn sodium and nedocromil sodium are 2 structurally unrelated compounds that have no bronchodilator activity but have similar bronchoprotective activity against EIB when inhaled.^{29,30,111} Several mechanisms have been proposed for these agents, including interference with mast cell mediator release of PGD₂.¹²³ The bronchoprotective effect is rapid⁴⁴⁷ but of short duration (1–2 hours).^{448,449} These agents

may be effective when taken alone or when inhaled shortly before, and perhaps simultaneously with, exercise and may increase overall inhibition of EIB when combined with other drugs used to diminish EIB.^{29,377,448,450} Significant intersubject and between-study variability has been observed in the ability of these agents to attenuate EIB. Some studies found few or no subjects protected, whereas other studies showed complete protection.^{108,451} Effectiveness of cromolyn may be dose related.^{384,451,452} The formulations (1 mg per actuation) available in the United States did not deliver sufficient doses of cromolyn unless many inhalations were given (eg, as many as 20 at a time). Long-term use of either drug is not accompanied by tolerance. For this reason and because of their excellent safety profiles and rapidity of action, these agents can be used repeatedly to attenuate EIB in responsive individuals.^{29,453}

Inhaled Corticosteroids

Summary Statement 44. Although ICS therapy can decrease the frequency and severity of EIB, its use does not necessarily eliminate the need for additional acute therapy with β_2 -adrenergic agonists or other agents. A

Exercise-induced bronchoconstriction in otherwise symptomatic asthmatic patients is best controlled by maintenance anti-inflammatory treatment alone^{9,41,42} or in combination with other short-term preventive treatment.^{23,33,207} Inhaled corticosteroids improve overall asthma control in most patients with chronic persistent asthma. Use of ICS also is associated with attenuation of hyperresponsiveness to direct and indirect stimuli, including exercise.¹³ The effect of ICS on asthma and EIB is dose^{9,105} and time dependent and may be associated with decreases in inflammatory mediators.³² The relationship between improvement in control of persistent asthma and bronchoprotection, however, is imperfect. Nevertheless, the degree of EIB is considered a reflection of asthma control (or lack of control), and moderate to severe EIB, in particular, strongly suggests need for reassessment of therapy or another diagnosis.

Some bronchoprotective effect with ICS has been recorded as early as 4 hours after the first dose.^{32,106} After 1 week of long-term therapy, efficacy begins to plateau^{9,103,105}; however, bronchoprotection may increase over weeks or even months until it reaches its final plateau.^{8,41,454,455} Bronchoprotection has been shown to occur in 30% to 60% of asthmatic patients with EIB, with marked individual variability ranging from "complete" protection to little or no evidence of protection.⁸ In the absence of definitive dose-ranging and repetitive studies in individual patients, it is not clear whether this reflects distinct subpopulations of responders and nonresponders (eg, a reflection of genetic differences).

Inhaled corticosteroids do not necessarily obviate the need for acute bronchoprotection for EIB. β_2 -Adrenergic agonists can be added if necessary for short-term prevention of EIB.^{371,372} When maintenance ICS are not sufficiently effective, LTRAs can be used to obtain added protection, while

also administering β_2 -agonists for acute bronchoprotection if necessary.^{21,22,35,207}

Summary Statement 45. Inhaled corticosteroid therapy does not prevent the occurrence of tolerance from daily β_2 -agonist use. A

The preponderance of evidence indicates little amelioration by ICS of tolerance to β_2 -agonist bronchoprotection^{26,28,412,426,456} and a shortened degree of bronchoprotection remains when ICS and LABAs are given together. Nevertheless, one study that assessed the combination of an ICS and LABA (fluticasone and salmeterol) for maintenance therapy in adult patients indicated better bronchoprotection at 1 and 8.5 hours after dosing compared with the same dose of fluticasone alone during 4 weeks. In that study, most patients receiving the combined therapy also exhibited greater complete (<10% decrease of FEV₁) protection and overall asthma control.²⁸ A somewhat similar study with the same agents in children and adolescents also indicated a small persistent effect of bronchoprotection when the combination was used compared with the ICS alone.³¹

Anticholinergic Agents

Summary Statement 46. Although ipratropium bromide has been inconsistent in attenuating EIB, a few patients may be responsive to this agent. A

Anticholinergic agents (atropine sulfate and ipratropium bromide) have bronchodilator activity⁴²² by blocking vagally mediated tone and have been used alone and in conjunction with SABAs with some success in treating acute exacerbations of asthma.⁴⁵⁷ Efficacy of anticholinergic agents to prevent EIB,⁴⁵⁸ however, has not been consistent in double-blind studies, especially in placebo-controlled trials.⁴⁵⁹ Not all patients appear to respond to anticholinergic agents,^{29,450,460,461} and responsiveness may be variable in the same patient.⁴⁶² Studies should be performed to determine the characteristics of the responder population (perhaps based on increased cholinergic contributions to EIB in some patients).⁴²²

Methylxanthines, Antihistamines, and Other Agents

Summary Statement 47. Drugs in several other pharmacotherapeutic classes, including theophylline, antihistamines, calcium channel blockers, α -adrenergic receptor antagonists, inhaled furosemide, heparin, and hyaluronic acid, have been examined for actions against EIB with inconsistent results. B

Theophylline and aminophylline are methylxanthines that have been used for long-term maintenance therapy to treat persistent asthma. In recent years these agents have only been used as adjunct therapy to ICS or similar maintenance therapy when further control of asthma is needed.^{33,207} Caffeine also belongs to this class of drugs. Methylxanthines are nonselective phosphodiesterase inhibitors of the cyclic AMP and cyclic guanine monophosphate pathways that play a role in the pathophysiology of asthma. Methylxanthines are mild bronchodilators and modify EIB in some patients possibly in part due to their bronchodilator action.^{463,464} There are studies,

Table 4. Bronchial Provocation Challenge Comparisons for EIB

	Methacholine bronchial provocation challenge	Histamine challenge	Laboratory-based exercise challenge	Sports- specific exercise challenge	Eucapnic voluntary hyperpnea	Hypertonic saline challenge	Inhaled powdered mannitol	Inhaled adenosine monophosphate
Direct (acts directly on tissue) vs indirect (mediator release) challenge	D	D	ID	ID	ID	ID	ID	ID
Sensitivity for diagnosing EIB	Fair	Fair	VG-E	VG-E	VG-E	Fair	Fair	Fair
Sensitivity for diagnosing asthma	VG	VG	Good	Good	Good	VG	VG	VG
Specificity for diagnosing EIB	Fair	Fair	E	E	E	VG	VG	VG
Specificity for diagnosing asthma	Good	Good	VG	VG	VG	VG	VG	VG
PPV for diagnosing EIB	Fair	Fair	VG	VG	VG	Fair-Good	Fair-Good	Good
Fair-Good PPV for diagnosing asthma	Fair	Fair	VG	VG	VG	VG	VG	VG
NPV for diagnosing EIB	Fair	Fair	VG	VG	VG	Fair	Fair	Fair
NPV for diagnosing asthma	VG	VG	Fair	Fair	Fair	Fair	Fair	Fair
Produces dose-response curve	Yes	Yes	No	No	No	Yes	Yes	Yes
FDA approved	Yes	No	N/A	N/A	N/A	N/A	Yes	No
Correlates with airway inflammation	Mi	Mi	H	H	H	H	H	H
Correlates with current EIB symptoms	Mi	Mi	Mi	Mi	Mi	Mi	Mi	Mi
Requires subjects to be capable of exertion (eg, relatively free of cardiac disease)	No	No	Yes	Yes	No	No	No	No
Causes severe decreases in FEV ₁ (eg, greater than 30%)	No	No	Yes	Yes	Yes	No	No	No
Commonly causes cough	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Can perform sputum collection in conjunction with challenge	No	No	No	No	No	Yes	Yes	No
Provides response dose ratio	Yes	Yes	No	No	No	Yes	Yes	Yes

Abbreviations: D, direct; E, excellent; EIB, exercise-induced bronchoconstriction; F, fair; FDA, US Food and Drug Administration; FEV₁, forced expiratory volume in 1 second; F, frequently; G, good; H, high; ID, indirect; Mi, minimal; Mo, moderate; NPV, negative predictive value; PPV, positive predictive value; VG, very good.

however, that clearly show no benefit from methylxanthines given orally.⁴⁶⁵ Methylxanthines exhibit a relatively narrow therapeutic index with potentially serious adverse events such as seizures. Selective phosphodiesterase inhibitors are safer and may have efficacy similar to the methylxanthines. One such agent, roflumilast, is a phosphodiesterase 4 inhibitor that has been reported to attenuate mild EIB.⁴⁶⁶

Some antihistamines have been reported to attenuate EIB^{107,110,429,467–470} but with inconsistent results.^{113,471} Other antihistamines appear ineffective.⁴⁷² A possible explanation for this variability may relate to differences in the intensity and duration of the exercise stimulus, with greater intensity required for participation of histamine in the pathogenesis of EIB.¹²⁵ Also, histamine is only 1 of the 3 main mediators that contribute to EIB. In addition, the antihistamine class has pharmacodynamic diversity. For example, antihistamines may inhibit mediator activation and release and act on end organs and other histamine receptors.⁴⁷³ Different routes of administration and dosages of antihistamines may also be confounding factors in previous studies.⁴⁷⁴

Allergic rhinitis is a common finding in atopic asthmatic patients, and there is some evidence that effective treatment of nasal congestion and obstruction by nasal ICS is associated with at least mild reduction in EIB.^{56,454} However, other evidence⁴⁷² suggests that antihistamines used in children with allergic rhinitis and EIB may not be bronchoprotective. The conclusion is that there is an absence of definitive studies that have determined effectiveness of nasal or systemic antihistamines, used to treat allergic rhinitis, for an effect on EIB. It is likely to remain common practice to use antihistamines to treat allergic rhinitis in the hope that there will be some effect on EIB. Definitive studies are still needed to confirm the utility of this practice.

Numerous other compounds have been examined for activity against EIB and may have moderate effectiveness in some situations.³⁷⁸ These include calcium channel blockers,^{475,476} inhaled furosemide,^{477–479} some α -adrenergic receptor antagonists (oral and inhaled),^{458,480} inhaled heparin,⁴⁸¹ and hyaluronic acid.⁴⁸² These agents do not always produce consistent results in preventing EIB. The effectiveness of some of these agents does not necessarily apply to other members of the same drug class, suggesting various mechanisms of action, not necessarily related to the obvious mechanisms attributed to each class of drug. The effectiveness of diverse kinds of drugs suggests that there are multiple mechanisms underlying EIB.¹² Although these drugs are not recommended for clinical use against EIB, these and other agents may be useful as probes in studying possible mechanisms underpinning EIB. In addition, it is important to recognize that these agents may interfere with clinical protocols that seek to examine the effects of other experimental drugs on EIB.

Nonpharmacologic Therapy

Summary Statement 48. Preexercise warm-up may be helpful in reducing the severity of EIB. A

Warm-up before exercise was studied on postexercise bronchoconstriction in athletes with EIB. Continuous

warm-up before exercise was shown to cause significant decrease in postexercise bronchoconstriction in some athletes. This has importance in patient education, and health care professionals should tell patients that preexercise warm-up should be done at 60% to 80% HR_{max} to provide partial attenuation of EIB; this refractory period may last typically from 1 to 3 hours and occasionally 4 hours.^{129–131} However, this does not alleviate the need for medications. Albuterol plus a warm-up gives better protection than the warm-up or albuterol alone.^{483,484}

Summary Statement 49. Reduction of sodium intake and ingestion of fish oil and ascorbic acid supplementation may be helpful in reducing the severity of EIB. A

A low sodium diet maintained for 1 to 2 weeks decreases EIB, but long-term effects of a low sodium diet on either the prevalence or severity of asthma or on EIB are unknown. These studies involve small numbers of individuals and need to be replicated before a low-salt diet is widely recommended. The low-sodium diet may be considered in addition to pharmacologic therapy of EIB.^{34,485,486}

There is some evidence that a variety of dietary factors, such as dietary ω -3 polyunsaturated fatty acids, can reduce EIB. If these data are reproducible, it may be a new and important therapy that is safe and effective.³⁴

Two weeks of ascorbic acid supplementation has been shown to improve pulmonary function, asthma symptom scores, fraction of eNO, and urinary LTs in asthmatic patients with EIB. This provides a potential nonpharmacologic recommendation for EIB patients.⁴⁸⁷

Competitive and Elite Athletes

Summary Statement 50. Exercise-induced bronchoconstriction alone in elite athletes may have different characteristics than EIB with asthma in elite athletes or EIB in the general population. These divergent characteristics may include pathogenesis, presentation, diagnosis, management, and the requirement by governing bodies to obtain permission to receive pharmaceutical agents. D

Exercise-induced bronchoconstriction is common in elite athletes who do not otherwise have symptoms of asthma.

Summary Statement 51. Airway inflammation in elite athletes may be related to the high intensity of physical training, high minute ventilation, and inhalation of airborne pollutants and allergens. D

Exercise-induced bronchoconstriction in elite athletes may be associated with a mixed neutrophil-eosinophil type airway inflammation in contrast to the more commonly observed eosinophil-dominant inflammatory cascade reported with chronic asthma.²³⁵ In addition, in elite athletes (especially those competing in high air pollution environments), Leukotrienes may play a predominant role in the EIB response.^{166,189} Endurance athletes, in particular, may develop respiratory symptoms and have BHR as a consequence of prolonged strenuous exercise and training at high rates of ventilation. The underlying airway inflammation in elite athletes may also be related to the high

intensity of the physical training with high minute ventilation, which has potential to cause injury to the airway epithelium. This injury is likely to occur when exercise is performed in adverse environmental conditions, such as cold dry air, or when there is prolonged exposure to chemicals at indoor swimming pools or exhaust fumes from ice resurfacers used to groom ice hockey rinks.^{158–160,162,163} The high minute ventilation may aggravate effects of environmental exposures during physical activity, permitting airborne allergens and irritants to enter the small airways. In the process of conditioning dry air in various sports activities, the airways may be injured, which is likely a consequence of the humidification process occurring deeper in the airways.^{10,13}

Summary Statement 52. The diagnosis of EIB, whether alone or with asthma, in elite athletes may be difficult because history and presentation are not reliable. Objective testing is necessary to diagnose the condition accurately. A

Diagnosing EIB in elite athletes may be complicated by their apparently excellent physical condition, which causes the athlete and his or her supporting personnel (eg, coaches, family members, teammates, primary care physicians, and trainers) not to appreciate that he or she may be experiencing symptoms of asthma. Many elite athletes may not acknowledge or may even deny typical daily asthma symptoms, such as cough, wheezing, tight chest, or prolonged shortness of breath. Moreover, baseline pulmonary function tests may not indicate airway obstruction in these individuals. In fact, many athletes have higher than normal baseline pulmonary function such that a normal percentage predicted FEV₁ may be observed even in the presence of airway obstruction.

Various bronchial provocation challenges may be useful in identifying EIB. The indirect bronchial provocation challenges are most useful in this setting and include exercise challenge in the athlete's sport (either in the natural setting or simulated conditions in the laboratory), EVH, and inhaled hyperosmolar aerosols (mannitol, saline). The exercise challenge in the athlete's venue may be logistically difficult to accomplish; however, reproducibility of the response to a standardized exercise challenge in the laboratory can also be an issue.⁴⁸⁸ Other tests, such as dry air laboratory exercise challenge, may be used as surrogates. In fact, because environmental and exercise intensity varies, a well-controlled dry air laboratory challenge may provide the most accurate and reliable test results. The challenge test recommended by the IOC-MC's Independent Panel on Asthma is EVH, and the same panel accepts mannitol testing.^{35,99,292} Table 4 compares challenges, and descriptions of how these challenges should be performed and can be found in the "Diagnosis" section.

In a recent comparative study of Scottish elite swimmers, mannitol was associated with high baseline eNO and inflammatory phenotype, whereas field-based exercise and chlorine challenge identified swimmer-specific bronchoconstrictive response. Thus, chlorine exposure, which was associated with a sustained decrease in eNO, may represent a neurogenic response in these elite athletes.⁴⁸⁹

Summary Statement 53. In general, the treatment of EIB in patients who have asthma is similar in both recreational and elite athletes. However, the efficacy of therapy for EIB alone in athletes at any level is not well established. A

The management of EIB in elite athletes is similar to that for recreational athletes and should include reducing relevant environmental exposures as much as possible; treating associated comorbid conditions; appropriate pharmacotherapy for control of symptoms, prophylaxis, and rescue; and patient education.³⁵ An individualized exercise prescription considering the athlete's venue may need to be designed by the athlete and the specialist to provide adequate control of EIB or EIB with asthma (eg, swimmers).

Unfortunately, the environmental exposures are often not controllable because athletes may have to practice or compete for long periods in air that is cold, polluted, or has high pollen allergen or chemical irritant levels.

It is recommended that controller pharmacotherapy for athletes who have EIB with asthma should include daily ICS. The combination of ICS plus LABAs is not recommended because of the potential for tolerance to develop with daily use of β_2 -agonists. This tolerance reduces the duration of bronchoprotection in exercise afforded by the β_2 -agonist. In some patients with concomitant moderate to severe persistent asthma, however, combination therapy may have added utility. The athlete's performance results should be monitored carefully because spirometry and symptoms alone may not be reliable end points to monitor control of asthma. If symptoms of EIB in athletes who have asthma are not controlled despite appropriate inhaler technique and adherence with therapy, treatment can be stepped up by increasing the dose of ICS or addition of other medications. However, symptoms of EIB in the elite athlete who does not otherwise appear to have asthma have been reported to be less responsive to ICS, perhaps reflecting neutrophilic inflammation.^{35,201} Consideration should be given to oral LT antagonists, inhaled cromolyn, and inhaled nedocromil.^{30,109,114} There may be reason to believe that some individuals may respond better than others to any of these various agents, based on genetics or other differences.⁴⁹⁰ Inhaled anticholinergics, such as ipratropium or tiotropium, may be considered, but studies on these drugs in asthma have given mixed results and they are not considered standard therapy for EIB.^{35,491} Ideally, elite athletes should only use β_2 -agonists intermittently due to the potential for tolerance described above.^{35,99} A lack of appropriate response to therapy requires additional evaluation of the athlete and consideration of the differential diagnosis (see the "Differential Diagnosis" section).

Athletes in sports with high ventilation rates (eg, swimming, mountain biking, rowing, biathlon, cross country skiing, and skating events) may develop respiratory symptoms compatible with EIB alone, with or without demonstrating a positive challenge test result indicative of asthma. It has been proposed that the repetitive epithelial injury repair cycle, in response to high ventilation rate, results in changes in the contractile properties of BSM as a result of exposure to

plasma-derived products from exudation.¹⁰ For some athletes who have allergy, this exposure may lead to in vivo passive sensitization of the BSM.¹⁰ This epithelial injury and repair cycle^{227,245} may represent a form of overuse syndrome, and therefore the athlete may benefit from limitation of activity; however, this may be unrealistic for the elite endurance athlete.¹⁸⁸ The pathogenesis of this type of BHR in athletes who only have EIB may be different from that classically observed in athletes who have chronic asthma, where inflammatory cells such as eosinophils are important. Thus, these athletes who have EIB alone may have different mechanisms for the cause of their symptoms and may not respond to the medications typically used for asthma.^{492,493}

These parameters are also available on the Internet at <http://www.jcaai.org>.

REFERENCES

- Rundell KW, Im J, Mayers LB, Wilber RL, Szmedra L, Schmitz HR. Self-reported symptoms and exercise-induced asthma in the elite athlete. *Med Sci Sports Exerc.* 2001;33:208–13. III.
- Parsons JP, Kaeding C, Phillips G, Jarjoura D, Wadley G, Mastronarde JG. Prevalence of exercise-induced bronchospasm in a cohort of varsity college athletes. *Med Sci Sports Exerc.* 2007;39:1487–92. III.
- Rundell KW, Slee JB. Exercise and other indirect challenges to demonstrate asthma or exercise-induced bronchoconstriction in athletes. *J Allergy Clin Immunol.* 2008;122:238–48. IV.
- Holzer K, Anderson SD, Douglass J. Exercise in elite summer athletes: challenges for diagnosis. *J Allergy Clin Immunol.* 2002;110:374–80. IV.
- Bye MR, Kerstein D, Barsh E. The importance of spirometry in the assessment of childhood asthma. *Am J Dis Child.* 1992;146:977–8. IV.
- Crapo RO, Casaburi R, Coates AL, et al. Guidelines for methacholine and exercise challenge testing—1999. This official statement of the American Thoracic Society was adopted by the ATS Board of Directors, July 1999. *Am J Respir Crit Care Med.* 2000;161:309–29. IV.
- Weiler JM, Bonini S, Coifman R, et al. American Academy of Allergy, Asthma & Immunology Work Group report: exercise-induced asthma. *J Allergy Clin Immunol.* 2007;119:1349–58. IV.
- Koh MS, Tee A, Lasserson TJ, Irving LB. Inhaled corticosteroids compared to placebo for prevention of exercise induced bronchoconstriction. *Cochrane Database Syst Rev.* 2007;CD002739. Ia.
- Subbarao P, Duong M, Adelroth E, et al. Effect of ciclesonide dose and duration of therapy on exercise-induced bronchoconstriction in patients with asthma. *J Allergy Clin Immunol.* 2006;117:1008–13. Ib.
- Anderson SD, Kippelen P. Airway injury as a mechanism for exercise-induced bronchoconstriction in elite athletes. *J Allergy Clin Immunol.* 2008;122:225–35. IV.
- Hallstrand TS, Henderson WR Jr. An update on the role of leukotrienes in asthma. *Curr Opin Allergy Clin Immunol.* 2010;10:60–6. IV.
- Anderson SD, Daviskas E. The mechanism of exercise-induced asthma is ... *J Allergy Clin Immunol.* 2000;106:453–9. IV.
- Anderson SD, Holzer K. Exercise-induced asthma: is it the right diagnosis in elite athletes? *J Allergy Clin Immunol.* 2000;106:419–28. IV.
- Ober C, Hoffjan S. Asthma genetics 2006: the long and winding road to gene discovery. *Genes Immun.* 2006;7:95–100. IV.
- Hilberg T, Deigner HP, Möller E, et al. Transcription in response to physical stress—clues to the molecular mechanisms of exercise-induced asthma. *FASEB J.* 2005;19:1492–4. IIa.
- Hallstrand TS, Debley JS, Farin FM, Henderson WR Jr. Role of MUC5AC in the pathogenesis of exercise-induced bronchoconstriction. *J Allergy Clin Immunol.* 2007;119:1092–8. IIb.
- Carlsen KH, Engh G, Mørk M. Exercise-induced bronchoconstriction depends on exercise load. *Respir Med.* 2000;94:750–5. III.
- Cockcroft D, Davis B. Direct and indirect challenges in the clinical assessment of asthma. *Ann Allergy Asthma Immunol.* 2009;103:363–9. IV.
- Abu-Hasan M, Tannous B, Weinberger M. Exercise-induced dyspnea in children and adolescents: if not asthma then what? *Ann Allergy Asthma Immunol.* 2005;94:366–71. IIb.
- Weinberger M, Abu-Hasan M. Perceptions and pathophysiology of dyspnea and exercise intolerance. *Pediatr Clin North Am.* 2009;56:33–48, ix. III.
- Grzelewski T, Stelmach I. Exercise-induced bronchoconstriction in asthmatic children: a comparative systematic review of the available treatment options. *Drugs.* 2009;69(12):1533–53. IV.
- Carlsen KH, Anderson SD, Bjermer L, et al. Treatment of exercise induced asthma, respiratory and allergic disorders in sports and the relationship to doping: Part II of the report from the Joint Task Force of European Respiratory Society (ERS) and European Academy of Allergy and Clinical Immunology (EAACI) in cooperation with GA(2)LEN. *Allergy.* 2008;63:492–505. IV.
- Stelmach I, Grzelewski T, Majak P, Jerzynska J, Stelmach W, Kuna P. Effect of different antiasthmatic treatments on exercise-induced bronchoconstriction in children with asthma. *J Allergy Clin Immunol.* 2008;121:383–9. Ib.
- Raissy HH, Harkins M, Kelly F, Kelly HW. Pretreatment with albuterol versus montelukast for exercise-induced bronchospasm in children. *Pharmacotherapy.* 2008;28:287–94. Ib.
- Ramage L, Lipworth BJ, Ingram CG, Cree IA, Dhillon DP. Reduced protection against exercise induced bronchoconstriction after chronic dosing with salmeterol. *Respir Med.* 1994;88:363–8. Ib.
- Simons FE, Gerstner TV, Cheang MS. Tolerance to the bronchoprotective effect of salmeterol in adolescents with exercise-induced asthma using concurrent inhaled glucocorticoid treatment. *Pediatrics.* 1997;99:655–9. Ib.
- Haney S, Hancox RJ. Rapid onset of tolerance to beta-agonist bronchodilation. *Respir Med.* 2005;99:566–71. IIa.
- Weiler JM, Nathan RA, Rupp NT, Kalberg CJ, Emmett A, Dorinsky PM. Effect of fluticasone/salmeterol administered via a single device on exercise-induced bronchospasm in patients with persistent asthma. *Ann Allergy Asthma Immunol.* 2005;94:65–72. Ia.
- Spooner CH, Spooner GR, Rowe BH. Mast-cell stabilising agents to prevent exercise-induced bronchoconstriction. *Cochrane Database Syst Rev.* 2003;4:CD002307. Ia.
- Kippelen P, Larsson J, Anderson SD, Brannan JD, Dahlén B, Dahlén SE. Effect of sodium cromoglycate on mast cell mediators during hyperpnea in athletes. *Med Sci Sports Exerc.* 2010;42:1853–60. IIa.
- Pearlman D, Qaundah P, Matz J, Yancey SW, Stempel DA, Ortega HG. Fluticasone propionate/salmeterol and exercise-induced asthma in children with persistent asthma. *Pediatr Pulmonol.* 2009;44:429–35. Ib.
- Kippelen P, Larsson J, Anderson SD, et al. Acute effects of beclomethasone on hyperpnea-induced bronchoconstriction. *Med Sci Sports Exerc.* 2010;42:273–80. IIa.
- National Asthma Education and Prevention Program. Expert Panel Report 3 (EPR-3): Guidelines for the Diagnosis and Management of Asthma—Summary Report 2007. *J Allergy Clin Immunol.* 2007;120(suppl 5): S94–S138. IV.
- Mickleborough TD. A nutritional approach to managing exercise-induced asthma. *Exerc Sport Sci Rev.* 2008;36:135–44. Ib.
- Fitch KD, Sue-Chu M, Anderson SD, et al. Asthma and the elite athlete: summary of the International Olympic Committee's consensus conference, Lausanne, Switzerland, January 22–24, 2008. *J Allergy Clin Immunol.* 2008;122:254–60, e1–7. IV.
- Sterk PJ, Fabbri LM, Quanjer PH, et al. Airway response. Standardized provocation tests in adults: pharmacological, physical and sensitizing stimuli. Work Group on Standardization of Respiratory Function Tests. European Community for Coal and Steel. Official position of the European Respiratory Society. *Eur Respir J* 1993;6 (Supp 16):53–83. IV.
- Godfrey S, Springer C, Bar-Yishay E, Avital A. Cut-off points defining normal and asthmatic bronchial reactivity to exercise and inhalation

- challenges in children and young adults. *Eur Respir J*. 1999;14:659–68. Ia.
38. Kattan M, Keens TG, Mellis CM, Levison H. The response to exercise in normal and asthmatic children. *J Pediatr*. 1978;92:718–21. IIA.
 39. Anderson SD, Charlton B, Weiler JM, et al. Comparison of mannitol and methacholine to predict exercise-induced bronchoconstriction and a clinical diagnosis of asthma. *Respir Res*. 2009;10:4.Ib.
 40. Haby MM, Peat JK, Mellis CM, Anderson SD, Woolcock AJ. An exercise challenge for epidemiological studies of childhood asthma: validity and repeatability. *Eur Respir J*. 1995;8:729–36. IIA.
 41. Hofstra WB, Neijens HJ, Duiverman EJ, et al. Dose-response over time to inhaled fluticasone propionate treatment of exercise- and methacholine-induced bronchoconstriction in children with asthma. *Pediatr Pulmonol*. 2000;29:415–23. IIB.
 42. Jonasson G, Carlsen KH, Hultquist C. Low-dose budesonide improves exercise-induced bronchospasm in schoolchildren. *Pediatr Allergy Immunol*. 2000;11:120–5. Ib.
 43. Niinimaa V, Cole P, Mintz S, Shephard RJ. The switching point from nasal to oronasal breathing. *Respir Physiol*. 1980;42:61–71. III.
 44. Anderson SD, Togias A. Dry air and hyperosmolar challenge in asthma and rhinitis. In: Busse WW, Holgate ST, eds. *Asthma and Rhinitis*, 2nd ed. Malden, MA: Blackwell Science Ltd; 1995:1449–68. IV.
 45. Anderson SD. Asthma provoked by exercise, hyperventilation, and the inhalation of non-isotonic aerosols. In: Barnes PJ, Rodger IW, Thomson NC, eds. *Asthma: Basic Mechanisms and Clinical Management*. 2nd ed. London, England: Academic Press Ltd; 1992:473–90. IV.
 46. Bar-Or O, Neuman I, Dotan R. Effects of dry and humid climates on exercise-induced asthma in children and preadolescents. *J Allergy Clin Immunol*. 1977;60:163–8. IIA.
 47. Chen WY, Horton DJ. Heat and water loss from the airways and exercise-induced asthma. *Respiration*. 1977;34:305–13. IIB.
 48. Strauss RH, McFadden ER Jr, Ingram RH Jr, Deal EC Jr, Jaeger JJ, Stearns D. Influence of heat and humidity on the airway obstruction induced by exercise in asthma. *J Clin Invest*. 1978;61:433–40. IIB.
 49. Deal EC Jr, McFadden ER Jr, Ingram RH Jr, Jaeger JJ. Hyperpnea and heat flux: initial reaction sequence in exercise-induced asthma. *J Appl Physiol*. 1979;46:476–83. IIB.
 50. McFadden ER Jr, Pichurko BM, Bowman HF, et al. Thermal mapping of the airways in humans. *J Appl Physiol*. 1985;58:564–70. IIB.
 51. Daviskas E, Gonda I, Anderson SD. Local airway heat and water vapour losses. *Respir Physiol*. 1991;84:115–32. IV.
 52. Anderson SD, Daviskas E, Schoeffel RE, Unger SF. Prevention of severe exercise-induced asthma with hot humid air. *Lancet*. 1979;2:(8143)629. IIB.
 53. Anderson SD, Schoeffel RE, Follet R, Perry CP, Daviskas E, Kendall M. Sensitivity to heat and water loss at rest and during exercise in asthmatic patients. *Eur J Respir Dis*. 1982;63:459–71. IIB.
 54. Silverman M, Anderson SD. Standardization of exercise tests in asthmatic children. *Arch Dis Child*. 1972;47:882–9. IIA.
 55. Hallstrand TS, Bates PW, Schoene RB. Aerobic conditioning in mild asthma decreases the hyperpnea of exercise and improves exercise and ventilatory capacity. *Chest*. 2000;118:1460–9. IIA.
 56. Shturman-Ellstein R, Zeballos RJ, Buckley JM, Souhrada JF. The beneficial effect of nasal breathing on exercise-induced bronchoconstriction. *Am Rev Respir Dis*. 1978;118:65–73. IIB.
 57. Schachter EN, Lach E, Lee M. The protective effect of a cold weather mask on exercise-induced asthma. *Ann Allergy*. 1981;46:12–6. IIB.
 58. Millqvist E, Bengtsson U, Löwhagen O. Combining a beta2-agonist with a face mask to prevent exercise-induced bronchoconstriction. *Allergy*. 2000 Jul;55:672–5. IIB.
 59. Daviskas E, Gonda I, Anderson SD. Mathematical modeling in heat and water transport in the human respiratory tract. *J Appl Physiol*. 1990;69:362–72. IV.
 60. Siddiqui S, Mistry V, Doe C, et al. Airway hyperresponsiveness is dissociated from airway wall structural remodeling. *J Allergy Clin Immunol*. 2008;122:335–41, 341.e1-3. IIA.
 61. Carroll NG, Mutavdzic S, James AL. Distribution and degranulation of airway mast cells in normal and asthmatic subjects. *Eur Respir J*. 2002 May;19:879–85. IIA.
 62. Freed AN, Davis MS. Hyperventilation with dry air increases airway surface fluid osmolality in canine peripheral airways. *Am J Respir Crit Care Med*. 1999;159:1101–7. LB.
 63. Gilbert IA, Fouke JM, McFadden ER Jr. Heat and water flux in the intrathoracic airways and exercise-induced asthma. *J Appl Physiol*. 1987;63:1681–91. IIA.
 64. McFadden ER Jr. Hypothesis: exercise-induced asthma as a vascular phenomenon. *Lancet*. 1990;335(8694):880–3. IV.
 65. McFadden ER Jr, Pichurko BM. Intraairway thermal profiles during exercise and hyperventilation in normal man. *J Clin Invest*. 1985;76:1007–10. IIB.
 66. McFadden ER Jr, Lenner KA, Strohl KP. Postexertional airway re-warming and thermally induced asthma: new insights into pathophysiology and possible pathogenesis. *J Clin Invest*. 1986;78:18–25. IIA.
 67. Schoeffel RE, Anderson SD, Altounyan RE. Bronchial hyperreactivity in response to inhalation of ultrasonically nebulised solutions of distilled water and saline. *Br Med J (Clin Res Ed)*. 1981;283:1285–7. IIA.
 68. Anderson SD. Is there a unifying hypothesis for exercise-induced asthma? *J Allergy Clin Immunol*. 1984;73:660–5. IV.
 69. Anderson SD, Schoeffel RE, Black JL, Daviskas E. Airway cooling as the stimulus to exercise-induced asthma—a re-evaluation. *Eur J Respir Dis*. 1985;67:20–30. IIA.
 70. Hahn A, Anderson SD, Morton AR, Black JL, Fitch KD. A reinterpretation of the effect of temperature and water content of the inspired air in exercise-induced asthma. *Am Rev Respir Dis*. 1984;130:575–9. IIB.
 71. Aitken ML, Marini JJ. Effect of heat delivery and extraction on airway conductance in normal and in asthmatic subjects. *Am Rev Respir Dis*. 1985;131:357–61. IIA.
 72. Kivity S, Greif J, Reisner B, Fireman E, Topilsky M. Bronchial inhalation challenge with ultrasonically nebulized saline; comparison to exercise-induced asthma. *Ann Allergy*. 1986;57:355–8. IIA.
 73. Belcher NG, Rees PJ, Clark TJM, Lee TH. A comparison of the refractory periods induced by hypertonic airway challenge and exercise in bronchial asthma. *Am Rev Respir Dis*. 1987;135:822–5. IIB.
 74. Boulet LP, Turcotte H. Comparative effects of hyperosmolar saline inhalation and exercise in asthma. *Immunol Allergy Practice*. 1989;11:93–100. IIB.
 75. Anderson SD, Daviskas E. The airway microvasculature and exercise-induced asthma. *Thorax*. 1992;47:748–52. IV.
 76. Ingenito E, Solway J, Lafleur J, Lombardo A, Drazen JM, Pichurko B. Dissociation of temperature-gradient and evaporative heat loss during cold gas hyperventilation in cold-induced asthma. *Am Rev Respir Dis*. 1988;138:540–6. IIB.
 77. Argyros GJ, Phillips YY, Rayburn DB, Rosenthal RR, Jaeger JJ. Water loss without heat flux in exercise-induced bronchospasm. *Am Rev Respir Dis*. 1993;147:1419–24. IIA.
 78. Freed AN, Omori C, Hubbard WC, Adkinson NF. Dry air- and hypertonic aerosol-induced bronchoconstriction and cellular responses in the canine lung periphery. *Eur Respir J*. 1994;7:1308–16. Ib.
 79. Reiss TF, Hill JB, Harman E, et al. Increased urinary excretion of LTE4 after exercise and attenuation of exercise-induced bronchospasm by montelukast, a cysteinyl leukotriene receptor antagonist. *Thorax*. 1997;52:1030–5. Ib.
 80. O'Sullivan S, Roquet A, Dahlén B, et al. Evidence for mast cell activation during exercise-induced bronchoconstriction. *Eur Respir J*. 1998;12:345–50. IIB.
 81. Brannan JD, Gulliksson M, Anderson SD, Chew N, Kumlin M. Evidence of mast cell activation and leukotriene release after mannitol inhalation. *Eur Respir J*. 2003;22:491–6. IIA.
 82. Eggleston PA, Kagey-Sobotka A, Lichtenstein LM. A comparison of the osmotic activation of basophils and human lung mast cells. *Am Rev Respir Dis*. 1987;135:1043–8. Ib.
 83. Gulliksson M, Palmberg L, Nilsson G, Ahlstedt S, Kumlin M. Release of prostaglandin D2 and leukotriene C4 in response to hyperosmolar stimulation of mast cells. *Allergy*. 2006;61:1473–9. Ib.

84. Moloney ED, Griffin S, Burke CM, Poulter LW, O'Sullivan S. Release of inflammatory mediators from eosinophils following a hyperosmolar stimulus. *Respir Med*. 2003;97:928–32. LB.
85. Davis MS, Daviskas E, Anderson SD. Airway surface fluid desiccation during isocapnic hyperpnea. *J Appl Physiol*. 2003;94:2545–6. LB.
86. Daviskas E, Anderson SD, Gonda I, Chan HK, Cook P, Fulton R. Changes in mucociliary clearance during and after isocapnic hyperventilation in asthmatic and healthy subjects. *Eur Respir J*. 1995;8:742–51. IIa.
87. Eschenbacher WL, Sheppard D. Respiratory heat loss is not the sole stimulus for bronchoconstriction induced by isocapnic hyperpnea with dry air. *Am Rev Respir Dis*. 1985;131:894–901. IIb.
88. Tabka Z, Ben Jebria A, Vergeret J, Guenard H. Effect of dry warm air on respiratory water loss in children with exercise-induced asthma. *Chest*. 1988;94:81–6. IIa.
89. Chan-Yeung MM, Vyas MN, Grzybowski S. Exercise-induced asthma. *Am Rev Respir Dis*. 1971;104:915–23. IIb.
90. Kivity S, Souhrada JF. Hyperpnea: the common stimulus for bronchospasm in asthma during exercise and voluntary isocapnic hyperpnea. *Respiration*. 1980;40:169–77. IIb.
91. Phillips YY, Jaeger JJ, Laube BL, Rosenthal RR. Eucapnic voluntary hyperventilation of compressed gas mixture. A simple system for bronchial challenge by respiratory heat loss. *Am Rev Respir Dis*. 1985;131:31–5. IIa.
92. Eliasson AH, Phillips YY, Rajagopal KR, Howard RS. Sensitivity and specificity of bronchial provocation testing. An evaluation of four techniques in exercise-induced bronchospasm. *Chest*. 1992;102:347–55. IIa.
93. Mannix ET, Manfredi F, Farber MO. A comparison of two challenge tests for identifying exercise-induced bronchospasm in figure skaters. *Chest*. 1999;115:649–53. IIb.
94. Rundell KW, Anderson SD, Spiering BA, Judelson DA. Field exercise vs laboratory eucapnic voluntary hyperventilation to identify airway hyperresponsiveness in elite cold weather athletes. *Chest*. 2004;125:909–15. IIb.
95. Pedersen L, Winther S, Backer V, Anderson SD, Larsen KR. Airway responses to eucapnic hyperpnea, exercise and methacholine in elite swimmers. *Med Sci Sports Exerc*. 2008;40:1567–72. IIb.
96. Dickinson J, McConnell A, Whyte G. Diagnosis of exercise-induced bronchoconstriction: eucapnic voluntary hyperpnea challenges identify previously undiagnosed elite athletes with exercise-induced bronchoconstriction. *Br J Sports Med*. 2010;Jul 20. [Epub ahead of print]. IIb.
97. Argyros GJ, Roach JM, Hurwitz KM, Eliasson AH, Phillips YY. Eucapnic voluntary hyperventilation as a bronchoprovocation technique: development of a standardized dosing schedule in asthmatics. *Chest*. 1996;109:1520–4. III.
98. Hurwitz KM, Argyros GJ, Roach JM, Eliasson AH, Phillips YY. Interpretation of eucapnic voluntary hyperventilation in the diagnosis of asthma. *Chest*. 1995;108:1240–5. IIa.
99. Carlsen KH, Anderson SD, Bjermer L, et al. Exercise-induced asthma, respiratory and allergic disorders in elite athletes: epidemiology, mechanisms and diagnosis: part I of the report from the Joint Task Force of the European Respiratory Society (ERS) and the European Academy of Allergy and Clinical Immunology (EAACI) in cooperation with GA2LEN. *Allergy*. 2008;63:387–403. IV.
100. Yoshikawa T, Shoji S, Fujii T, et al. Severity of exercise-induced bronchoconstriction is related to airway eosinophilic inflammation in patients with asthma. *Eur Respir J*. 1998;12:879–84. IIb.
101. Kivity S, Argaman A, Onn A, et al. Eosinophil influx into the airways in patients with exercise-induced asthma. *Respir Med*. 2000;94:1200–5. IIa.
102. Gauvreau GM, Ronnen GM, Watson RM, O'Byrne PM. Exercise-induced bronchoconstriction does not cause eosinophilic airway inflammation or airway hyperresponsiveness in subjects with asthma. *Am J Respir Crit Care Med*. 2000;162:1302–7. Ia.
103. Duong M, Subbarao P, Adelroth E, et al. Sputum eosinophils and the response of exercise-induced bronchoconstriction to corticosteroid in asthma. *Chest*. 2008;133:404–11. III.
104. Lee SY, Kim HB, Kim JH, et al. Eosinophils play a major role in the severity of exercise-induced bronchoconstriction in children with asthma. *Pediatr Pulmonol*. 2006;41:1161–6. IIa.
105. Pedersen S, Hansen OR. Budesonide treatment of moderate and severe asthma in children: a dose-response study. *J Allergy Clin Immunol*. 1995;95(1 pt 1):29–33. Ib.
106. Thio BJ, Slingerland GL, Nagelkerke AF, Roord JJ, Mulder PGH, Dankert-Roelse JE. Effects of single-dose fluticasone on exercise-induced asthma in asthmatic children: a pilot study. *Pediatr Pulmonol*. 2001;32:115–21. IIa.
107. Patel KR. Terfenadine in exercise-induced asthma. *Br Med J (Clin Res Ed)*. 1984;288:1496–7. Ib.
108. Tullett WM, Tan KM, Wall RT, Patel KR. Dose-response effect of sodium cromoglycate pressurised aerosol in exercise induced asthma. *Thorax*. 1985;40:41–4. Ib.
109. Oseid S, Mellbye E, Hem E. Effect of nedocromil sodium on exercise-induced bronchoconstriction exacerbated by inhalation of cold air. *Scand J Med Sci Sports*. 1995;5:88–93. Ib.
110. Baki A, Orhan F. The effect of loratadine in exercise-induced asthma. *Arch Dis Child*. 2002;86:38–9. Ib.
111. Kelly KD, Spooner CH, Rowe BH. Nedocromil sodium versus sodium cromoglycate in treatment of exercise-induced bronchoconstriction: a systematic review. *Eur Respir J*. 2001;17:39–45. Ia.
112. Pearlman DS, van Adelsberg J, Philip G, et al. Onset and duration of protection against exercise-induced bronchoconstriction by a single oral dose of montelukast. *Ann Allergy Asthma Immunol*. 2006;97:98–104. Ia.
113. Dahlén B, Roquet A, Inman MD, et al. Influence of zafirlukast and loratadine on exercise-induced bronchoconstriction. *J Allergy Clin Immunol*. 2002;109(5 pt 1):789–93. Ib.
114. Rundell KW, Spiering BA, Baumann JM, Evans TM. Effects of montelukast on airway narrowing from eucapnic voluntary hyperventilation and cold air exercise. *Br J Sports Med*. 2005;39:232–6. Ib.
115. Nagakura T, Obata T, Shichijo K, et al. GC/MS analysis of urinary excretion of 9 α ,11 β PGF₂ in acute and exercise-induced asthma in children. *Clin Exp Allergy*. 1998;28:181–6. Ib.
116. Hallstrand TS, Moody MW, Wurfel MM, Schwartz LB, Henderson WR Jr, Aitken ML. Inflammatory basis of exercise-induced bronchoconstriction. *Am J Respir Crit Care Med*. 2005;172:679–86. Ib.
117. Haverkamp HC, Dempsey JA, Pegelow DF, et al. Treatment of airway inflammation improves exercise pulmonary gas exchange and performance in asthmatic subjects. *J Allergy Clin Immunol*. 2007;120:39–47. Ib.
118. Patel KR, Berkin KE, Kerr JW. Dose-response study of sodium cromoglycate in exercise-induced asthma. *Thorax*. 1982;37:663–6. IIa.
119. Eggleston PA, Kagey-Sobotka A, Schleimer RP, Lichtenstein LM. Interaction between hyperosmolar and IgE-mediated histamine release from basophils and mast cells. *Am Rev Respir Dis*. 1984;130:86–91. LB.
120. Van Rensen EL, Hiemstra PS, Rabe KF, Sterk PJ. Assessment of microvascular leakage via sputum induction: the role of substance P and neurokinin A in patients with asthma. *Am J Respir Crit Care Med*. 2002;165:1275–9. IIb.
121. Hartley JP, Charles TJ, Monie RD, et al. Arterial plasma histamine after exercise in normal individuals and in patients with exercise-induced asthma. *Clin Sci (Lond)*. 1981;61:151–7. IIa.
122. Anderson SD, Bye PTP, Schoeffel RE, Seale JP, Taylor KM, Ferris L. Arterial plasma histamine levels at rest, during and after exercise in patients with asthma: effects of terbutaline aerosol. *Thorax*. 1981;36:259–67. IIa.
123. Brannan JD, Gulliksson M, Anderson SD, Chew N, Seale JP, Kumlin M. Inhibition of mast cell PGD₂ release protects against mannitol-induced airway narrowing. *Eur Respir J*. 2006;27:944–50. IIa.

124. Leff JA, Busse WW, Pearlman D, et al. Montelukast, a leukotriene receptor antagonist, for the treatment of mild asthma and exercise-induced bronchoconstriction. *N Engl J Med.* 1998;339:147–52. Ib.
125. Anderson SD, Brannan JD. Exercise induced asthma: is there still a case for histamine? *J Allergy Clin Immunol.* 2002;109(5 pt1):771–3. IV.
126. Kanazawa H, Asai K, Hirata K, Yoshikawa J. Vascular involvement in exercise-induced airway narrowing in patients with bronchial asthma. *Chest.* 2002;122:166–70. Iia.
127. Kanazawa H, Hirata K, Yoshikawa J. Involvement of vascular endothelial growth factor in exercise induced bronchoconstriction in asthmatic patients. *Thorax.* 2002;57:885–8. Iia.
128. Kanazawa H, Tochino Y, Asai K. Angiopoietin-2 as a contributing factor of exercise-induced bronchoconstriction in asthmatic patients receiving inhaled corticosteroid therapy. *J Allergy Clin Immunol.* 2008;121:390–5. LB.
129. Edmunds AT, Tooley M, Godfrey S. The refractory period after exercise-induced asthma: its duration and relation to the severity of exercise. *Am Rev Respir Dis.* 1978;117:247–54. Iib.
130. Schoeffel RE, Anderson SD, Gillam I, Lindsay DA. Multiple exercise and histamine challenge in asthmatic patients. *Thorax.* 1980;35:164–70. Iib.
131. Anderson SD, Schoeffel RE. Respiratory heat and water loss during exercise in patients with asthma. Effect of repeated exercise challenge. *Eur J Respir Dis.* 1982;63:472–80. Iib.
132. Larsson J, Perry CP, Brannan JD, Anderson SD, Dahlén SE, Dahlén B. Mast cell mediator release during refractoriness to mannitol induced bronchoconstriction. *Eur Respir J.* 2009;34:1978. Iia.
133. O'Byrne PM, Jones GL. The effect of indomethacin on exercise-induced bronchoconstriction and refractoriness after exercise. *Am Rev Respir Dis.* 1986;134:69–72. Iia.
134. Margolskee DJ, Bigby BG, Boushey HA. Indomethacin blocks airway tolerance to repetitive exercise but not to eucapnic hyperpnea in asthmatic subjects. *Am Rev Respir Dis.* 1988;137:842–6. Iia.
135. Wilson BA, Bar-Or O, Seed LG. Effects of humid air breathing during arm or treadmill exercise on exercise-induced bronchoconstriction and refractoriness. *Am Rev Respir Dis.* 1990;142:349–52. Iib.
136. Wilson BA, Bar-Or O, O'Byrne PM. The effects of indomethacin on refractoriness following exercise both with and without bronchoconstriction. *Eur Respir J.* 1994;7:2174–8. Iia.
137. Torres-Galvan SM, Cumpido JA, Buset N, et al. 5-Lipoxygenase pathway gene polymorphisms: lack of association with asthma in a Spanish population. *J Investig Allergol Clin Immunol.* 2009;19:453–8. Iia.
138. Kang MJ, Lee SY, Kim HB, et al. Association of IL-13 polymorphisms with leukotriene receptor antagonist drug responsiveness in Korean children with exercise-induced bronchoconstriction. *Pharmacogenet Genomics.* 2008;18:551–8. III.
139. Fahy JV. Goblet cell and mucin gene abnormalities in asthma. *Chest.* 2002;122(suppl 6):320S–6S. IV.
140. Hallstrand TS, Wurfel MM, Lai Y, et al. Transglutaminase 2, a novel regulator of eicosanoid production in asthma revealed by genome-wide expression profiling of distinct asthma phenotypes. *PLoS One.* 2010;5:e8583. Iia.
141. Hallstrand TS, Chi EY, Singer AG, Gelb MH, Henderson WR Jr. Secreted phospholipase A2 group X overexpression in asthma and bronchial hyperresponsiveness. *Am J Respir Crit Care Med.* 2007;176:1072–8. Iia.
142. Ciencewicz J, Trivedi S, Kleeberger SR. Oxidants and the pathogenesis of lung diseases. *J Allergy Clin Immunol.* 2008;122:456–68. IV.
143. Barreto M, Villa MP, Olita C, Martella S, Ciabattani G, Montuschi P. 8-Isoprostane in exhaled breath condensate and exercise-induced bronchoconstriction in asthmatic children and adolescents. *Chest.* 2008;135:66–73. Iib.
144. Rundell KW, Slee JB, Caviston R, Hollenbach AM. Decreased lung function after inhalation of ultrafine and fine particulate matter during exercise is related to decreased total nitrate in exhaled breath condensate. *Inhal Toxicol.* 2008;20:1–9. Iia.
145. Hazucha MJ, Folinsee LJ, Bromberg PA. Distribution and reproducibility of spirometric response to ozone by gender and age. *J Appl Physiol.* 2003;95:1917–25. III.
146. Folinsee LJ, Horstman DH, Kehrl HR, Harder S, Abdul-Salaam S, Ives PJ. Respiratory responses to repeated prolonged exposure to 0.12 ppm ozone. *Am J Respir Crit Care Med.* 1994;149:98–105. Iib.
147. Horvath SM, Gliner JA, Folinsee LJ. Adaptation to ozone: duration of effect. *Am Rev Respir Dis.* 1981;123:496–9. Iib.
148. McDonnell WF, Horstman DH, Hazucha MJ, et al. Pulmonary effects of ozone exposure during exercise: dose-response characteristics. *J Appl Physiol.* 1983;54:1345–52. Iib.
149. Ercan H, Birben E, Dizdar EA, et al. Oxidative stress and genetic and epidemiologic determinants of oxidant injury in childhood asthma. *J Allergy Clin Immunol.* 2006;118:1097–104. III.
150. Hayes JD, McLellan LI. Glutathione and glutathione-dependent enzymes represent a co-ordinately regulated defence against oxidative stress. *Free Radic Res.* 1999;31:273–300. LB.
151. Islam T, Berhane K, McConnell R, et al. Glutathione-S-transferase (GST) P1, GSTM1, exercise, ozone and asthma incidence in school children. *Thorax.* 2009;64:197–202. III.
152. Ware JH. Particulate air pollution and mortality--clearing the air. *N Engl J Med.* 2000;343:1798–9. IV.
153. Werz O, Szellas D, Steinhilber D. Reactive oxygen species released from granulocytes stimulate 5-lipoxygenase activity in a Blymphocytic cell line. *Eur J Biochem.* 2000;267:1263–9. LB.
154. Atkinson RW, Anderson HR, Sunyer J, et al. Acute effects of particulate air pollution on respiratory admissions: results from APHEA 2 project. Air Pollution and Health: a European Approach. *Am J Respir Crit Care Med.* 2001;164(10 pt 1):1860–6. IV.
155. Tolbert PE, Mulholland JA, MacIntosh DL, et al. Air quality and pediatric emergency room visits for asthma in Atlanta, Georgia, USA. *Am J Epidemiol.* 2000;151:798–810. III.
156. Hiltermann TJ, Stolk J, van der Zee SC, et al. Asthma severity and susceptibility to air pollution. *Eur Respir J.* 1998;11:686–93. IV.
157. McCreanor J, Cullinan P, Nieuwenhuijsen MJ, et al. Respiratory effects of exposure to diesel traffic in persons with asthma. *N Engl J Med.* 2007;357:2348–58. Iib.
158. Rundell KW. High levels of airborne ultrafine and fine particulate matter in indoor ice arenas. *Inhal Toxicol.* 2003;15:237–50. III.
159. Rundell K. Pulmonary function decay in women ice hockey players: is there a relationship to ice rink air quality? *Inhal Toxicol.* 2004;16:117–23. Iia.
160. Rundell KW, Spiering BA, Evans TM, Baumann JM. Baseline lung function, exercise-induced bronchoconstriction, and asthma-like symptoms in elite women ice hockey players. *Med Sci Sports Exerc.* 2004;36:405–10. III.
161. Rundell KW, Caviston R, Hollenbach AM, Murphy K. Vehicular air pollution, playgrounds, and youth athletic fields. *Inhal Toxicol.* 2006;18:541–7. Iib.
162. Mannix ET, Farber MO, Palange P, Galassetti P, Manfredi F. Exercise-induced asthma in figure skaters. *Chest.* 1996;109:312–5. Iia.
163. Provost-Craig MA, Arbour KS, Sestili DC, Chabalko JJ, Ekinci E. The incidence of exercise-induced bronchospasm in competitive figure skaters. *J Asthma.* 1996;33:67–71. III.
164. Wilber RL, Rundell KW, Szmedra L, Jenkinson DM, Im J, Drake SD. Incidence of exercise-induced bronchospasm in Olympic Winter Sport athletes. *Med Sci Sports Exerc.* 2000;32:732–7. Iib.
165. Anderson SD, Fitch K, Perry CP, et al. Responses to bronchial challenge submitted for approval to use inhaled beta2-agonists before an event at the 2002 Winter Olympics. *J Allergy Clin Immunol.* 2003;111:45–50. III.
166. Rundell KW, Spiering BA, Baumann JM, Evans TM. Bronchoconstriction provoked by exercise in a high-particulate-matter environment is attenuated by montelukast. *Inhal Toxicol.* 2005;17:99–105. Ib.
167. Singh P, Madden M, Gilmour MI. Effects of diesel exhaust particles and carbon black on induction of dust mite allergy in brown Norway rats. *J Immunotoxicol.* 2005;2:41–9. LB.

168. Rabinovitch N, Strand M, Stuhlman K, Gelfand EW. Exposure to tobacco smoke increases leukotriene E4-related albuterol usage and response to montelukast. *J Allergy Clin Immunol.* 2008;121:1365–71. Ib.
169. Anderson SD, Sue-Chu M, Perry CP, et al. Bronchial challenges in athletes applying to inhale a beta2-agonist at the 2004 Summer Olympics. *J Allergy Clin Immunol.* 2006;117:767–73. III.
170. Larsson K, Ohlsén P, Larsson L, Malmberg P, Rydström P-O, Ulriksen H. High prevalence of asthma in cross country skiers. *BMJ.* 1993;307:1326–9. IIa.
171. Helenius IJ, Tikkanen HO, Haahtela T. Occurrence of exercise induced bronchospasm in elite runners: dependence on atopy and exposure to cold air and pollen. *Br J Sports Med.* 1998;32:125–9. III.
172. Davis MS, Lockard AJ, Marlin DJ, Freed AN. Airway cooling and mucosal injury during cold weather exercise. *Equine Vet J Suppl.* 2002;34:413–6. LB.
173. Davis MS, Schofield B, Freed AN. Repeated peripheral airway hyperpnea causes inflammation and remodeling in dogs. *Med Sci Sports Exerc.* 2003;35:608–16. LB.
174. Helenius IJ, Tikkanen HO, Haahtela T. Association between type of training and risk of asthma in elite athletes. *Thorax.* 1997;52:157–60. III.
175. Helenius I, Tikkanen HO, Helenius M, Lumme A, Remes V, Haahtela T. Exercise-induced changes in pulmonary function of healthy, elite long-distance runners in cold air and pollen season exercise challenge tests. *Int J Sports Med.* 2002;23:252–61. III.
176. Anderson SD, Kippelen P. Exercise-induced bronchoconstriction: pathogenesis. *Curr Allergy Asthma Rep.* 2005;5:116–22. IV.
177. Bjermer L, Anderson SD. Bronchial hyperresponsiveness in athletes; mechanisms for development. In: Carlsen K-H, Delgado L, Del Giacco S, eds. *Diagnosis, Prevention and Treatment of Exercise-Related Asthma, Respiratory and Allergic Disorders in Sports.* Wakefield, England: European Respiratory Society; IV.2005–34.
178. Stensrud T, Mykland KV, Gabrielsen K, Carlsen KH. Bronchial hyperresponsiveness in skiers: field test versus methacholine provocation? *Med Sci Sports Exerc.* 2007;39:1681–6. Ib.
179. Freed AN, Omori C, Schofield BH, Mitzner W. Dry air-induced mucosal cell injury and bronchovascular leakage in canine peripheral airways. *Am J Respir Cell Mol Biol.* 1994;11:724–32. LB.
180. Johnson PR, Ammit AJ, Carlin SM, Armour CL, Caughey GH, Black JL. Mast cell tryptase potentiates histamine induced contraction in human sensitized bronchus. *Eur Respir J.* 1996;10:38–43. LB.
181. Anticevich SZ, Hughes JM, Black JL, Armour CL. Induction of human airway hyperresponsiveness by tumor necrosis factor-alpha. *Eur J Pharmacol.* 1995;284:221–5. LB.
182. Anticevich SZ, Hughes JM, Black JL, Armour CL. Induction of hyperresponsiveness in human airway tissue by neutrophils—mechanism of action. *Clin Exp Allergy.* 1996;26:549–56. IV.
183. Ammit AJ, Bekir SS, Johnson PR, Hughes JM, Armour CL, Black JL. Mast cell numbers are increased in the smooth muscle of human sensitized isolated bronchi. *Am J Respir Crit Care Med.* 1997;155:1123–9. LB.
184. Sue-Chu M, Larsson L, Bjermer L. Prevalence of asthma in young cross-country skiers in central Scandinavia: differences between Norway and Sweden. *Respir Med.* 1996;90:99–105. IIb.
185. Leuppi JD, Kuhn M, Comminot C, Reinhart WH. High prevalence of bronchial hyperresponsiveness and asthma in ice hockey players. *Eur Respir J.* 1998;12:13–6. IIb.
186. Sue-Chu M, Brannan JD, Anderson SD, Chew N, Bjermer L. Airway hyperresponsiveness to methacholine, adenosine 5-monophosphate, mannitol, eucapnic voluntary hyperpnea and field exercise challenge in elite cross country skiers. *Brit J Sports Med.* 2010;44:827–32. IIb.
187. Hemingson HB, Davis BE, Cockcroft DW. Seasonal fluctuations in airway responsiveness in elite endurance athletes. *Can Respir J.* 2004; 1:399–401. IIb.
188. Helenius I, Ryttilä P, Sarna S, et al. Effect of continuing or finishing high-level sports on airway inflammation, bronchial hyperresponsiveness, and asthma: a 5-year prospective follow-up study of 42 highly trained swimmers. *J Allergy Clin Immunol.* 2002;109:962–8. IIb.
189. Caillaud C, Le Creff C, Legros P, Denjean A. Strenuous exercise increases plasmatic and urinary leukotriene E4 in cyclists. *Can J Appl Physiol.* 2003;28:793–806. IIa.
190. Bradding P, Walls AF, Holgate ST. The role of the mast cell in the pathophysiology of asthma. *J Allergy Clin Immunol.* 2006;117:1277–84. LB.
191. Greiff L, Wollmer P, Andersson M, Svensson C, Persson CG. Effects of formoterol on histamine induced plasma exudation in induced sputum from normal subjects. *Thorax.* 1998;53:1010–3. IIa.
192. Scola AM, Chong LK, Suvarna SK, Chess-Williams R, Peachell PT. Desensitisation of mast cell β 2-adrenoceptor-mediated responses by salmeterol and formoterol. *Br J Pharmacol.* 2004;141:163–71. LB.
193. Chong LK, Suvarna K, Chess-Williams R, Peachell PT. Desensitization of β 2-adrenoceptor-mediated responses by short-acting β 2-adrenoceptor agonists in human lung mast cells. *Br J Pharmacol.* 2003;138:512–20. LB.
194. Mak JC, Roffel AF, Katsunuma T, Elzinga CR, Zaagsma J, Barnes PJ. Up-regulation of airway smooth muscle histamine H1 receptor mRNA, protein, and function by beta 2-adrenoceptor activation. *Mol Pharmacol.* 2000;57:857–64. LB.
195. McGraw DW, Elwing JM, Fogel KM, et al. Cross talk between Gi and Gq/Gs pathways in airway smooth muscle regulates bronchial contractility and relaxation. *J Clin Invest.* 2007;117:1391–8. LB.
196. Heir T, Aanestad G, Carlsen KH, Larsen S. Respiratory tract infection and bronchial responsiveness in elite athletes and sedentary control subjects. *Scand J Med Sci Sports.* 1995;5:94–9. IIa.
197. Nieman DC, Henson DA, Fagoaga OR, et al. Change in salivary IgA following a competitive marathon race. *Int J Sports Med.* 2002;23:69–75. IIa.
198. Sue-Chu M, Karjalainen EM, Altraja A, et al. Lymphoid aggregates in endobronchial biopsies from young elite cross-country skiers. *Am J Respir Crit Care Med.* 1998;158:597–601. IIa.
199. Delventhal S, Hensel A, Petzoldt K, Pabst R. Effects of microbial stimulation on the number, size and activity of bronchus-associated lymphoid tissue (BALT) structures in the pig. *Int J Exp Pathol.* 1992; 73:351–7. LB.
200. Tschernig T, Pabst R. Bronchus-associated lymphoid tissue (BALT) is not present in the normal adult lung but in different diseases. *Pathobiology.* 2000;68:1–8. LB.
201. Sue-Chu M, Karjalainen E-M, Laitinen A, Larsson L, Laitinen LA, Bjermer L. Placebo-controlled study of inhaled budesonide on indices of airways inflammation in bronchoalveolar lavage fluid and bronchial biopsies in cross country skiers. *Respiration.* 2000;67:417–25. Ib.
202. Helenius I, Lumme A, Ounap J, et al. No effect of montelukast on asthma-like symptoms in elite ice hockey players. *Allergy.* 2004;59:39–44. Ib.
203. Simpson JL, Powell H, Boyle MJ, Scott RJ, Gibson PG. Clarithromycin targets neutrophilic airway inflammation in refractory asthma. *Am J Respir Crit Care Med.* 2008;177:148–55. Ib.
204. Parsons JP, Mastrorade JG. Exercise-induced bronchoconstriction in athletes. *Chest.* 2005;128:3966–74. IV.
205. Bavarian B, Mehrkhani F, Ziaee V, Yousefi A, Nourian R. Sensitivity and specificity of self-reported symptoms for exercise-induced bronchospasm diagnosis in children. *Iran J Pediatr.* 2009;19:47–51. III.
206. Holzer K, Brukner P. Screening of athletes for exercise-induced bronchoconstriction. *Clin J Sport Med.* 2004;14:134–8. IV.
207. National Institutes of Health, Lung and Blood Institute. *Global Initiative for Asthma: Global Strategy for Asthma Management and Prevention.* NHLBI/WHO Workshop Report. Bethesda, MD: Medical Communication Resources; 2007:16–19. <http://www.ginasthma.org>. Accessed October 16, 2010. IV.
208. Sano F, Solé D, Nasipit CK. Prevalence and characteristics of exercise-induced asthma in children. *Pediatr Allergy Immunol.* 1998;9:181–5. IIa.

209. Cabral ALB, Conceição GM, Fonseca-Guedes CHF, Martins MA. Exercise-induced bronchospasm in children: effects of asthma severity. *Am J Respir Crit Care Med*. 1999;159:1819–23. IIB.
210. Burr ML, Eldridge BA, Borysiewicz LK. Peak expiratory flow rates before and after exercise in schoolchildren. *Arch Dis Child*. 1974;49:923–6. IIA.
211. Burr ML, Butland BK, King S, Vaughan-Williams E. Changes in asthma prevalence: two surveys 15 years apart. *Arch Dis Child*. 1989;64:1452–6. IIA.
212. Mannix ET, Roberts M, Fagin DP, Reid B, Farber MO. The prevalence of airways hyperresponsiveness in members of an exercise training facility. *J Asthma*. 2003;40:349–55. IIB.
213. Mannix ET, Roberts MA, Dukes HJ, Magnes CJ, Farber MO. Airways hyperresponsiveness in high school athletes. *J Asthma*. 2004;41:567–74. IIB.
214. Rupp NT, Guill MF, Brudno DS. Unrecognized exercise-induced bronchospasm in adolescent athletes. *Am J Dis Child*. 1992;146:941–4. IIA.
215. Sudhir P, Prasad CE. Prevalence of exercise-induced bronchospasm in schoolchildren: an urban-rural comparison. *J Trop Pediatr*. 2003;49:104–8. IIA.
216. Hallstrand TS, Curtis JR, Koepsell TD, et al. Effectiveness of screening examinations to detect unrecognized exercise-induced bronchoconstriction. *J Pediatr*. 2002;141:343–8. IV.
217. Godfrey S, König P. Exercise-induced bronchial lability in wheezy children and their families. *Pediatrics*. 1975;56(5 pt, suppl 2):851–5. IIA.
218. Daga MK, Ahuja VM, Bajaj SK, et al. Bronchial responsiveness in normal first-degree relatives of asthmatics and patients of allergic rhinitis. *J Indian Acad Clin Med*. 2007;8:36–41. IIA.
219. Sallaoui R, Chamari K, Mossa A, et al. Exercise-induced bronchoconstriction and atopy in Tunisian athletes. *BMC Pulm Med*. 2009;9:8. IIA.
220. Brutsche M, Britschgi D, Dayer E, Tschopp JM. Exercise-induced bronchospasm (EIB) in relation to seasonal and perennial specific IgE in young adults. *Allergy*. 1995;50:905–9. IIA.
221. Gani F, Passalacqua G, Senna G, Mosca Frezet M. Sport, immune system and respiratory infections. *Eur Ann Allergy Clin Immunol*. 2003;35:41–6. IV.
222. Weiler JM, Layton T, Hunt M. Asthma in United States Olympic athletes who participated in the 1996 Summer Games. *J Allergy Clin Immunol*. 1998;102:722–6. III.
223. Weiler JM, Ryan EJ 3rd. Asthma in United States Olympic athletes who participated in the 1998 Olympic Winter Games. *J Allergy Clin Immunol*. 2000;106:267–71. III.
224. Anderson SD, Connolly N, Godfrey S. Comparison of bronchoconstriction induced by cycling and running. *Thorax*. 1971;26:396–401. IIA.
225. Fitch KD, Morton AR. Specificity of exercise in exercise-induced asthma. *Br Med J*. 1971;4:577–81. IIA.
226. Rundell KW, Wilber RL, Szmedra L, Jenkinson DM, Mayers LB, Im J. Exercise-induced asthma screening of elite athletes: field versus laboratory exercise challenge. *Med Sci Sports Exerc*. 2000;32:309–16. III.
227. Sue-Chu M, Larsson L, Moen T, Rennard SI, Bjermer L. Bronchoscopy and bronchoalveolar lavage findings in cross-country skiers with and without “ski asthma.” *Eur Respir J*. 1999;13:626–32. IIA.
228. Pohjantähti H, Laitinen J, Parkkari J. Exercise-induced bronchospasm among healthy elite cross country skiers and non-athletic students. *Scand J Med Sci Sports*. 2005;15:324–8. IIA.
229. Randolph CC, Dreyfus D, Rundell KW, Bangladore D, Fraser B. Prevalence of allergy and asthma symptoms in recreational roadrunners. *Med Sci Sports Exerc*. 2006;38:2053–7. III.
230. Rundell KW, Spiering BA, Judelson DA, Wilson MH. Bronchoconstriction during cross-country skiing: is there really a refractory period? *Med Sci Sports Exerc*. 2003;35:18–26. IV.
231. Helenius I, Haahntela T. Allergy and asthma in elite summer sport athletes. *J Allergy Clin Immunol*. 2000;106:444–52. IV.
232. Bernard A, Nickmilder M, Voisin C, Sardella A. Impact of chlorinated swimming pool attendance on the respiratory health of adolescents. *Pediatrics*. 2009;124:1110–8. IIA.
233. Uçok K, Dane S, Gokbel H, Akar S. Prevalence of exercise-induced bronchospasm in long distance runners trained in cold weather. *Lung*. 2004;182:265–70. IIA.
234. Rupp NT, Brudno S, Guill MF. The value of screening for risk of exercise-induced asthma in high school athletes. *Ann Allergy*. 1993;70:339–42. IIB.
235. Sue-Chu M, Henriksen AH, Bjermer L. Non-invasive evaluation of lower airway inflammation in hyper-responsive elite cross-country skiers and asthmatics. *Respir Med*. 1999;93:719–25. IIA.
236. Castricum A, Holzer K, Brukner P, Irving L. The role of bronchial provocation challenge tests in the diagnosis of exercise-induced bronchoconstriction in elite swimmers. *Br J Sports Med*. 2010;44:736–40. IIB.
237. De Baets F, Bodart E, Dramaix-Wilmet M, et al. Exercise-induced respiratory symptoms are poor predictors of bronchoconstriction. *Pediatr Pulmonol*. 2005;39:301–5. IIB.
238. Schatz M, Clark S, Emond JA, Schreiber D, Camargo CA Jr. Sex differences among children 2–13 years of age presenting at the emergency department with acute asthma. *Pediatr Pulmonol*. 2004;37:523–9. III.
239. Lin RY, Lee GB. The gender disparity in adult asthma hospitalizations dynamically relates to age. *J Asthma*. 2008;45:931–5. III.
240. Bardagi S, Agudo A, Gonzalez CA, Romero PV. Prevalence of exercise-induced airway narrowing in schoolchildren from a Mediterranean town. *Am Rev Respir Dis*. 1993;147:1112–5. IIB.
241. Langdeau JB, Day A, Turcotte H, Boulet LP. Gender differences in the prevalence of airway hyperresponsiveness and asthma in athletes. *Respir Med*. 2009;103:401–6. III.
242. Keeley DJ, Neill P, Gallivan S. Comparison of the prevalence of reversible airways obstruction in rural and urban Zimbabwean children. *Thorax*. 1991;46:549–53. IIA.
243. Kukafka DS, Lang DM, Porter S, et al. Exercise-induced bronchospasm in high school athletes via a free running test: incidence and epidemiology. *Chest*. 1998;114:1613–22. IIA.
244. Jones CO, Qureshi S, Rona RJ, Chinn S. Exercise-induced bronchoconstriction by ethnicity and presence of asthma in British nine year olds. *Thorax*. 1996;51:1134–6. III.
245. Karjalainen EM, Laitinen A, Sue-Chu M, Altraja A, Bjermer L, Laitinen LA. Evidence of airway inflammation and remodeling in ski athletes with and without bronchial hyperresponsiveness to methacholine. *Am J Respir Crit Care Med*. 2000;161:2086–91. IIA.
246. Henriksen AH, Tveit KH, Holmen TL, Sue-Chu M, Bjermer L. A study of the association between exercise-induced wheeze and exercise versus methacholine-induced bronchoconstriction in adolescents. *Pediatr Allergy Immunol*. 2002;13:203–8. III.
247. Brannan JD, Koskela H, Anderson SD, Chew N. Responsiveness to mannitol in asthmatic subjects with exercise- and hyperventilation-induced asthma. *Am J Respir Crit Care Med*. 1998;158:1120–6. III.
248. Evans TM, Rundell KW, Beck KC, Levine AM, Baumann JM. Airway narrowing measured by spirometry and impulse oscillometry following room temperature and cold temperature exercise. *Chest*. 2005;128:2412–9. III.
249. Evans TM, Rundell KW, Beck KC, Levine AM, Baumann JM. Cold air inhalation does not affect the severity of EIB after exercise or eucapnic voluntary hyperventilation. *Med Sci Sports Exerc*. 2005;37:544–9. III.
250. Rundell KW, Spiering BA. Inspiratory stridor in elite athletes. *Chest*. 2003;123:468–74. IIB.
251. Newman KB, Mason UG 3rd, Schmaling KB. Clinical features of vocal cord dysfunction. *Am J Respir Crit Care Med*. 1995;152(4 pt 1):1382–6. IV.
252. Godfrey S, Silverman M, Anderson SD. The use of the treadmill for assessing exercise-induced asthma and the effect of varying the severity and the duration of exercise. *Pediatrics*. 1975;56(5, pt 2 suppl):893–8. IV.

253. Haby MM, Anderson SD, Peat JK, Mellis CM, Toelle BG, Woolcock AJ. An exercise challenge protocol for epidemiological studies of asthma in children: comparison with histamine challenge. *Eur Respir J*. 1994;7:43–9. III.
254. [No authors listed]. Clinical exercise testing with reference to lung diseases: indications, standardization and interpretation strategies. ERS Task Force on Standardization of Clinical Exercise Testing. European Respiratory Society. *Eur Respir J*. 1997;10:2662–89. IV.
255. Anderson SD, Silverman M, Konig P, Godfrey S. Exercise-induced asthma. *Brit J Dis Chest*. 1975;69:1–39. IIa.
256. Eggleston PA, Rosenthal RR, Anderson SA, et al. Guidelines for the methodology of exercise challenge testing of asthmatics. Study Group on Exercise Challenge, Bronchoprovocation Committee, American Academy of Allergy. *J Allergy Clin Immunol*. 1979;64:642–5. IV.
257. König P. Exercise challenge: indications and techniques. *Allergy Proc*. 1989;10:345–8. IV.
258. Mahler DA. Exercise-induced asthma. *Med Sci Sports Exerc*. 1993;25:554–61. IV.
259. Tanaka H, Monahan KD, Seals DR. Age-predicted maximal heart rate revisited. *J Am Coll Cardiol*. 2001;37:153–6. IV.
260. Warren JB, Jennings SJ, Clark TJ. Effect of adrenergic and vagal blockade on the normal human airway response to exercise. *Clin Sci (Lond)*. 1984;66:79–85. IV.
261. Zeitoun M, Wilk B, Matsuzaka A, KnOpfli BH, Wilson BA, Bar-Or O. Facial cooling enhances exercise-induced bronchoconstriction in asthmatic children. *Med Sci Sports Exerc*. 2004;36:767–71. IIb.
262. Koskela H, Tukiainen H. Facial cooling, but not nasal breathing of cold air, induces bronchoconstriction: a study in asthmatic and healthy subjects. *Eur Respir J*. 1995;8:2088–93. IIa.
263. Miyazoe H, Harada Y, Yamasaki S, Tsuji Y. Clinical study on accentuated antagonism in the regulation of heart rate in children. *Jpn Heart J*. 1998;39:481–7. IIb.
264. Ogston J, Butcher JD. A sport-specific protocol for diagnosing exercise-induced asthma in cross-country skiers. *Clin J Sport Med*. 2002;12:291–5. III.
265. Souza AC, Pereira CA. [Bronchial provocation tests using methacholine, cycle ergometer exercise and free running in children with intermittent asthma.] [Article in Portuguese] *J Pediatr (Rio J)*. 2005;81:65–72. IV.
266. Stensrud T, Berntsen S, Carlsen KH. Humidity influences exercise capacity in subjects with exercise-induced bronchoconstriction (EIB). *Respir Med*. 2006;100:1633–41. III.
267. Spiering BA, Judelson DA, Rundell KW. An evaluation of standardizing target ventilation for eucapnic voluntary hyperventilation using FEV₁. *J Asthma*. 2004;41:745–9. III.
268. Silverman M, Anderson SD. Metabolic cost of treadmill exercise in children. *J Appl Physiol*. 1972;33:696–8. IV.
269. Givoni B, Goldman RF. Predicting metabolic energy cost. *J Appl Physiol*. 1971;30:429–33. IV.
270. Jones NL. *Clinical Exercise Testing*. 4th ed. Philadelphia, PA: WB Saunders Company; 1997. IV.
271. Anderson SD, Lambert S, Brannan JD, et al. Laboratory protocol for exercise asthma to evaluate salbutamol given by two devices. *Med Sci Sports Exerc*. 2001;33:893–900. IV.
272. Powers SK, Martin D, Cicale M, Collop N, Huang D, Criswell D. Exercise-induced hypoxemia in athletes: role of inadequate hyperventilation. *Eur J Appl Physiol Occup Physiol*. 1992;65:37–42. IV.
273. Roach JM, Hurwitz KM, Argyros GJ, Eliasson AH, Phillips YY. Eucapnic voluntary hyperventilation as a bronchoprovocation technique. Comparison with methacholine inhalation in asthmatics. *Chest*. 1994;105:667–72. IV.
274. Anderson SD, Argyros GJ, Magnussen H, Holzer K. Provocation by eucapnic voluntary hyperpnea to identify exercise induced bronchoconstriction. *Br J Sports Med*. 2001;35:344–7. III.
275. Sterling GM. The mechanism of bronchoconstriction due to hypocapnia in man. *Clin Sci*. 1968;34:277–85. IV.
276. Suman OE, Beck KC, Babcock MA, Pegelow DF, Reddan AW. Airway obstruction during exercise and isocapnic hyperventilation in asthmatic subjects. *J Appl Physiol*. 1999;87:1107–13. IV.
277. Argyros GJ, Roach JM, Hurwitz KM, Eliasson AH, Phillips YY. The refractory period after eucapnic voluntary hyperventilation challenge and its effect on challenge technique. *Chest*. 1995;108:419–24. IV.
278. Finnerty JP, Wilmont C, Holgate ST. Inhibition of hypertonic saline-induced bronchoconstriction by terfenadine and flurbiprofen. Evidence for the predominant role of histamine. *Am Rev Respir Dis*. 1989;140:593–7. III.
279. Riedler J, Reade T, Dalton M, Holst D, Robertson C. Hypertonic saline challenge in an epidemiologic survey of asthma in children. *Am J Respir Crit Care Med*. 1994;150:1632–9. III.
280. Rabone SJ, Phoon WO, Anderson SD, et al. Hypertonic saline challenge in an adult epidemiological survey. *Occup Med. (Lond)* 1996;46:177–85. III.
281. Smith CM, Anderson SD. A comparison between the airway response to isocapnic hyperventilation and hypertonic saline in subjects with asthma. *Eur Respir J*. 1989;2:36–43. IIb.
282. Belcher NG, Murdoch RD, Dalton N, et al. A comparison of mediator and catecholamine release between exercise- and hypertonic saline-induced asthma. *Am Rev Respir Dis*. 1988;137:1026–32. IIb.
283. Anderson SD, Brannan JD, Chan HK. Use of aerosols for bronchial provocation testing in the laboratory: where we have been and where we are going. *J Aerosol Med*. 2002;15:313–24. IV.
284. Anderson SD, Brannan JD. Methods for “indirect” challenge tests including exercise, eucapnic voluntary hyperpnea and hypertonic aerosols. *Clin Rev Allergy Immunol*. 2003;24:27–54. IV.
285. Jones SL, Herbison P, Cowan JO, et al. Exhaled NO and assessment of anti-inflammatory effects of inhaled steroid: dose-response relationship. *Eur Respir J*. 2002;20:601–8. III.
286. Riedler J, Gamper A, Eder W, Oberfeld G. Prevalence of bronchial hyperresponsiveness to 4.5% saline and its relation to asthma and allergy symptoms in Austrian children. *Eur Respir J*. 1998;11:355–60. III.
287. Brannan JD, Anderson SD, Perry CP, Freed-Martens R, Lässig AR, Charlton B. The safety and efficacy of inhaled dry powder mannitol as a bronchial provocation test for airway hyperresponsiveness: a phase 3 comparison study with hypertonic (4.5%) saline. *Respir Res*. 2005;6:144. IIb.
288. Anderson SD, Brannan J, Trevillion L, Young IH. Lung function and bronchial provocation testing for intending divers with a history of asthma. *SPUMS J*. 1995;25:233–48. IV.
289. Anderson SD. Challenge tests to assess airway hyperresponsiveness and efficacy of drugs used in the treatment of asthma. *J Aerosol Med*. 1996;9:95–109. IV.
290. Anderson SD, Brannan J, Spring J, et al. A new method for bronchial provocation testing in asthmatic subjects using a dry powder of mannitol. *Am J Respir Crit Care Med*. 1997;156:758–65. III.
291. Muñoz PA, Gómez FP, Manrique HA, et al. Pulmonary gas exchange response to exercise- and mannitol-induced bronchoconstriction in mild asthma. *J Appl Physiol*. 2008;105:1477–85. IV.
292. Holzer K, Anderson SD, Chan HK, Douglass J. Mannitol as a challenge test to identify exercise-induced bronchoconstriction in elite athletes. *Am J Respir Crit Care Med*. 2003;167:534–7. IV.
293. Cowan DC, Hewitt RS, Cowan JO, et al. Exercise-induced wheeze: fraction of exhaled nitric oxide-directed management. *Respirology*. 2010;15:683–90. IV.
294. Brannan JD, Koskela H, Anderson SD, Chan HK. Budesonide reduces sensitivity and reactivity to inhaled mannitol in asthmatic subjects. *Respirology*. 2002;7:37–44. IIb.
295. International Olympic Medical Committee. β_2 -adrenoceptor agonists and the Olympic Games in Beijing. 2008. Available at http://multimedia.olympic.org/pdf/en_report_1302.pdf. Accessed October 16, 2010. IV.
296. Dickinson JW, Whyte GP, McConnell AK, Harries MG. Screening elite winter athletes for exercise induced asthma: a comparison of three challenge methods. *Br J Sports Med*. 2006;40:179–82. IV.

297. Christopher KL, Wood RP 2nd, Eckert RC, Blager FB, Raney RA, Souhrada JF. Vocal-cord dysfunction presenting as asthma. *N Engl J Med*. 1983;308:1566–70. Iib.
298. Björnisdóttir US, Gudmundsson K, Hjartarson H, Bröndbo K, Magnússon B, Juliusson S. Exercise-induced laryngomalacia: an imitator of exercise-induced bronchospasm. *Ann Allergy Asthma Immunol*. 2000; 85:387–91. III.
299. Chemery L, Le Clech G, Delaval P, Carré F, Gogibu J, Dassonville J. [Exercise-induced laryngomalacia] [Article in French]. *Rev Mal Respir*. 2002;19(5 pt 1):641–3. III.
300. Bittleman DB, Smith RJ, Weiler JM. Abnormal movement of the arytenoid region during exercise presenting as exercise-induced asthma in an adolescent athlete. *Chest*. 1994;106:615–6. III.
301. McFadden ER Jr, Zawadski DK. Vocal cord dysfunction masquerading as exercise-induced asthma. A physiologic cause for “choking” during athletic activities. *Am J Respir Crit Care Med*. 1996;153:942–7. Iib.
302. Morris MJ, Deal LE, Bean DR, Grbach VX, Morgan JA. Vocal cord dysfunction in patients with exertional dyspnea. *Chest*. 1999;116: 1676–82. Iia.
303. Perkins PJ, Morris MJ. Vocal cord dysfunction induced by methacholine challenge testing. *Chest*. 2002;122:1988–93. Iia.
304. Davis RS, Brugman SM, Larsen GL. Use of videography in the diagnosis of exercise-induced vocal cord dysfunction: a case report with video clips. *J Allergy Clin Immunol*. 2007;119:1329–31. III.
305. Heimdal JH, Roksund OD, Halvorsen T, Skadberg BT, Olofsson J. Continuous laryngoscopy exercise test: a method for visualizing laryngeal dysfunction during exercise. *Laryngoscope*. 2006;116:52–7. III.
306. Fahey JT, Bryant NJ, Karas D, Goldberg B, Destefano R, Gracco LC. Exercise-induced stridor due to abnormal movement of the arytenoids area: videoendoscopic diagnosis and characterization of the “at risk” group. *Pediatr Pulmonol*. 2005;39:51–5. III.
307. Richter GT, Rutter MJ, deAlarcon A, Orvidas LJ, Thompson DM. Late-onset laryngomalacia: a variant of disease. *Arch Otolaryngol Head Neck Surg*. 2008;134:75–80. Iib.
308. Bent JP 3rd, Miller DA, Kim JW, Bauman NM, Wilson JS, Smith RJ. Pediatric exercise-induced laryngomalacia. *Ann Otol Rhinol Laryngol*. 1996;105:169–75. III.
309. Mandell DL, Arjmand EM. Laryngomalacia induced by exercise in a pediatric patient. *Int J Pediatr Otorhinolaryngol*. 2003;67:999–1003. III.
310. Smith RJ, Bauman NM, Bent JP, Kramer M, Smits WL, Ahrens RC. Exercise-induced laryngomalacia. *Ann Otol Rhinol Laryngol*. 1995; 104:537–41. IV.
311. Lakin RC, Metzger WJ, Haughey BH. Upper airway obstruction presenting as exercise-induced asthma. *Chest*. 1984;86:499–501. III.
312. Gessler EM, Simko EJ, Greinwald JH Jr. Adult laryngomalacia: an uncommon clinical entity. *Am J Otolaryngol*. 2002;23:386–9. III.
313. Patel NJ, Jorgensen C, Kuhn J, Merati AL. Concurrent laryngeal abnormalities in patients with paradoxical vocal fold dysfunction. *Otolaryngol Head Neck Surg*. 2004;130:686–9. Iib.
314. Powell DM, Karanfilov BI, Beechler KB, Treole K, Trudeau MD, Forrest LA. Paradoxical vocal cord dysfunction in juveniles. *Arch Otolaryngol Head Neck Surg*. 2000;126:29–34. Iib.
315. Ford CN. Evaluation and management of laryngopharyngeal reflux. *JAMA*. 2005;294:1534–40. IV.
316. Hammo AH, Weinberger MM. Exercise-induced hyperventilation: a pseudoasthma syndrome. *Ann Allergy Asthma Immunol*. 1999;82: 574–8. III.
317. Weinberger MM. Etiology of exercise-induced dyspnea: not just exercise-induced asthma or vocal cord dysfunction. *J Allergy Clin Immunol*. 2008;121:269. IV.
318. Weinberger M, Abu-Hasan M. Pseudo-asthma: when cough, wheezing, and dyspnea are not asthma. *Pediatrics*. 2007;120:855–64. III.
319. Weinberger M. Exercise induced dyspnoea: if not asthma, then what? *Arch Dis Child*. 2006;91:543–4. III.
320. Meyer T, Faude O, Scharhag J, Urhausen A, Kindermann W. Is lactic acidosis a cause of exercise induced hyperventilation at the respiratory compensation point? *Br J Sports Med*. 2004;38:622–5. III.
321. White KM, Quinn JM, Hagan LL, Johnson TL 2nd. Exercise-induced hyperventilation. *Ann Allergy Asthma Immunol*. 2008;100:171–2. III.
322. Jack S, Rossiter HB, Pearson MG, Ward SA, Warburton CJ, Whipp BJ. Ventilatory responses to inhaled carbon dioxide, hypoxia, and exercise in idiopathic hyperventilation. *Am J Respir Crit Care Med*. 2004; 170:118–25. Iia.
323. Lopes WA, Radominski RB, Rosário Filho NA, Leite N. Exercise induced bronchospasm in obese adolescents. *Allergol Immunopathol (Madr)*. 2009;37:175–9. Iia.
324. Fonkalsrud EW, DeUgarte D, Choi E. Repair of pectus excavatum and carinatum deformities in 116 adults. *Ann Surg*. 2002;236:304–12. III.
325. Koumbourlis AC, Stolar CJ. Lung growth and function in children and adolescents with idiopathic pectus excavatum. *Pediatr Pulmonol*. 2004; 38:339–43. III.
326. Smyth RJ, Chapman KR, Wright TA, Crawford JS, Rebuck AS. Ventilatory patterns during hypoxia, hypercapnia, and exercise in adolescents with mild scoliosis. *Pediatrics*. 1986;77:692–7. III.
327. Ben-Dov I, Kaminski N, Reichert N, Rosenman J, Shulimzon T. Diaphragmatic paralysis: a clinical imitator of cardiorespiratory diseases. *Isr Med Assoc J*. 2008;10(8–9):579–83. III.
328. Marciniuk DD, Sridhar G, Clemens RE, Zintel TA, Gallagher CG. Lung volumes and expiratory flow limitation during exercise in interstitial lung disease. *J Appl Physiol*. 1994;77:963–73. III.
329. Harris-Eze AO, Sridhar G, Clemens RE, Zintel TA, Gallagher CG, Marciniuk DD. Role of hypoxemia and pulmonary mechanics in exercise limitation in interstitial lung disease. *Am J Respir Crit Care Med*. 1996;154(4 pt 1):994–1001. III.
330. Sheffer AL, Austen KF. Exercise-induced anaphylaxis. *J Allergy Clin Immunol*. 1980;66:106–11. III.
331. Sheffer AL, Austen KF. Exercise-induced anaphylaxis. *J Allergy Clin Immunol*. 1984;73(5 pt 2):699–703. IV.
332. Maulitz RM, Pratt DS, Schocket AL. Exercise-induced anaphylactic reaction to shellfish. *J Allergy Clin Immunol*. 1979;63:433–4. III.
333. Kidd JM 3rd, Cohen SH, Sosman AJ, Fink JN. Food-dependent exercise-induced anaphylaxis. *J Allergy Clin Immunol*. 1983;71:407–11. III.
334. Kivity S, Sneh E, Greif J, Topilsky M, Mekori YA. The effect of food and exercise on the skin response to compound 48/80 in patients with food-associated exercise-induced urticaria-angioedema. *J Allergy Clin Immunol*. 1988;81:1155–8. III.
335. Schwartz HJ. Elevated serum tryptase in exercise-induced anaphylaxis. *J Allergy Clin Immunol*. 1995;95:917–9. III.
336. Du Toit G. Food-dependent exercise-induced anaphylaxis in childhood. *Pediatr Allergy Immunol*. 2007;18:455–63. IV.
337. Palosuo K, Alenius H, Varjonen E, et al. A novel wheat gliadin as a cause of exercise-induced anaphylaxis. *J Allergy Clin Immunol*. 1999; 103(5 pt 1):912–7. Iib.
338. Palosuo K, Varjonen E, Nurkka J, et al. Transglutaminase-mediated cross-linking of a peptic fraction of omega-5 gliadin enhances IgE reactivity in wheat-dependent, exercise-induced anaphylaxis. *J Allergy Clin Immunol*. 2003;111:1386–92. Iib.
339. Matsuo H, Kohno K, Niihara H, Morita E. Specific IgE determination to epitope peptides of omega-5 gliadin and high molecular weight glutenin subunit is a useful tool for diagnosis of wheat-dependent exercise-induced anaphylaxis. *J Immunol*. 2005;175:8116–22. Iib.
340. Matsuo H, Dahlström J, Tanaka A, et al. Sensitivity and specificity of recombinant omega-5 gliadin-specific IgE measurement for the diagnosis of wheat-dependent exercise-induced anaphylaxis. *Allergy*. 2008; 63:233–6. Iia.
341. Oyefara BI, Bahna SL. Delayed food-dependent, exercise-induced anaphylaxis. *Allergy Asthma Proc*. 2007;28:64–6. III.
342. Meyer FJ, Ewert R, Hoepfer MM, et al. Peripheral airway obstruction in primary pulmonary hypertension. *Thorax*. 2002;57:473–6. Iia.
343. D’Alonzo GE, Gianotti LA, Pohil RL, et al. Comparison of progressive exercise performance of normal subjects and patients with primary pulmonary hypertension. *Chest*. 1987;92:57–62. Iia.

344. Rastogi D, Ngai P, Barst RJ, Koumbourlis AC. Lower airway obstruction, bronchial hyperresponsiveness, and primary pulmonary hypertension in children. *Pediatr Pulmonol.* 2004;37:50–5. Iib.
345. Achouh L, Montani D, Garcia G, et al. Pulmonary arterial hypertension masquerading as severe refractory asthma. *Eur Respir J.* 2008;32:513–6. III.
346. Reindl I, Kleber FX. Exertional hyperpnea in patients with chronic heart failure is a reversible cause of exercise intolerance. *Basic Res Cardiol.* 1996;91(suppl 1):37–43. Iib.
347. Maron BJ. Hypertrophic cardiomyopathy: a systematic review. *JAMA.* 2002;287:1308–20. IV.
348. Frank MJ, Abdulla AM, Watkins LO, Prisant L, Stefadouros MA. Long-term medical management of hypertrophic cardiomyopathy: usefulness of propranolol. *Eur Heart J.* 1983;4(suppl F):155–64. Iib.
349. Besley DC, McWilliams GJ, Moodie DS, Castle LW. Long-term follow-up of young adults following permanent pacemaker placement for complete heart block. *Am Heart J.* 1982;103:332–7. III.
350. Parker JM, Cary-Freitas B, Berg BW. Symptomatic vascular rings in adulthood: an uncommon mimic of asthma. *J Asthma.* 2000;37:275–80. III.
351. Chuang ML, Chang HC, Lim KE, Vintch JR. Gas exchange detection of right-to-left shunt in dyspneic patients: report of three cases. *Int J Cardiol.* 2006;108:117–9. III.
352. Gossage JR, Kanj G. Pulmonary arteriovenous malformations. A state of the art review. *Am J Respir Crit Care Med.* 1998;158:643–61. IV.
353. Ofir D, Laveneziana P, Webb KA, Lam YM, O'Donnell DE. Mechanisms of dyspnea during cycle exercise in symptomatic patients with GOLD stage I chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2008;177:622–9. Iia.
354. Dempsey JA, McKenzie DC, Haverkamp HC, Eldridge MW. Update in the understanding of respiratory limitations to exercise performance in fit, active adults. *Chest.* 2008;134:613–22. IV.
355. Ferrari M, Bonella F, Benini L, et al. Acid reflux into the oesophagus does not influence exercise-induced airway narrowing in bronchial asthma. *Br J Sports Med.* 2008;42:845–9. Iib.
356. Weiner P, Konson N, Sternberg A, Zamir D, Fireman Z. Is gastroesophageal reflux a factor in exercise-induced asthma? *Respir Med.* 1998;92:1071–5. Iia.
357. Wright RA, Sagatelian MA, Simons ME, McClave SA, Roy TM. Exercise-induced asthma. Is gastroesophageal reflux a factor? *Dig Dis Sci.* 1996;41:921–5. Iib.
358. Peterson KA, Samuelson WM, Ryujin DT, et al. The role of gastroesophageal reflux in exercise-triggered asthma: a randomized controlled trial. *Dig Dis Sci.* 2009;54:564–71. Ib.
359. Peters HP, De Kort AF, Van Krevelen H, et al. The effect of omeprazole on gastro-oesophageal reflux and symptoms during strenuous exercise. *Aliment Pharmacol Ther.* 1999;13:1015–22. Ib.
360. Haden JR, Khan DA. Psychiatric syndromes that mimic asthma. *Adv Psychosom Med.* 2003;24:72–85. IV.
361. Ringsberg KC, Akerlind I. Presence of hyperventilation in patients with asthma-like symptoms but negative asthma test responses: provocation with voluntary hyperventilation and mental stress. *J Allergy Clin Immunol.* 1999;103:601–8. Iia.
362. Babb TG, Ranasinghe KG, Comeau LA, Semon TL, Schwartz B. Dyspnea on exertion in obese women: association with an increased oxygen cost of breathing. *Am J Respir Crit Care Med.* 2008;178:116–23. Iib.
363. Davies CT, Godfrey S, Light M, Sargeant AJ, Zeidifard E. Cardiopulmonary responses to exercise in obese girls and young women. *J Appl Physiol.* 1975;38:373–6. III.
364. Haller RG, Lewis SF, Estabrook RW, DiMauro S, Servidei S, Foster DW. Exercise intolerance, lactic acidosis, and abnormal cardiopulmonary regulation in exercise associated with adult skeletal muscle cytochrome c oxidase deficiency. *J Clin Invest.* 1989;84:155–61. III.
365. Hooper RG, Thomas AR, Kearl RA. Mitochondrial enzyme deficiency causing exercise limitation in normal-appearing adults. *Chest.* 1995;107:317–22. III.
366. Flaherty KR, Wald J, Weisman IM, et al. Unexplained exertional limitation: characterization of patients with a mitochondrial myopathy. *Am J Respir Crit Care Med.* 2001;164:425–32. Iia.
367. Rossing TH, Weiss JW, Breslin FJ, Ingram RH Jr, McFadden ER Jr. Effects of inhaled sympathomimetics on obstructive response to respiratory heat loss. *J Appl Physiol.* 1982;52:1119–23. Ib.
368. Latimer KM, O'Byrne PM, Morris MM, Roberts R, Hargreave FE. Bronchoconstriction stimulated by airway cooling. Better protection with combined inhalation of terbutaline sulphate and cromolyn sodium than with either alone. *Am Rev Respir Dis.* 1983;128:440–3. Ib.
369. Food and Drug Association: Center for Drug Evaluation and Research Draft Guidance for Industry. Exercise-induced bronchospasm (EIB). Development of drugs to prevent EIB. Draft Guidance. Washington, DC: US Dept of Health and Human Services; February 2002. IV.
370. Kemp JP, Dockhorn RJ, Shapiro GG, et al. Montelukast once daily inhibits exercise-induced bronchoconstriction in 6- to 14-year-old children with asthma. *J Pediatr.* 1998;133:424–8. Ib.
371. Anderson S, Seale JP, Ferris L, Schoeffel R, Lindsay DA. An evaluation of pharmacotherapy for exercise-induced asthma. *J Allergy Clin Immunol.* 1979;64(6 pt 2):612–24. IV.
372. Godfrey S, König P. Suppression of exercise-induced asthma by salbutamol, theophylline, atropine, cromolyn, and placebo in a group of asthmatic children. *Pediatrics.* 1975;56:930–4. Ib.
373. Godfrey S, König P. Inhibition of exercise-induced asthma by different pharmacological pathways. *Thorax.* 1976;31:137–43. Ib.
374. Hofstra WB, Sont JK, Sterk PJ, Neijens HJ, Kuethe MC, Duiverman EJ. Sample size estimation in studies monitoring exercise-induced bronchoconstriction in asthmatic children. *Thorax.* 1997;52:739–41. Iia.
375. Clinical exercise testing with reference to lung diseases: indications, standardization and interpretation strategies. ERS Task Force on Standardization of Clinical Exercise Testing. European Respiratory Society. *Eur Respir J.* 1997 Nov;10:2662–89. IV.
376. Anderson SD, Seale JP, Rozea P, Bandler L, Theobald G, Lindsay DA. Inhaled and oral salbutamol in exercise-induced asthma. *Am Rev Respir Dis.* 1976;114:493–500. Iia.
377. McFadden ER Jr, Gilbert IA. Exercise-induced asthma. *N Engl J Med.* 1994;330:1362–7. IV.
378. Hendrickson CD, Lynch JM, Gleeson K. Exercise induced asthma: a clinical perspective. *Lung.* 1994;172:1–14. IV.
379. Moraes TJ, Selvadurai H. Management of exercise-induced bronchospasm in children: the role of leukotriene antagonists. *Treat Respir Med.* 2004;3:9–15. IV.
380. Tan RA, Spector SL. Exercise-induced asthma: diagnosis and management. *Ann Allergy Asthma Immunol.* 2002;89:226–35. IV.
381. Kelly HW. What is new in the prevention of exercise-induced bronchospasm (EIB) in children? *Pediatr Asthma Allergy Immunol.* 2008;21:40–3. IV.
382. Anderson SD, Caillaud C, Brannan JD. β_2 -agonists and exercise-induced asthma. *Clin Rev Allergy Immunol.* 2006;31(2–3):163–80. IV.
383. Peachell P. Regulation of mast cells by β_2 -agonists. *Clin Rev Allergy Immunol.* 2006;31(2–3):131–42. IV.
384. Schoeffel RE, Anderson SD, Lindsay DA. Sodium cromoglycate as a pressurized aerosol (vicrom) in exercise-induced asthma. *Aust N Z J Med.* 1983;13:157–61. Iia.
385. Anderson SD, Rodwell LT, Du Toit J, Young IH. Duration of protection by inhaled salmeterol in exercise-induced asthma. *Chest.* 1991;100:1254–60. Ib.
386. Schoeffel RE, Anderson SD, Seale JP. The protective effect and duration of action of metaproteronol aerosol on exercise-induced asthma. *Ann Allergy.* 1981;46:273–5. Ib.
387. Aebischer JC, Benoit RC, Scherrer M. [Pirbuterol and salbutamol aerosol for exercise-induced bronchoconstriction] [Article in German]. *Schweiz Med Wochenschr.* 1984;114:1660–4. Ib.
388. Pearlman DS, Rees W, Schaefer K, Huang H, Andrews WT. An evaluation of levalbuterol HFA in the prevention of exercise-induced bronchospasm. *J Asthma.* 2007;44:729–33. Ib.

389. Ferrari M, Balestreri F, Baratieri S, Biasin C, Oldani V, Lo Cascio V. Evidence of the rapid protective effect of formoterol dry-powder inhalation against exercise-induced bronchospasm in athletes with asthma. *Respiration*. 2000;67:510–3. Ib.
390. Ferrari M, Segattini C, Zanon R, et al. Comparison of the protective effect of formoterol and of salmeterol against exercise-induced bronchospasm when given immediately before a cycloergometric test. *Respiration*. 2002; 69:509–12. Ib.
391. Bisgaard H. Long-acting beta2-agonists in management of childhood asthma: a critical review of the literature. *Pediatr Pulmonol*. 2000; 29:221–34. IV.
392. Kemp JP, Dockhorn RJ, Busse WW, Bleecker ER, Van As A. Prolonged effect of inhaled salmeterol against exercise-induced bronchospasm. *Am J Respir Crit Care Med*. 1994;150:1612–5. Ib.
393. Nelson JA, Strauss L, Skowronski M, Ciuffo R, Novak R, McFadden ER Jr. Effect of long-term salmeterol treatment on exercise-induced asthma. *N Engl J Med*. 1998;339:141–6. Ib.
394. Carlsen KH, Røksund O, Olsholt K, Njå F, Leegaard J, Bratten G. Overnight protection by inhaled salmeterol on exercise-induced asthma in children. *Eur Respir J*. 1995;8:1852–5. Ib.
395. Newnham DM, Ingram CG, Earnshaw J, Palmer JB, Dhillon DP. Salmeterol provides prolonged protection against exercise-induced bronchoconstriction in a majority of subjects with mild, stable asthma. *Respir Med*. 1993;87:439–44. Ib.
396. Boner AL, Spezia E, Piovesan P, Chiocca E, Maiocchi G. Inhaled formoterol in the prevention of exercise-induced bronchoconstriction in asthmatic children. *Am J Respir Crit Care Med*. 1994;149:935–9. Ib.
397. Vilsvik J, Ankerst J, Palmqvist M, et al. Protection against cold air and exercise-induced bronchoconstriction while on regular treatment with Oxis. *Respir Med*. 2001;95:484–90. Ib.
398. Bronsky EA, Yegen Ü, Yeh CM, Larsen LV, Della Cioppa G. Formoterol provides long lasting protection against exercise-induced bronchospasm. *Ann Allergy Asthma Immunol*. 2002; 89:407–12. Ib.
399. Villaran C, O'Neill SJ, Helbling A, et al. Montelukast versus salmeterol in patients with asthma and exercise-induced bronchoconstriction. Montelukast/Salmeterol Exercise Study Group. *J Allergy Clin Immunol*. 1999;104(3 Part 1):547–53. Ib.
400. Edelman JM, Turpin JA, Bronsky EA, et al. Oral montelukast compared with inhaled salmeterol to prevent exercise-induced bronchoconstriction. A randomized, double-blind trial. Exercise Study Group. *Ann Intern Med*. 2000;132:97–104. Ib.
401. Hancox RJ, Subbarao P, Kamada D, Watson RM, Hargreave FE, Inman MD. Beta2-agonist tolerance and exercise-induced bronchospasm. *Am J Respir Crit Care Med*. 2002;165:1068–70. Ib.
402. Inman MD, O'Byrne PM. The effect of regular inhaled albuterol on exercise-induced bronchoconstriction. *Am J Respir Crit Care Med*. 1996;153:65–9. Ib.
403. Wraight JM, Hancox RJ, Herbison GP, Cowan JO, Flannery EM, Taylor DR. Bronchodilator tolerance: the impact of increasing bronchoconstriction. *Eur Respir J*. 2003;21:810–5. Ib.
404. Hancox RJ, Aldridge RE, Cowan JO, et al. Tolerance to beta-agonists during acute bronchoconstriction. *Eur Respir J*. 1999;14:283–7. Ib.
405. Hancox RJ, Cowan JO, Flannery EM, Herbison GP, McLachlan CR, Taylor DR. Bronchodilator tolerance and rebound bronchoconstriction during regular inhaled beta-agonist treatment. *Respir Med*. 2000;94: 767–71. Ib.
406. Haney S, Hancox RJ. Overcoming beta-agonist tolerance: high dose salbutamol and ipratropium bromide. Two randomised controlled trials. *Respir Res*. 2007;8:19. Ib.
407. Aziz I, Tan KS, Hall IP, Devlin MM, Lipworth BJ. Subsensitivity to bronchoprotection against adenosine monophosphate challenge following regular once-daily formoterol. *Eur Respir J*. 1998;12:580–4. Ib.
408. Drotar DE, Davis EE, Cockcroft DW. Tolerance to the bronchoprotective effect of salmeterol 12 hours after starting twice daily treatment. *Ann Allergy Asthma Immunol*. 1998;80:31–4. Ib.
409. Bhagat R, Kalra S, Swystun VA, Cockcroft DW. Rapid onset of tolerance to the bronchoprotective effect of salmeterol. *Chest*. 1995; 108:1235–9. Ib.
410. Davis BE, Reid JK, Cockcroft DW. Formoterol thrice weekly does not result in the development of tolerance to bronchoprotection. *Can Respir J*. 2003;10:23–6. Ib.
411. Giannini D, Carletti A, Dente FL, et al. Tolerance to the protective effect of salmeterol on allergen challenge. *Chest*. 1996;110:1452–7. Ib.
412. Storms W, Chervinsky P, Ghannam AF, Bird S, Hustad CM, Edelman JM. A comparison of the effects of oral montelukast and inhaled salmeterol on response to rescue bronchodilation after challenge. *Respir Med*. 2004;98:1051–62. Ib.
413. Garcia R, Guerra P, Feo F, et al. Tachyphylaxis following regular use of formoterol in exercise-induced bronchospasm. *J Investig Allergol Clin Immunol*. 2001;11:176–82. Ib.
414. Johnson M. Molecular mechanisms of β_2 -adrenergic receptor function, response, and regulation. *J Allergy Clin Immunol*. 2006;117:18–24. IV.
415. Hayes MJ, Qing F, Rhodes CG, et al. In vivo quantification of human pulmonary beta-adrenoceptors: effect of beta-agonist therapy. *Am J Respir Crit Care Med*. 1996;154:1277–83. III.
416. McGraw DW, Liggett SB. Heterogeneity in beta-adrenergic receptor kinase expression in the lung accounts for cell-specific desensitization of the beta2-adrenergic receptor. *J Biol Chem*. 1997;272:7338–44. IIa.
417. O'Connor BJ, Aikman SL, Barnes PJ. Tolerance to the nonbronchodilator effects of inhaled beta 2-agonists in asthma. *N Engl J Med*. 1992;327:1204–8. Ib.
418. Swystun VA, Gordon JR, Davis EB, Zhang X, Cockcroft DW. Mast cell tryptase release and asthmatic responses to allergen increase with regular use of salbutamol. *J Allergy Clin Immunol*. 2000;106:57–64. Ia.
419. Adler A, Uziel Y, Mei-Zahav M, Horowitz I. Formoterol induces tolerance to the bronchodilating effect of salbutamol following methacholine-provocation test in asthmatic children. *Pulm Pharmacol Ther*. 2006;19:281–5. Ia.
420. Anderson GP. Current issues with β_2 -adrenoceptor agonists: pharmacology and molecular and cellular mechanisms. *Clin Rev Allergy Immunol*. 2006;31(2–3):119–30. IV.
421. Finnerty JP, Holgate ST. The contribution of histamine release and vagal reflexes, alone and in combination, to exercise-induced asthma. *Eur Respir J*. 1993;6:1132–7. Ib.
422. Knöpfli BH, Bar-Or O, Araújo CG. Effect of ipratropium bromide on EIB in children depends on vagal activity. *Med Sci Sports Exerc*. 2005;37:354–9. IIa.
423. Sarria B, Naline E, Zhang Y, et al. Muscarinic M2 receptors in acetylcholine-isoproterenol functional antagonism in human isolated bronchus. *Am J Physiol Lung Cell Mol Physiol*. 2002;283:L1125–32. IIa.
424. Taylor DR. The beta-agonist saga and its clinical relevance: on and on it goes. *Am J Respir Crit Care Med*. 2009;179:976–8. IV.
425. Hanania NA, Singh S, El-Wali R, et al. The safety and effects of the beta-blocker, nadolol, in mild asthma: an open-label pilot study. *Pulm Pharmacol Ther*. 2008;21:134–41. IIa.
426. Kalra S, Swystun VA, Bhagat R, Cockcroft DW. Inhaled corticosteroids do not prevent the development of tolerance to the bronchoprotective effect of salmeterol. *Chest*. 1996;109:953–6. Ib.
427. Food and Drug Association (FDA). FDA Drug Safety Communication: New safety requirements for long-acting inhaled asthma medications called Long-Acting Beta-Agonists (LABAs) 2010. <http://www.fda.gov/Drugs/DrugSafety/PostmarketDrugSafetyInformationforPatientsandProviders/ucm200776.htm>. Accessed October 16, 2010. IV.
428. Meltzer SS, Hasday JD, Cohn J, Bleecker ER. Inhibition of exercise-induced bronchospasm by zileuton: a 5-lipoxygenase inhibitor. *Am J Respir Crit Care Med*. 1996;153:931–5. Ib.
429. Finnerty JP, Holgate ST. Evidence for the roles of histamine and prostaglandins as mediators in exercise-induced asthma: the inhibitory effect of terfenadine and flurbiprofen alone and in combination. *Eur Respir J*. 1990;3:540–7. Ib.
430. O'Byrne PM. Leukotriene bronchoconstriction induced by allergen and exercise. *Am J Respir Crit Care Med*. 2000;161(2 pt 2):S68–72. IV.
431. Fogel RB, Rosario N, Aristizabal G, et al. Effect of montelukast or salmeterol added to inhaled fluticasone on exercise-induced broncho-

- constriction in children. *Ann Allergy Asthma Immunol.* 2010;104:511–7. Ib.
432. O'Byrne PM. Initiation, dose reduction, and duration of inhaled corticosteroid therapy. *Immunol Allergy Clin North Am.* 2005;25:511–21. IV.
433. Smith LJ, Greenberger PA, Patterson R, Krell RD, Bernstein PR. The effect of inhaled leukotriene D4 in humans. *Am Rev Respir Dis.* 1985;131:368–72. Iia.
434. Finnerty JP, Wood-Baker R, Thomson H, Holgate ST. Role of leukotrienes in exercise-induced asthma. Inhibitory effect of ICI 204219, a potent leukotriene D4 receptor antagonist. *Am Rev Respir Dis.* 1992;145:746–9. Ib.
435. Mastalerz L, Gawlewicz-Mroccka A, Nizankowska E, Cmiel A, Szczeklik A. Protection against exercise-induced bronchoconstriction by montelukast in aspirin-sensitive and aspirin-tolerant patients with asthma. *Clin Exp Allergy.* 2002;32:1360–5. Ib.
436. Coreno A, Skowronski M, Kotaur C, McFadden ER Jr. Comparative effects of long-acting β 2-agonists, leukotriene receptor antagonists, and a 5-lipoxygenase inhibitor on exercise-induced asthma. *J Allergy Clin Immunol.* 2000;106:500–6. Ib.
437. Philip G, Villarán C, Pearlman DS, Loeys T, Dass SB, Reiss TF. Protection against exercise-induced bronchoconstriction two hours after a single oral dose of montelukast. *J Asthma.* 2007;44:213–7. Ib.
438. Philip G, Pearlman DS, Villarán C, et al. Single-dose montelukast or salmeterol as protection against exercise-induced bronchoconstriction. *Chest.* 2007;132:875–83. Ib.
439. Bronsky EA, Kemp JP, Zhang J, Guerreiro D, Reiss TF. Dose-related protection of exercise bronchoconstriction by montelukast, a cysteinyl leukotriene-receptor antagonist, at the end of a once-daily dosing interval. *Clin Pharmacol Ther.* 1997;62:556–61. Ib.
440. Peroni DG, Piacentini GL, Ress M, et al. Time efficacy of a single dose of montelukast on exercise-induced asthma in children. *Pediatr Allergy Immunol.* 2002;13:434–7. Ib.
441. de Benedictis FM, del Giudice MM, Foreza N, Decimo F, de Benedictis D, Capristo A. Lack of tolerance to the protective effect of montelukast in exercise-induced bronchoconstriction in children. *Eur Respir J.* 2006;28:291–5. Ib.
442. Drazen JM, Silverman EK, Lee TH. Heterogeneity of therapeutic responses in asthma. *Br Med Bull.* 2000;56:1054–70. Ib.
443. Kim JH, Lee SY, Kim HB, et al. TBXA2R gene polymorphism and responsiveness to leukotriene receptor antagonist in children with asthma. *Clin Exp Allergy.* 2008;38:51–9. Iia.
444. Lehnigk B, Rabe KF, Dent G, Herst RS, Carpentier PJ, Magnussen H. Effects of a 5-lipoxygenase inhibitor, ABT-761, on exercise-induced bronchoconstriction and urinary LTE4 in asthmatic patients. *Eur Respir J.* 1998;11:617–23. Ib.
445. Van Schoor J, Joos GF, Kips JC, Drajesk JF, Carpentier PJ, Pauwels RA. The effect of ABT-761, a novel 5-lipoxygenase inhibitor, on exercise- and adenosine-induced bronchoconstriction in asthmatic subjects. *Am J Respir Crit Care Med.* 1997;155:875–80. Ib.
446. Manning PJ, Watson RM, Margolskee DJ, Williams VC, Schwartz JI, O'Byrne PM. Inhibition of exercise-induced bronchoconstriction by MK-571, a potent leukotriene D4-receptor antagonist. *N Engl J Med.* 1990;323:1736–9. Ib.
447. Silverman M, Andrea T. Time course of effect of disodium cromoglycate on exercise-induced asthma. *Arch Dis Child.* 1972;47:419–22. Ib.
448. Woolley M, Anderson SD, Quigley BM. Duration of protective effect of terbutaline sulphate and cromolyn sodium alone and in combination on exercise-induced asthma. *Chest.* 1990;97:39–45. Ib.
449. Comis A, Valletta EA, Sette L, Andreoli A, Boner AL. Comparison of nedocromil sodium and sodium cromoglycate administered by pressurized aerosol, with and without a spacer device in exercise-induced asthma in children. *Eur Respir J.* 1993;6:523–6. Ib.
450. de Benedictis FM, Tuteri G, Pazzelli P, Solinas LF, Niccoli A, Parente C. Combination drug therapy for the prevention of exercise-induced bronchoconstriction in children. *Ann Allergy Asthma Immunol.* 1998;80:352–6. Ib.
451. Patel KR, Wall RT. Dose-duration effect of sodium cromoglycate aerosol in exercise-induced asthma. *Eur J Respir Dis.* 1986;69:256–60. Ib.
452. Patel KR, Tullett WM, Neale MG, Wall RT, Tan KM. Plasma concentrations of sodium cromoglycate given by nebulisation and metered dose inhalers in patients with exercise-induced asthma: relationship to protective effect. *Br J Clin Pharmacol.* 1986;21:231–3. Ib.
453. Kuzemko JA. Twenty years of sodium cromoglycate treatment: a short review. *Respir Med.* 1989;83:11–4. IV.
454. Henriksen JM, Wenzel A. Effect of an intranasally administered corticosteroid (budesonide) on nasal obstruction, mouth-breathing, and asthma. *Am Rev Respir Dis.* 1984;130:1014–8. Ib.
455. Henriksen JM. Effect of inhalation of corticosteroids on exercise induced asthma: randomised double blind crossover study of budesonide in asthmatic children. *Br Med J (Clin Res Ed).* 1985;291:248–9. Ib.
456. Yates DH, Kharitonov S, Barnes PJ. An inhaled glucocorticoid does not prevent tolerance to the protective effect of a long-acting inhaled beta 2-agonist. *Am J Respir Crit Care Med.* 1996;154:1603–7. Ib.
457. Blake K. Review of guidelines and the literature in the treatment of acute bronchospasm in asthma. *Pharmacotherapy.* 2006;26(9 pt 2):148S–55S. IV.
458. Beil M, de Kock MA. Role of alpha-adrenergic receptors in exercise-induced bronchoconstriction. *Respiration.* 1978;35:78–86. Iia.
459. Boulet L-P, Turcotte H, Tennina S. Comparative efficacy of salbutamol, ipratropium and cromoglycate in the prevention of bronchospasm induced by exercise and hyperosmolar challenges. *J Allergy Clin Immunol.* 1989;83:882–7. Ib.
460. Poppius H, Sovijärvi ARA, Tammilehto L. Lack of protective effect of high-dose ipratropium on bronchoconstriction following exercise with cold air breathing in patients with mild asthma. *Eur J Respir Dis.* 1986;68:319–25. Ib.
461. Magnussen H, Nowak D, Wiebicke W. Effect of inhaled ipratropium bromide on the airway response to methacholine, histamine, and exercise in patients with mild bronchial asthma. *Respiration.* 1992;59:42–7. Ib.
462. Boner AL, Vallone G, De Stefano G. Effect of inhaled ipratropium bromide on methacholine and exercise provocation in asthmatic children. *Pediatr Pulmonol.* 1989;6:81–5. Ib.
463. Ellis EF. Inhibition of exercise-induced asthma by theophylline. *J Allergy Clin Immunol.* 1984;73(5 pt 2):690–2. IV.
464. Iikura Y, Hashimoto K, Akasawa A, et al. Serum theophylline concentration levels and preventative effects on exercise-induced asthma. *Clin Exp Allergy.* 1996;26(suppl 2):38–41. Iia.
465. Seale JP, Anderson SD, Lindsay DA. A comparison of oral theophylline and oral salbutamol in exercise-induced asthma. *Aust N Z J Med.* 1977;7:270–4. Iia.
466. Timmer W, Leclerc V, Birraux G, et al. The phosphodiesterase 4 inhibitor roflumilast is efficacious in exercise-induced asthma and leads to suppression of LPS-stimulated TNF- α ex vivo. *J Clin Pharmacol.* 2002;42:297–303. Ib.
467. Clee MD, Ingram CG, Reid PC, Robertson AS. The effect of astemizole on exercise-induced asthma. *Br J Dis Chest.* 1984;78:180–3. Iia.
468. Magnussen H, Reuss G, Jörres R, Aurich R. The effect of azelastine on exercise-induced asthma. *Chest.* 1988;93:937–40. Ib.
469. Wiebicke W, Poynter A, Montgomery M, Chernick V, Pasterkamp H. Effect of terfenadine on the response to exercise and cold air in asthma. *Pediatr Pulmonol.* 1988;4:225–9. Ib.
470. Zielfinski J, Chodosowska E. Exercise-induced bronchoconstriction in patients with bronchial asthma. Its prevention with an antihistaminic agent. *Respiration.* 1977;34:31–5. Iia.
471. Peroni DG, Piacentini GL, Pietrobello A, et al. The combination of single-dose montelukast and loratadine on exercise-induced bronchospasm in children. *Eur Respir J.* 2002;20:104–7. Ib.
472. Manjra AI, Nel H, Maharaj B. Effect of desloratadine on patients with allergic rhinitis and exercise-induced bronchoconstriction: a placebo controlled study. *J Asthma.* 2009;46:156–9. Ib.

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473. Passalacqua G, Canonica GW, Bousquet J. Structure and classification of H1-antihistamines and overview of their activities. *Clin Allergy Immunol.* 2002;17:65–100. IV.
474. Ghosh SK, De Vos C, McIlroy I, Patel KR. Effect of cetirizine on exercise induced asthma. *Thorax.* 1991;46:242–4. Ib.
475. Barnes PJ, Wilson NM, Brown MJ. A calcium antagonist, nifedipine, modifies exercise-induced asthma. *Thorax.* 1981;36:726–30. Ib.
476. Corris PA, Nariman S, Gibson GJ. Nifedipine in the prevention of asthma induced by exercise and histamine. *Am Rev Respir Dis.* 1983; 128:991–2. Ib.
477. Lockhart A, Slutsky AS. Furosemide and loop diuretics in human asthma. *Chest.* 1994;106:244–9. IV.
478. Prandota J. Furosemide: progress in understanding its diuretic, anti-inflammatory, and bronchodilating mechanism of action, and use in the treatment of respiratory tract diseases. *Am J Ther.* 2002;9:317–28. IV.
479. Bianco S, Vaghi A, Robuschi M, Pasargiklian M. Prevention of exercise-induced bronchoconstriction by inhaled frusemide. *Lancet.* 1988; 2:252–5. Ib.
480. Barnes PJ, Wilson NM, Vickers H. Prazosin, an alpha 1-adrenoceptor antagonist, partially inhibits exercise-induced asthma. *J Allergy Clin Immunol.* 1981;68:411–5. Ib.
481. Ahmed T, Gonzalez BJ, Danta I. Prevention of exercise-induced bronchoconstriction by inhaled low-molecular-weight heparin. *Am J Respir Crit Care Med.* 1999;160:576–81. Ib.
482. Kunz LI, van Rensen EL, Sterk PJ. Inhaled hyaluronic acid against exercise-induced bronchoconstriction in asthma. *Pulm Pharmacol Ther.* 2006;19:286–91. Ib.
483. McKenzie DC, McLuckie SL, Stirling DR. The protective effects of continuous and interval exercise in athletes with exercise-induced asthma. *Med Sci Sports Exerc.* 1994;26:951–6. Ib.
484. Mickleborough TD, Lindley MR, Turner LA. Comparative effects of a high-intensity interval warm-up and salbutamol on the bronchoconstrictor response to exercise in asthmatic athletes. *Int J Sports Med.* 2007;28:456–62. Ib.
485. Mickleborough TD, Fogarty A. Dietary sodium intake and asthma: an epidemiological and clinical review. *Int J Clin Pract.* 2006;60: 1616–24. Ib.
486. Mickleborough TD, Lindley MR. Diet and exercise-induced bronchoconstriction. *Chest.* 2006;130:623–4. Ib.
487. Tecklenburg SL, Mickleborough TD, Fly AD, Bai Y, Stager JM. Ascorbic acid supplementation attenuates exercise-induced bronchoconstriction in patients with asthma. *Respir Med.* 2007;101:1770–8. Ib.
488. Anderson SD, Pearlman DS, Rundell KW, Perry CP, Boushey H, Sorkness CA, Nichols S, Weiler JM. Reproducibility of the airway response to an exercise protocol standardized for intensity, duration, and inspired air conditions, in subjects with symptoms suggestive of asthma. *Respir Res.* 2010 Sep 1;11:120. Iib.
489. Clearie KL, Williamson PA, Vaidyanathan S, et al. Disconnect between standardized field-based testing and mannitol challenge in Scottish elite swimmers. *Clin Exp Allergy.* 2010. May;40:731–7. Iib.
490. O'Byrne PM, Gauvreau GM, Brannan JD. Provoked models of asthma: what have we learnt? *Clin Exp Allergy.* 2009;39:181–92. IV.
491. National Asthma Education and Prevention Program. *Expert Panel Report 3. Guidelines for the Diagnosis and Management of Asthma.* Bethesda, MD: National Heart, Lung, and Blood Institute, US Dept of Health and Human Services, National Institutes of Health; 2007. NIH publication 07-4051. IV.
492. Bougault V, Turmel J, Boulet LP. Bronchial challenges and respiratory symptoms in elite swimmers and winter sport athletes. *Chest.* 2010; 138(suppl 2):31S–37S. Iib.
493. Hull JH, Hull PJ, Parsons JP, Dickinson JW, Ansley L. Approach to the diagnosis and management of suspected exercise-induced bronchoconstriction by primary care physicians. *BMC Pulm Med.* 2009;9:29. IV.

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